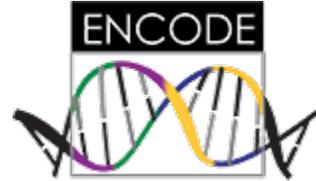


Using gkm-SVM to Detect Conserved Enhancers Where Alignment Fails

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Sept 2, 2017



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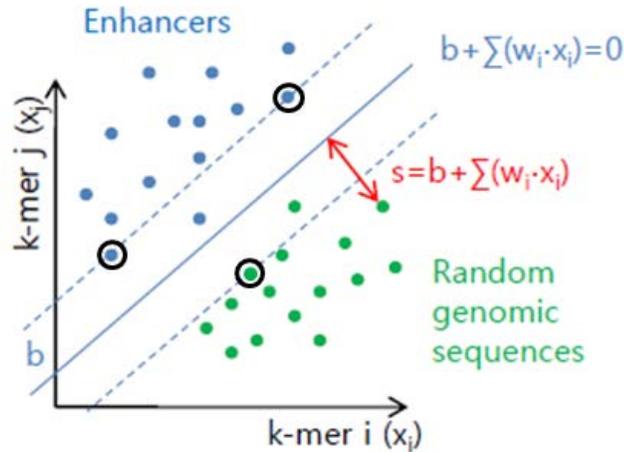
Felix Yu
Gianluca Silva Croso
Justin Shigaki
Dongwon Lee
M. Ghandi

validation experiments:
Dave Gorkin
Sarah McClymont
Andy McCallion

- ~25,000 non-coding genetic variants (SNPs) associated with common diseases, most in cell specific regulatory regions as detected by chromatin accessibility
- gapped kmer SVM (gkm-SVM) classifier trained on open chromatin can predict impact of GWAS SNPs
- relevant tissue/cell-type and orthologous mouse regulatory regions for functional evaluation often unknown
- gkm-SVM kernel can be used to detect conserved orthologous regions where alignment fails

gkm-SVM and deltaSVM Method Overview

- Define a set of cell type specific enhancers using functional genomics data: Dnase-seq ATAC-seq
- Train gkm-SVM to learn regulatory TF binding site vocabulary for given cell-type
- Calculate how each SNP changes gkm-SVM score to predict impact (deltaSVM)



2017 Beer, Human Mutation
 2016 gkm-SVM R package, Bioinformatics
 2015 Lee - Beer Nature Genetics (deltaSVM)
 2014 Lee, Ghandi, Beer, PLOS Comp Bio (gapped kmers)
 2013 Fletez-Brant, Lee, Beer, NAR
 2013 Ghandi, Beer, J Math Biol (gapped kmers)
 2011 Lee, Karchin, Beer, Genome Research (kmers)

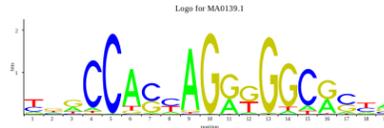
2004 Noble, Leslie

k-mer counts:

CACCAGGGGG
 CCACCAGGGG
 CCACCTGGTG
 CCACCAGGTG

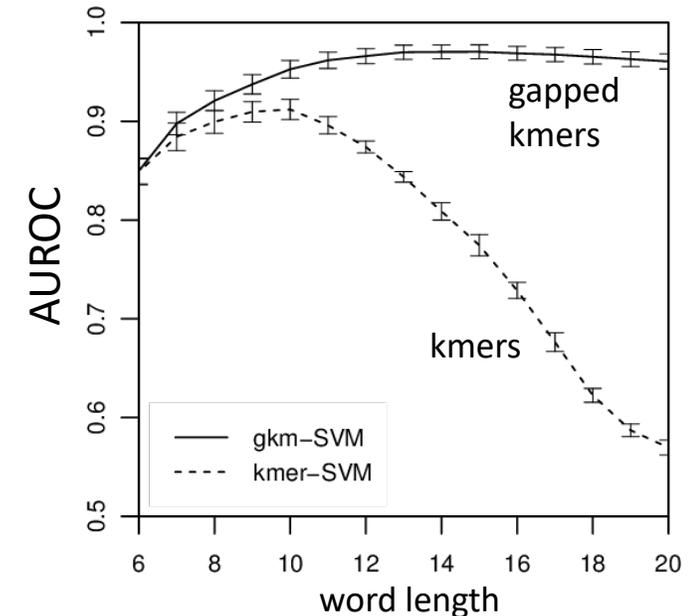
gapped kmer counts:

CA--AG--GG
 CC-C--G-GG
 CA-C-G--GG
 CCA---GG-G



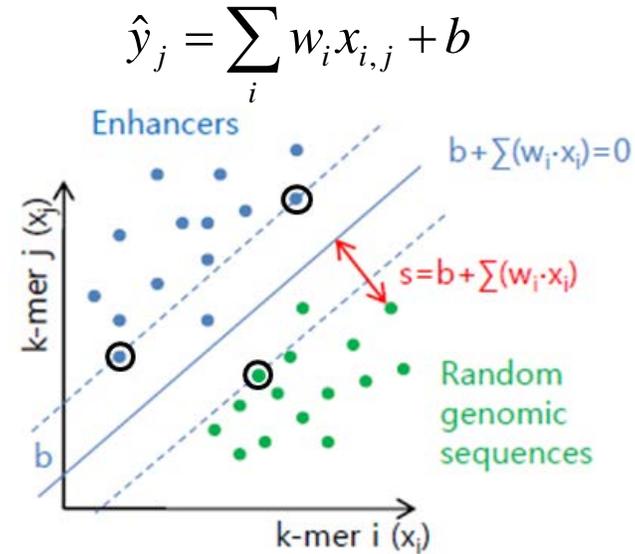
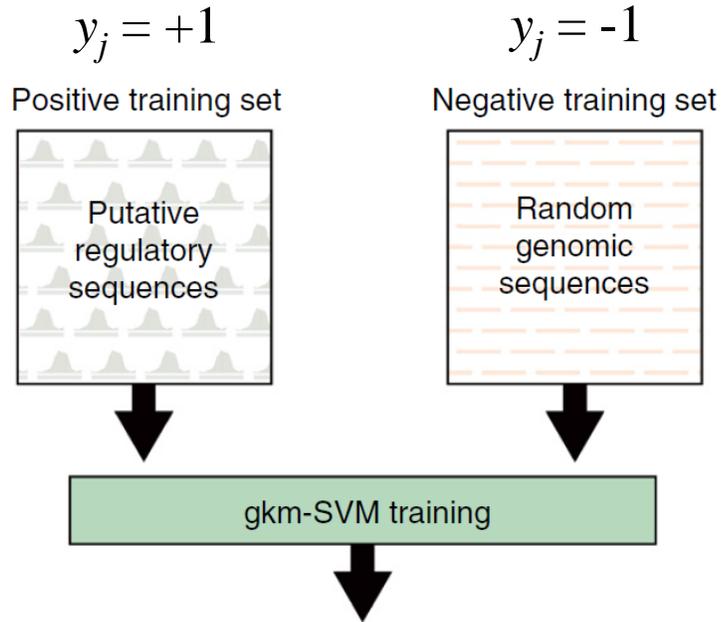
CTCF len=15

$$K(S_1, S_2) = \frac{\langle f^{S_1}, f^{S_2} \rangle}{\|f^{S_1}\| \|f^{S_2}\|}$$



