LARVA: An integrative framework for Large-scale Analysis of Recurrent Variants in noncoding Annotations



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Finding Key Variants in Cancer Genomes: the Needle in the Haystack



[Image credit: www.yourgrantauthority.com '15]

- Increasing number of whole genome sequenced for tumor/normal pairs
 - Eg >2500 for PCAWG
- Lots of somatic mutations in an average tumor (~5K/sample), particularly in non-coding regions
- A focus is distinguishing drivers & passengers
 - Canonical Drivers are mutations driving cancer progression
 - Thought to be under positive selection
 - · Recur in the same position, gene or functional element across tumors in different individuals
 - Passengers are thought not be significant to driving cancer progression
 - Collateral damage
 - Could result from impaired DNA repair processes
- Most driver work has focused on genes
 - eg Youn & Simon ('11). Bioinformatics; Lawrence et al. ('13). Nature

Noncoding Annotations





Ultra-sensitive & Ultraconserved elements Noncoding regions more conserved than expectation across the human population & between species [Bejerano et al. ('04). Science; Khurana et al., Science ('13)]

Identification of non-coding candidate drivers amongst somatic variants: FunSeq



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Candidate driver

From Funseq 1.0 to Funseq 2.0

- Elaborated features
 - $\circ~$ Motif disruption score: changes in PWMs
 - Network centrality analysis: PPI, regulatory, and phosphorylation networks



Mutation recurrence



Mutation recurrence







Cancer Somatic Mutational Heterogeneity, across cancer types, samples & regions





[Lochovsky et al. NAR ('15)]

1 Mbp genome regions (locations chosen at random)

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Cancer Somatic Mutation Modeling

- 3 models to evaluate the significance of mutation burden
- Suppose there are *k* genome elements. For element *i*, define:
 - *n_i*: total number of nucleotides
 - x_i: the number of mutations within the element
 - p: the mutation rate
 - R: the replication timing bin of the element

Model 1: Constant Background Mutation Rate (Model from Previous Work)

 $\mathbf{x_i}: Binomial(\mathbf{n_i}, \mathbf{p})$

Model 2: Varying Mutation Rate

 $\mathbf{x_i} | \mathbf{p_i} : Binomial(\mathbf{n_i}, \mathbf{p_i})$

 $\mathbf{p_i}: Beta(\mu, \sigma)$

Model 3: Varying Mutation Rate with Replication Timing Correction

 $\mathbf{x_i} | \mathbf{p_i} : Binomial(\mathbf{n_i}, \mathbf{p_i})$

 $\mathbf{p_i}: Beta(\mu|\mathbf{R}, \sigma|\mathbf{R})$

 $\mu|\mathbf{R},\sigma|\mathbf{R}:$ constant within the same \mathbf{R} bin

[[]Lochovsky et al. NAR ('15)]

LARVA Model Comparison

- Comparison of mutation count frequency implied by the binomial model (model 1) and the beta-binomial model (model 2) relative to the empirical distribution
- The beta-binomial distribution is significantly better, especially for accurately modeling the over-dispersion of the empirical distribution



Adding DNA replication timing correction further improves the beta-binomial model



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LARVA Results



[Lochovsky et al. NAR ('15)]

LARVA Implementation

- http://larva.gersteinlab.org/
- Freely downloadable C++ program
 - Verified compilation and correct execution on Linux
- A Docker image is also available to download
 - Runs on any operating system supported by Docker
- Running time on transcription factor binding sites (a worst case input size) is ~80 min
 - Running time scales linearly with the number of annotations in the input



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