Generate GC matched negative set, Train SVM to Identify Regulatory Features

Lee, Karchin, Beer Genome Research Dec 2011



Regulatory sequence vocabulary

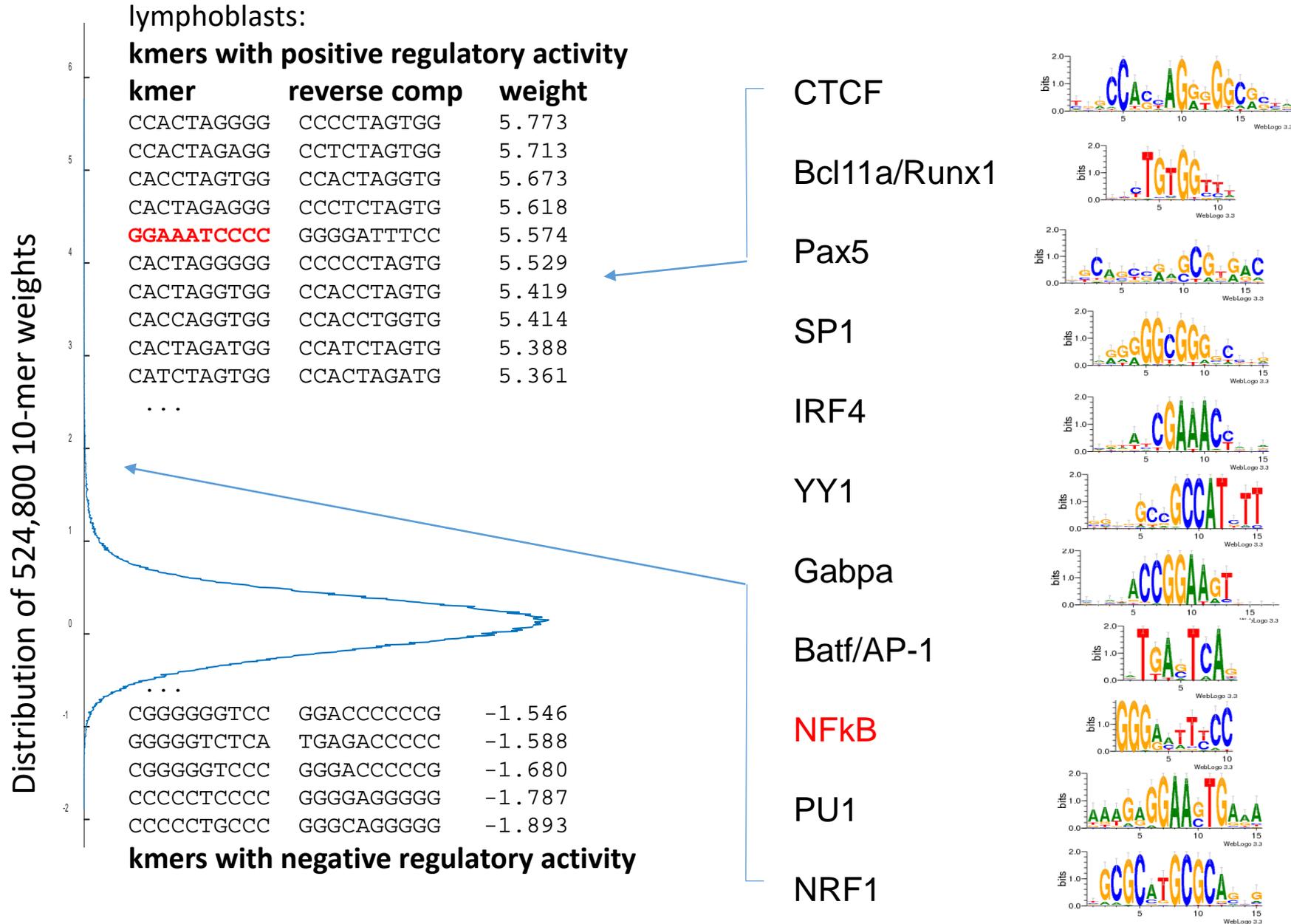
All unique 10-mers	SVM weights	
ATGACTCATC	3.275	Positive regulatory activity
ATGAGTCATC	3.147	
ATCATGTGAC	3.091	
GTCACATGAC	2.992	
⋮	⋮	
ACGAGAAACA	0.0002	Neutral
ACTATAACCA	0.0001	
ATTGCTAAGC	-0.0001	
GGATAAAATA	-0.0001	
⋮	⋮	
CAGGTGTGAG	-1.171	Negative regulatory activity
ATCACACCTG	-1.183	
ACACACCTGT	-1.253	
AATCCAGGTG	-1.282	

deltaSVM calculation

WT 10-mer	Weight	Variant 10-mer	Weight	Difference
TCTCTGCAAC	0.012	TCTCTGCAAA	0.298	0.286
CTCTGCAACA	0.082	CTCTGCAAAA	0.104	0.021
TCTGCAACAA	0.280	TCTGCAAAAA	0.114	-0.166
CTGCAACAAA	0.330	CTGCAAAAAA	-0.029	-0.359
TGCAACAAAG	0.441	TGCAAAAAAG	-0.025	-0.466
GCAACAAAGA	0.784	GCAAAAAAGA	0.008	-0.776
CAACAAAGAC	1.031	CAAAAAAGAC	0.109	-0.922
AACAAAGACA	0.545	AAAAAAGACA	-0.453	-0.998
ACAAAGACAG	0.671	AAAAAGACAG	-0.516	-1.187
CAAAGACAGA	-0.036	AAAAGACAGA	-0.478	-0.442

▲ deltaSVM = -5.007

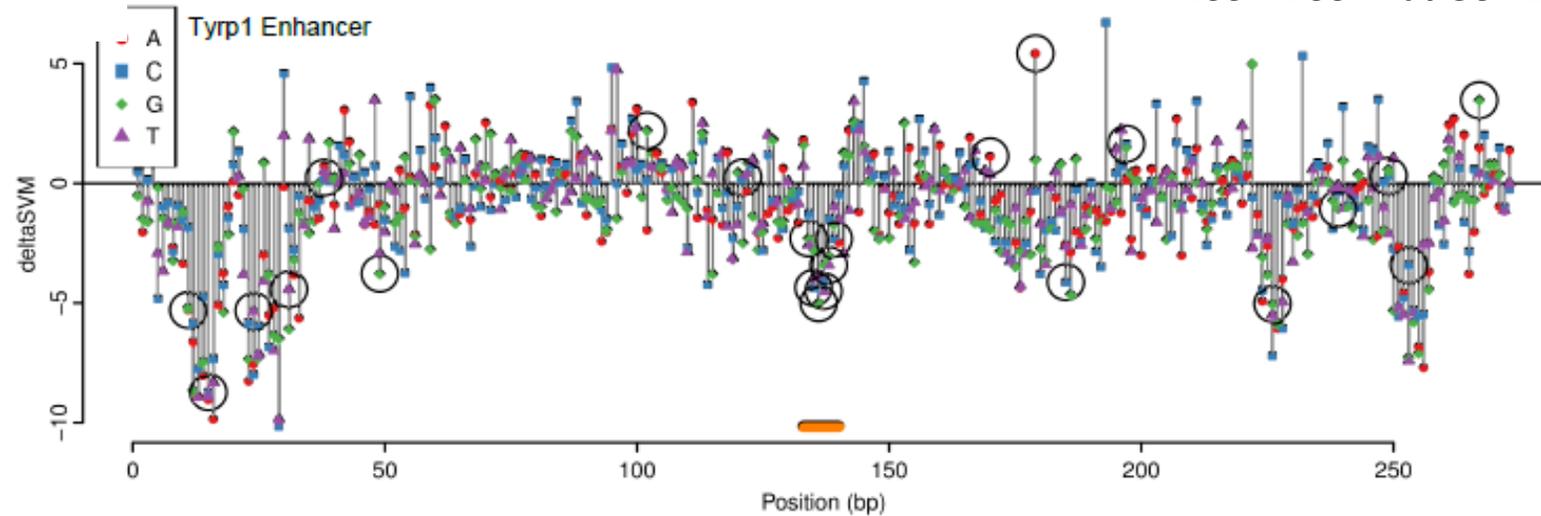
gkm-SVM Weights Encode Co-factor TFs Required to Predict Cell-specific Binding



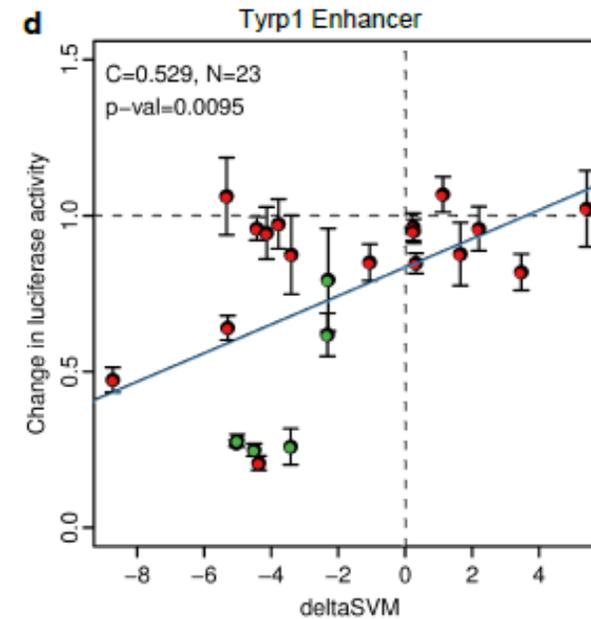
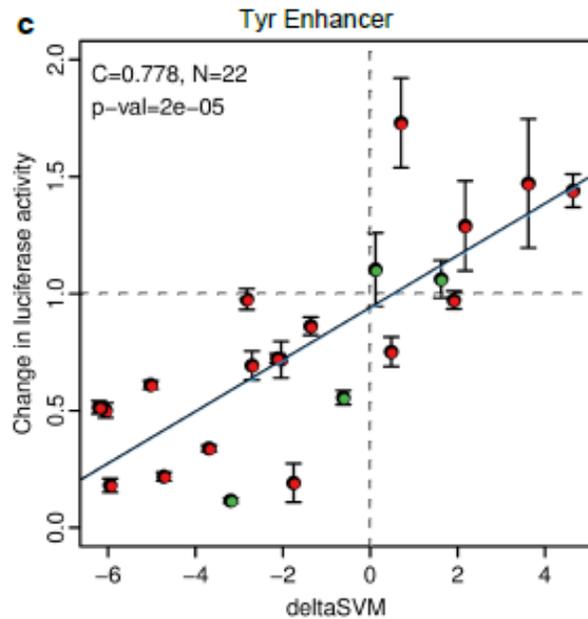
gkmSVM Predicts Effect of mutations in melanocyte enhancers

Direct Validation

Lee—Beer Nat Gen 2015

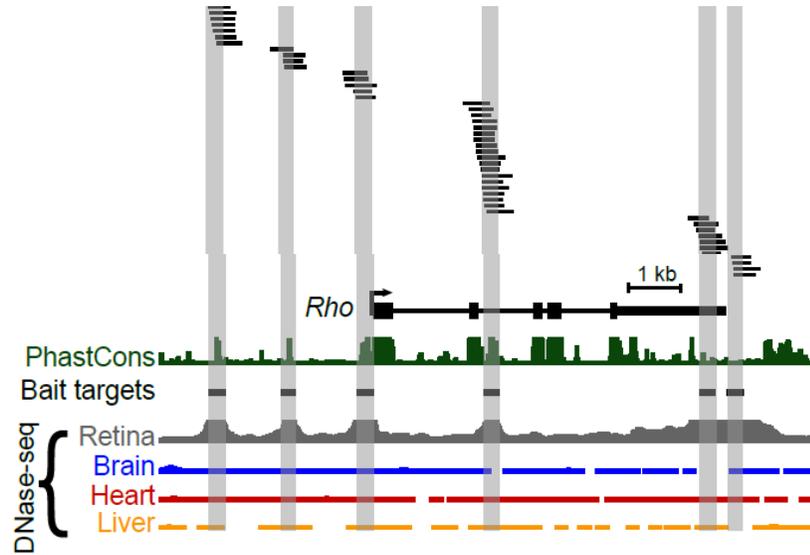


McCallion Lab



Similar results comparing to
MPRAs:
Patwardhan et al., 2012
Kheradpour et al, 2013

gkmSVM Predicts MPRA Expression *in vivo* in Mouse Retina

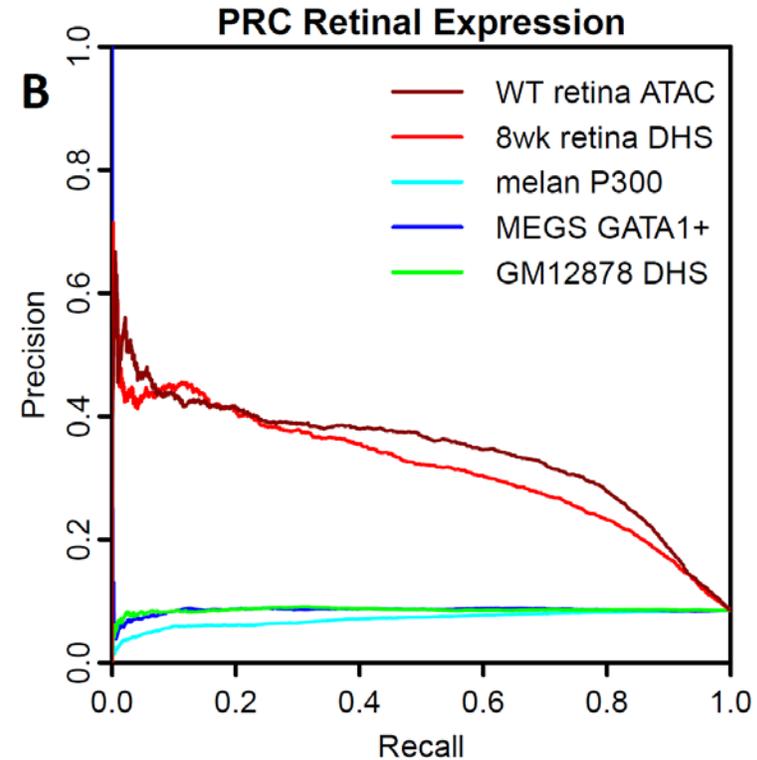
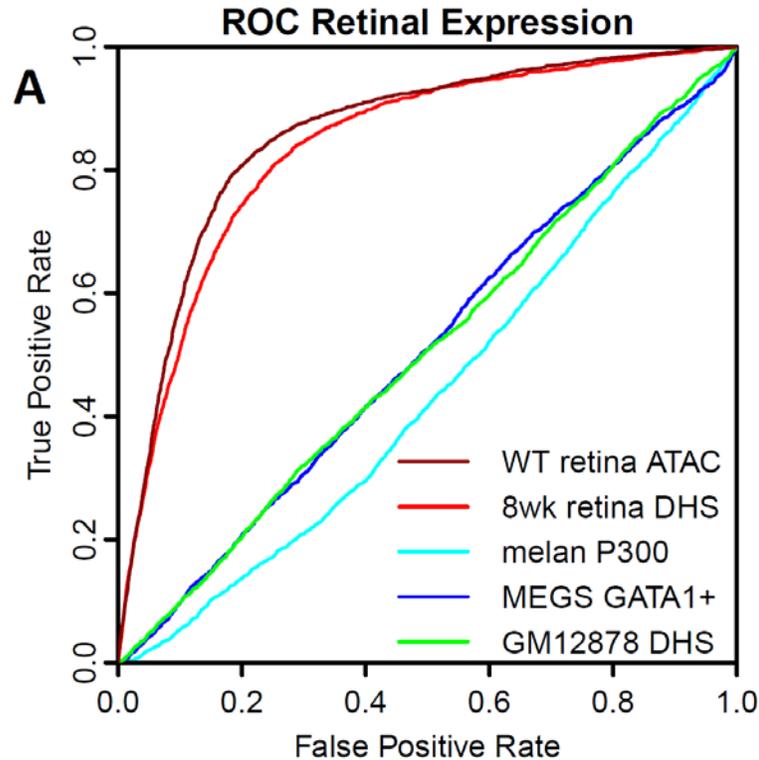


36005 DHS+ elements tested in mouse retina
(Shen—Corbo GR 2015)

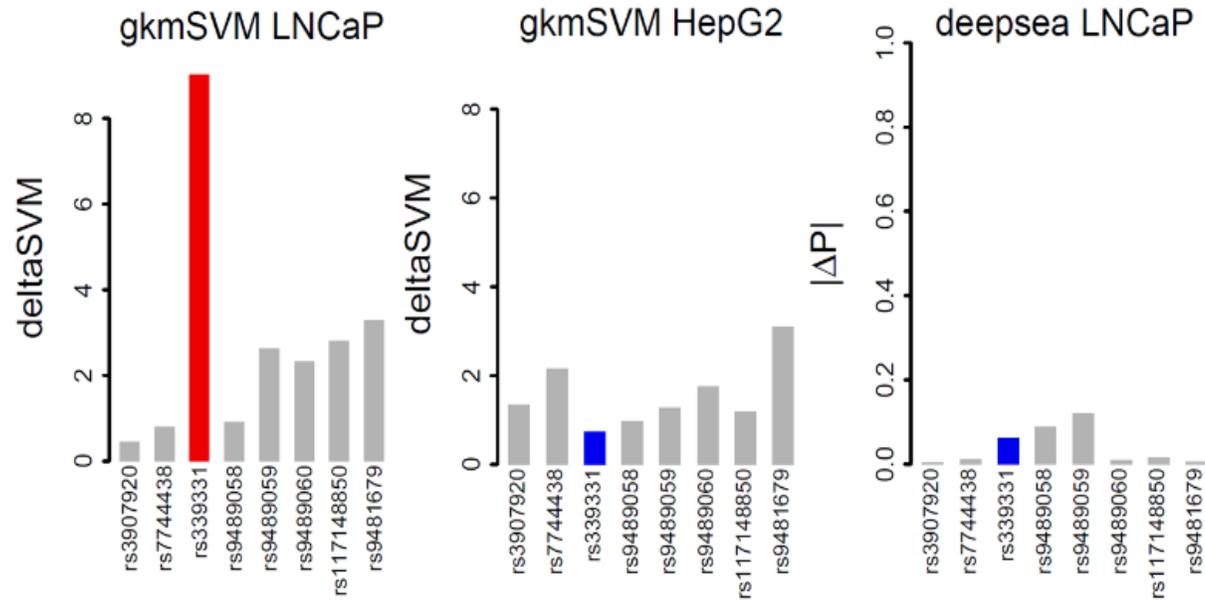
Pos: Strong expression 2156 seqs
Neg: Low expression 23147 seqs

gkmSVM trained on retina DHS or ATAC
predicts expression, but not other cell types

Beer Human Mut 2017



Validated causal disease SNPs only score higher than flanking negative SNPs when gkmSVM trained on relevant cell type



some cell types (immune) are over-represented in ENCODE data set

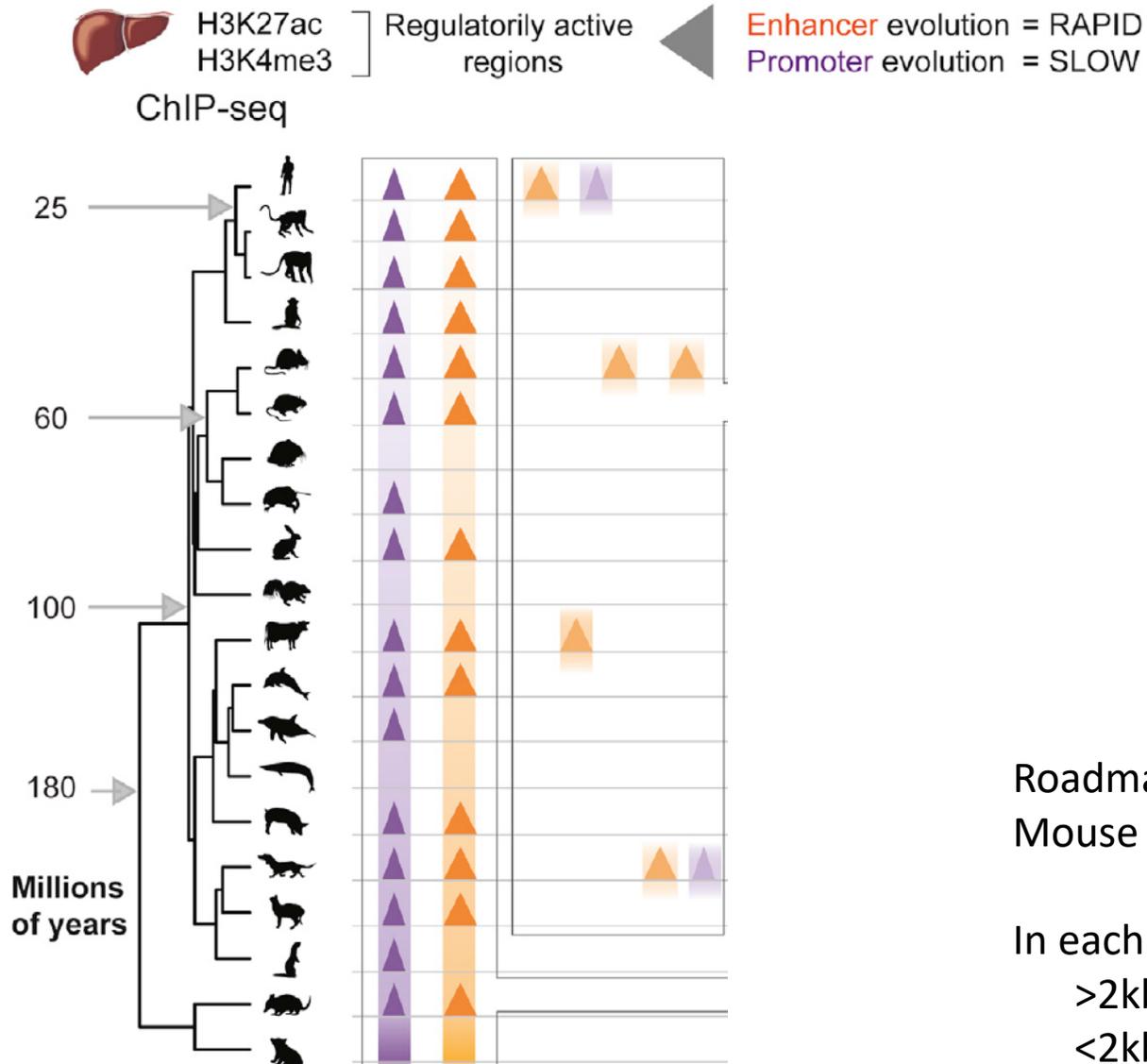
Deepsea misses *Rfx6* prostate cancer SNP, even though test set AUROC is high for LNCaP class

Beer Human Mutation 2017

gkm-SVM identifies *RFX6* prostate cancer SNP rs339331 (Huang NG 2014) (red) from among flanking SNPs when trained on:

- ENCODE LNCaP DHS (prostate, red)
- but not HepG2 DHS (liver, blue)
- Deepsea predicts *RFX6* SNP has low ΔP for LNCaP DHS.
- Imbalance in training data can skew predictive accuracy across classes
- models with similar test-set CV AUROC can give very different predictions for variant impact

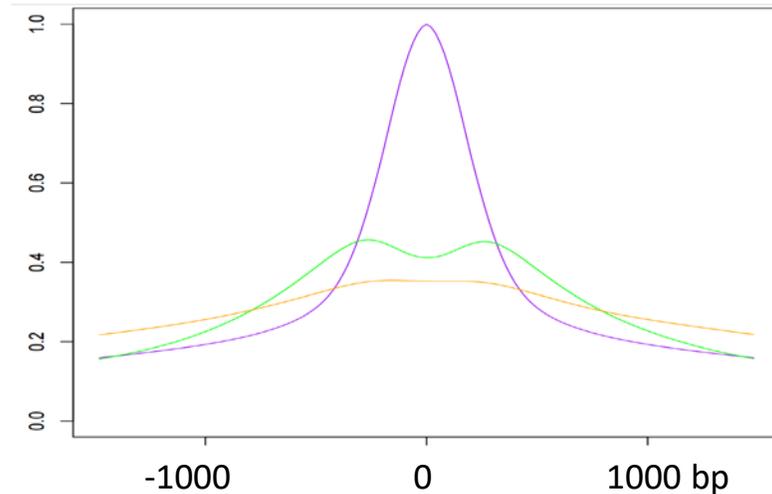
Enhancer and promoter evolution in twenty mammals



TFBS are conserved,
liver H3K27Ac activity is conserved
but enhancers are not alignable, why?

Genome-wide Cross-correlation of
DHS with chromatin marks

$$(f * g)(\Delta) = \sum_x f(x)g(x + \Delta)$$



DHS with:
H3K4me1
H3K27ac
DHS

Roadmap DHS (73 Human Tissues)

Mouse ENCODE DHS (53 Mouse Tissues)

In each tissue, select 10000 strongest cell-specific peaks

>2kb from TSS = enhancers

<2kb from TSS = promoters



train vs GC matched neg set

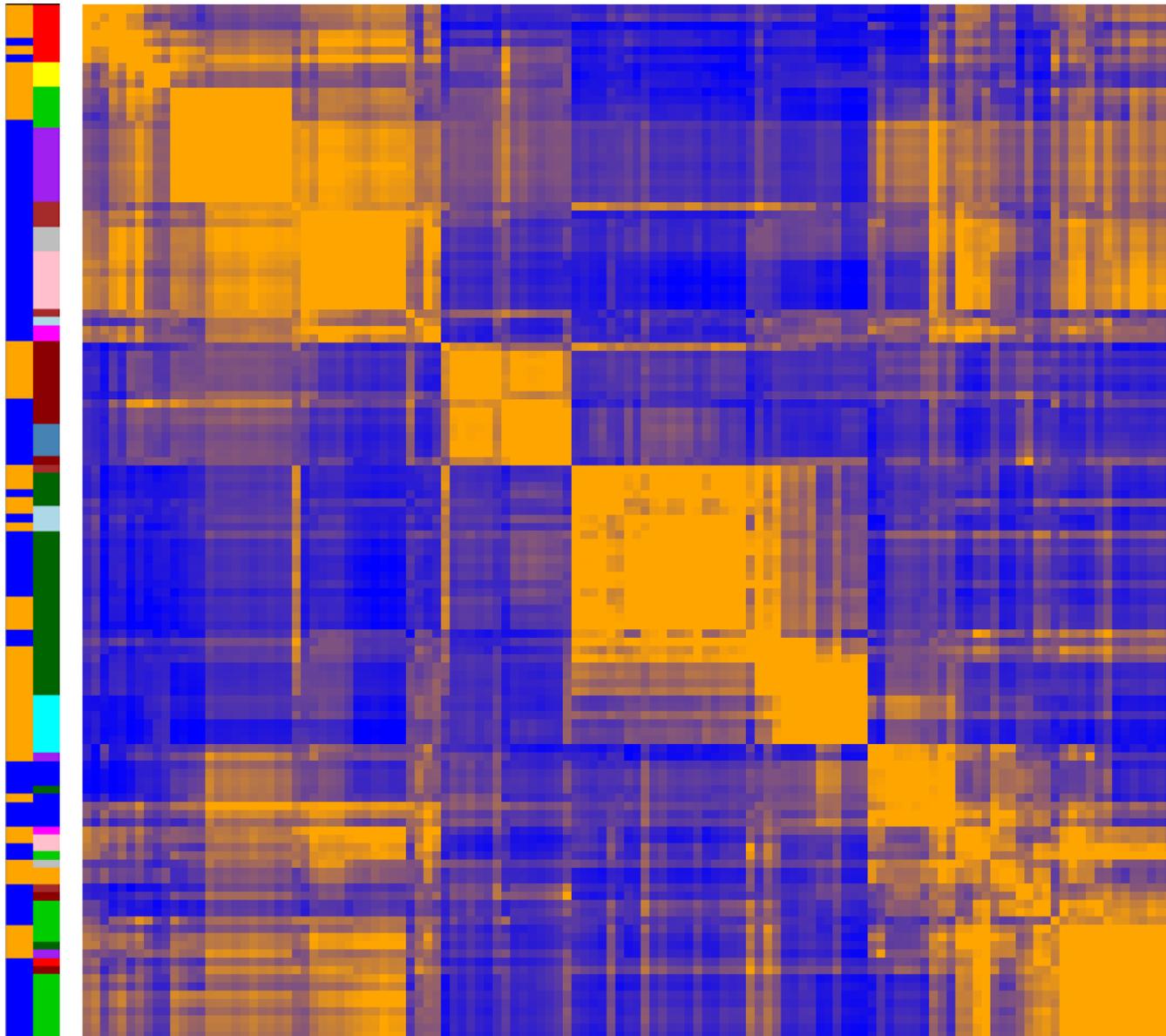
gkm-SVM Identifies Similar Sequence Features in Matched Human and Mouse Tissues

Train gkm-SVM on 10000 mouse ENCODE (53) and human Roadmap DHS (73, Stam)

Cluster by correlation across all gapped kmer weights $C(w_i, w_j)$:

median AUC > 0.9

■ human low corr
■ mouse high corr



Mm.WholebrainC57Bl6MAdult8wks
 Mm.CerebrumC57Bl6MAdult8wks
 Mm.CerebellumC57Bl6MAdult8wks
 Mm.WbBrainC57Bl6ME18half
 Hs.FetalBrain
 Mm.WholebrainC57Bl6ME14half
 Hs.FetalSpinalCord
 Mm.RetinaC57Bl6MNew1days
 Mm.RetinaC57Bl6MAdult8wks
 Mm.RetinaC57Bl6MAdult8wks
 Mm.MesodermC01ME11half
 Mm.HemibodyC01ME11half
 Mm.FibrotC01ME11half
 Mm.HibodyC01ME11half
 Hs.FetalSkin
 Hs.FetalRenalCortex
 Hs.FetalRenalPelvis
 Hs.FetalKidney_Left
 Hs.FetalKidney_Right
 Hs.FetalKidney
 Hs.FetalRenalPelvisRight
 Hs.FetalRenalPelvisLeft
 Hs.FetalRenalCortexLeft
 Hs.FetalRenalCortexRight
 Hs.FetalSpleen
 Hs.FetalTestes
 Hs.Ovary
 Hs.FetalLungRight
 Hs.FetalLungLeft
 Hs.FetalLung
 Hs.FetalMuscleUpperTrunk
 Hs.FetalMuscleLowerLimbSkeletal
 Hs.FetalMuscleUpperLimbSkeletal
 Hs.FetalMuscleTrunk
 Hs.FetalMuscleBack
 Hs.FetalMuscleLeg
 Hs.FetalMuscleArm
 Hs.FetalOvary
 Hs.FetalRenalGland
 Hs.Heart
 Hs.FetalHeart
 Mm.Zbtb41290aMEODfb6h
 Mm.Zbtb41290aME0
 Mm.Esiv75129ME0
 Mm.Esivw9UknME0
 Mm.Esiv141290aME0
 Mm.Esivw9UknME0
 Mm.Zbtb41290aMEODfb24h
 Hs.H1DerivedNeuronalProgenitor
 Hs.H1
 Hs.H9
 Hs.PDF9g
 Hs.PDF47
 Hs.PDF1811
 Hs.PDF107
 Hs.H1BM4DerivedMesoderm
 Mm.SpleenC57Bl6MAdult8wks
 Mm.Boeloid19cC57Bl6MAdult8wks
 Mm.Boeloid30cC57Bl6MAdult8wks
 Hs.CD10PrimaryCells
 Mm.C1224aFImmortal
 Mm.A20Balb/cImmortal8wks
 Hs.FetalThymus
 Mm.ThymusC57Bl6MAdult8wks
 Hs.MobilizedCD8PrimaryCells
 Hs.MobilizedCD4PrimaryCells
 Hs.CD4PrimaryCells
 Hs.MobilizedCD8PrimaryCells
 Hs.CD8PrimaryCells
 Hs.CD8PrimaryCells
 Hs.CD8PrimaryCells
 Hs.MobilizedCD8PrimaryCells
 Mm.TrnaC57Bl6MAdult8wks
 Mm.ThepacC57Bl6MAdult8wks
 Mm.TregC57Bl6MAdult8wks
 Mm.TregC57Bl6MAdult8wks
 Hs.CD14PrimaryCells
 Hs.MobilizedCD4PrimaryCells
 Mm.410bC57Bl6MAdult8wks
 Mm.EpogpmmC01ME14half
 Mm.MgeUknImmortalDRS24h
 Mm.MgeUknImmortal
 Mm.MgeUknImmortalDRS45h
 Mm.MeC57Bl6MAdult8wks
 Mm.LwerS129ME14half
 Mm.Lwer129ME14half
 Mm.LwerC57Bl6ME14half
 Mm.EcomycC01ME14half
 Mm.EpoppC01ME14half
 Mm.EpogpC01ME14half
 Mm.LwerC57Bl6MAdult8wks
 Mm.KidneyC57Bl6MAdult8wks
 Hs.SmallIntestine
 Hs.FetalIntestineLarge
 Hs.FetalIntestineSmall
 Hs.CD3PrimaryCells
 Mm.LgntC57Bl6MAdult8wks
 Hs.FetalStomach
 Hs.Pancreas
 Hs.Gastric
 Mm.HearC57Bl6MAdult8wks
 Mm.SkmuscleC57Bl6MAdult8wks
 Hs.PoodMuscle
 Hs.PenisForeskinFibroblastPrimar
 Mm.LungC57Bl6MAdult8wks
 Mm.FatC57Bl6MAdult8wks
 Mm.GbaC57Bl6MAdult8wks
 Hs.FetalPituitary
 Hs.H1BM4DerivedTrophoblastCu
 Hs.BreastHMEC
 Hs.PenisForeskinKeratinocytePrim
 Hs.PenisForeskinMelanocytePrim
 Mm.S134RImmortal
 Mm.FibroblastC57Bl6MAdult8wks
 Mm.Ni3S2NImmortal
 Mm.FatskSptb6Immortal
 Hs.IMR90
 Hs.H1DerivedMesenchymalStemC
 Hs.FibroblastsFetalSkinScalp
 Hs.FibroblastsFetalSkinBack
 Hs.FibroblastsFetalSkinQuadricep
 Hs.FibroblastsFetalSkinQuadricep
 Hs.FibroblastsFetalSkinBeepsLeft
 Hs.FibroblastsFetalSkinBeepsRight
 Hs.FibroblastsFetalSkinQuadricep
 Hs.FibroblastsFetalSkinAbdomen
 Hs.FibroblastsFetalSkinUpperBack

■ Brain
■ Retina
■ Epithelial
■ Renal
■ Other
■ Lung
■ Muscle
■ Heart
■ ES cells
■ IPSC
■ Adrenal
■ Blood
■ Liver
■ Digestive
■ Fat
■ Epithelial

Select reciprocal-best-hit: 11 matched Human-Mouse sample pairs

Human **ROADMAP** tissue/cell-type

Small_Intestine
 Fetal_Spinal_Cord
 Heart
 Psoas_Muscle
CD19_Primary_Cells 
 iPS_DF_19_7
 Fetal_Brain
 Penis_Foreskin_Fibroblast_Primary_Cells
 Mobilized_CD34_Primary_Cells
 Fetal_Thymus
 Mobilized_CD3_Primary_Cells

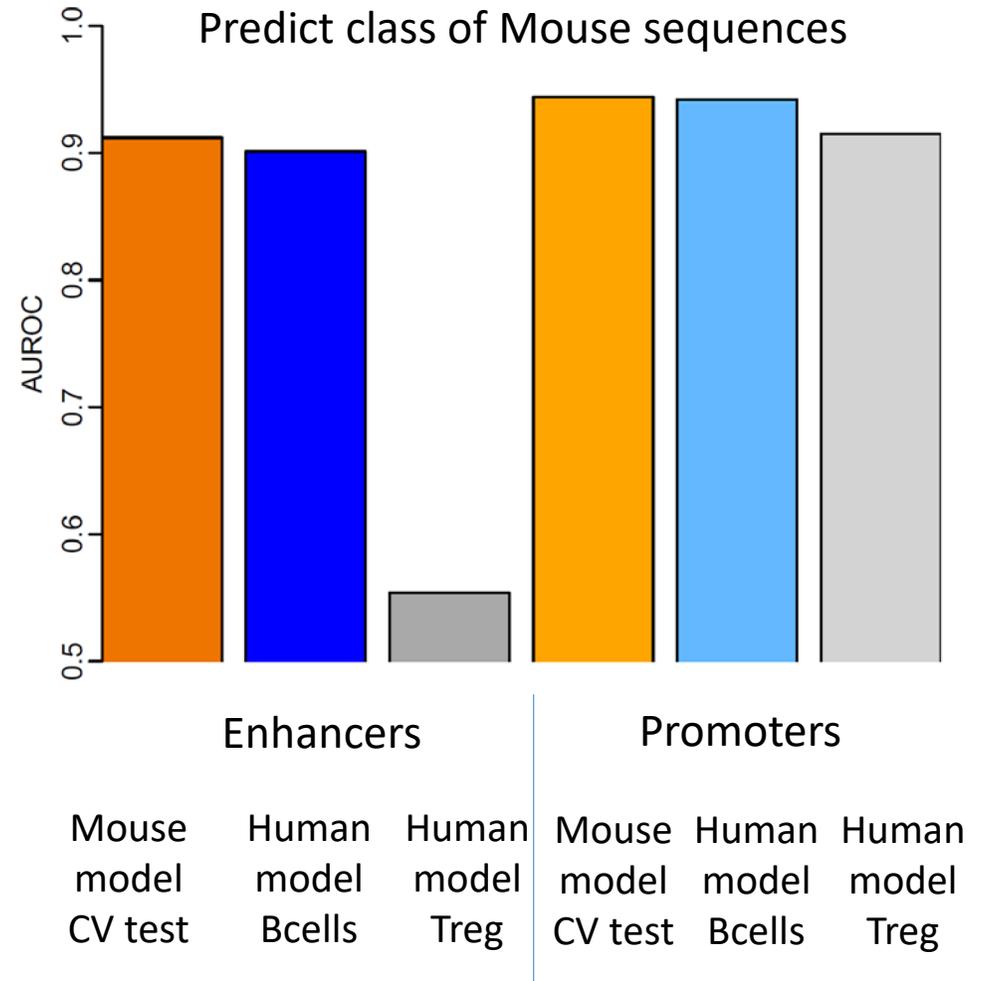
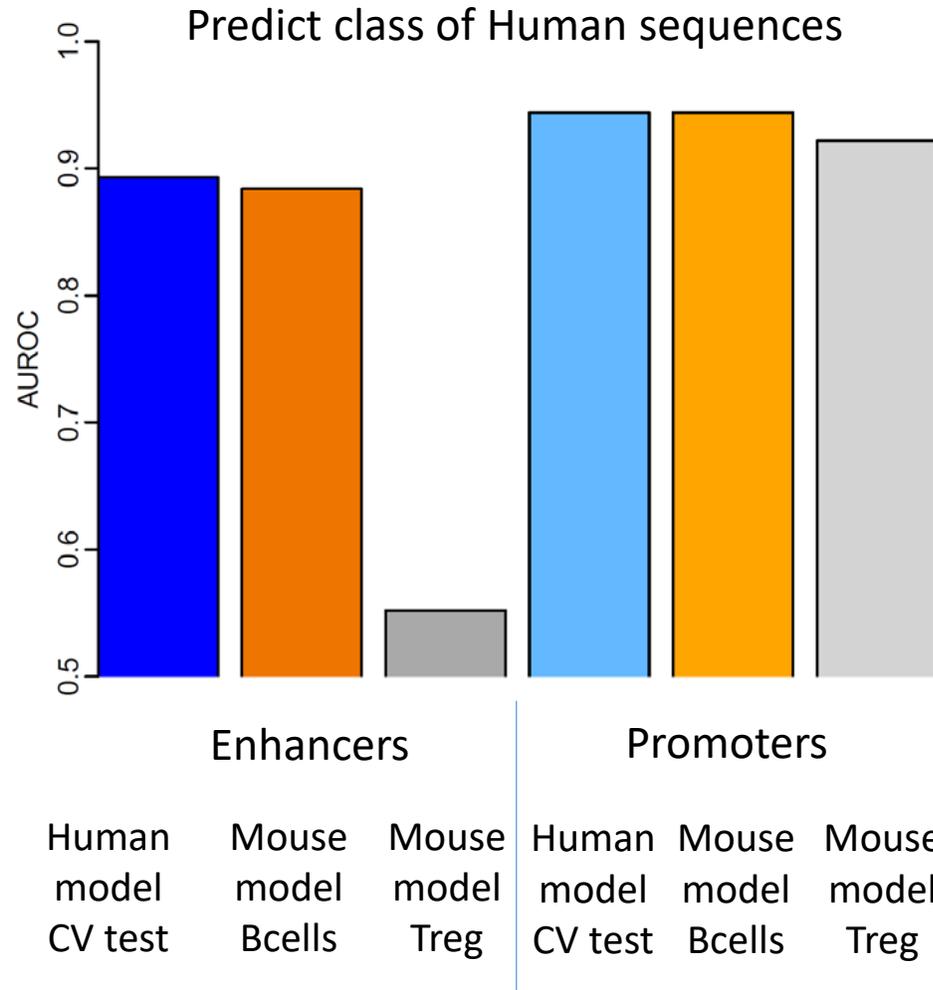
Mouse **ENCODE** tissue/cell-type

Lgint MAdult8wks
 Wholebrain ME14half
 Heart MAdult8wks
 Skmuscle MAdult8wks
Bcell CD19+ MAdult8wks
 ES-CJ7 S129 ME0
 Wbrain ME18half
 Nih3t3 Nihs MImmortal
 EpcpmmCd1ME14half
 Thymus MAdult8wks
 Treg MAdult8wks

Top wts → common TFBS

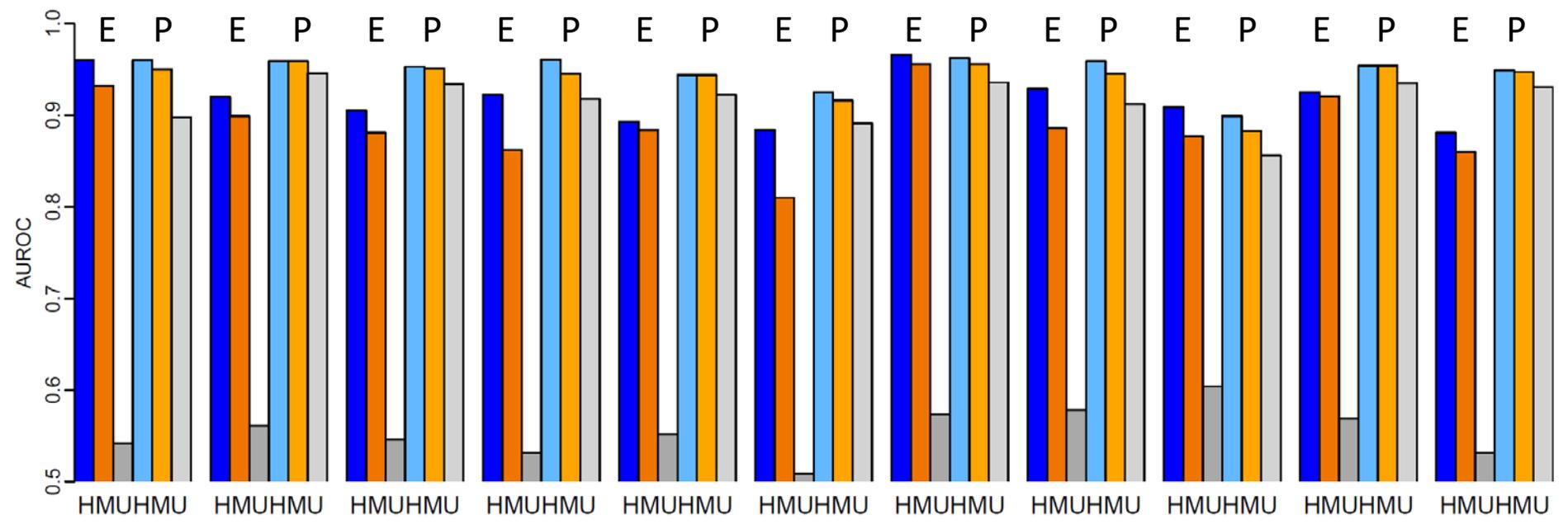
AP1, HNF4a, KLF
 Rfx2, HEB, Chx10
 Mef2, NF1/Tlx, AP1, Gata
 Mef2, AP1, MyoG
 SpiB, Runx1, NFkB, PU.1
 Oct, Sox, Klf, TEAD1
 Rfx2, Chx10, Olig2
 AP1, HEB, Runx1, TEAD1
 SpiB, Runx1, Gata, AP1
 Runx1, Tcf, ETS1
 BATF, ETS1, NFkB

Train gkm-SVM on B Cell regions in one species, score B Cell sequences in other species

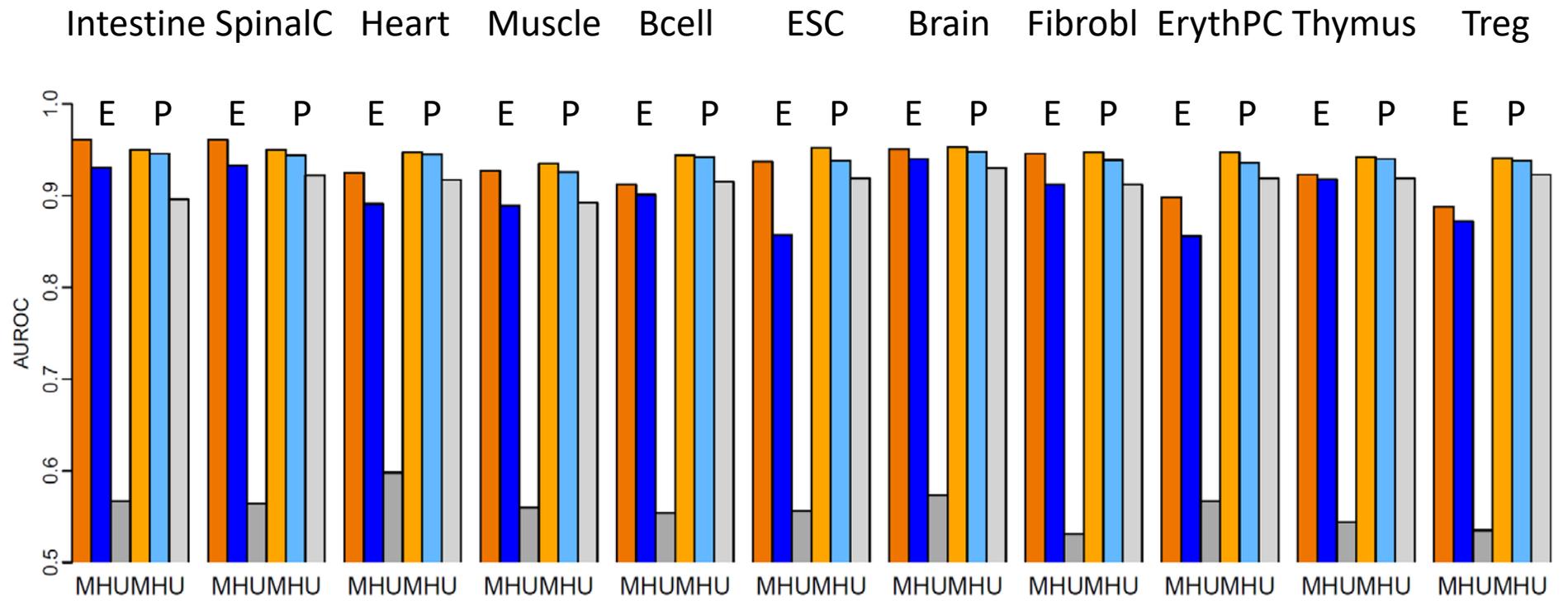


- Both cell-type specific enhancers and promoters are predictable from gapped kmer sequence features
- Regulatory vocabulary of cell types is conserved between human and mouse for **both enhancers and promoters**
- **Enhancer** regulatory vocabulary is cell-specific
- **Promoter** regulatory vocabulary is more cell-type independent

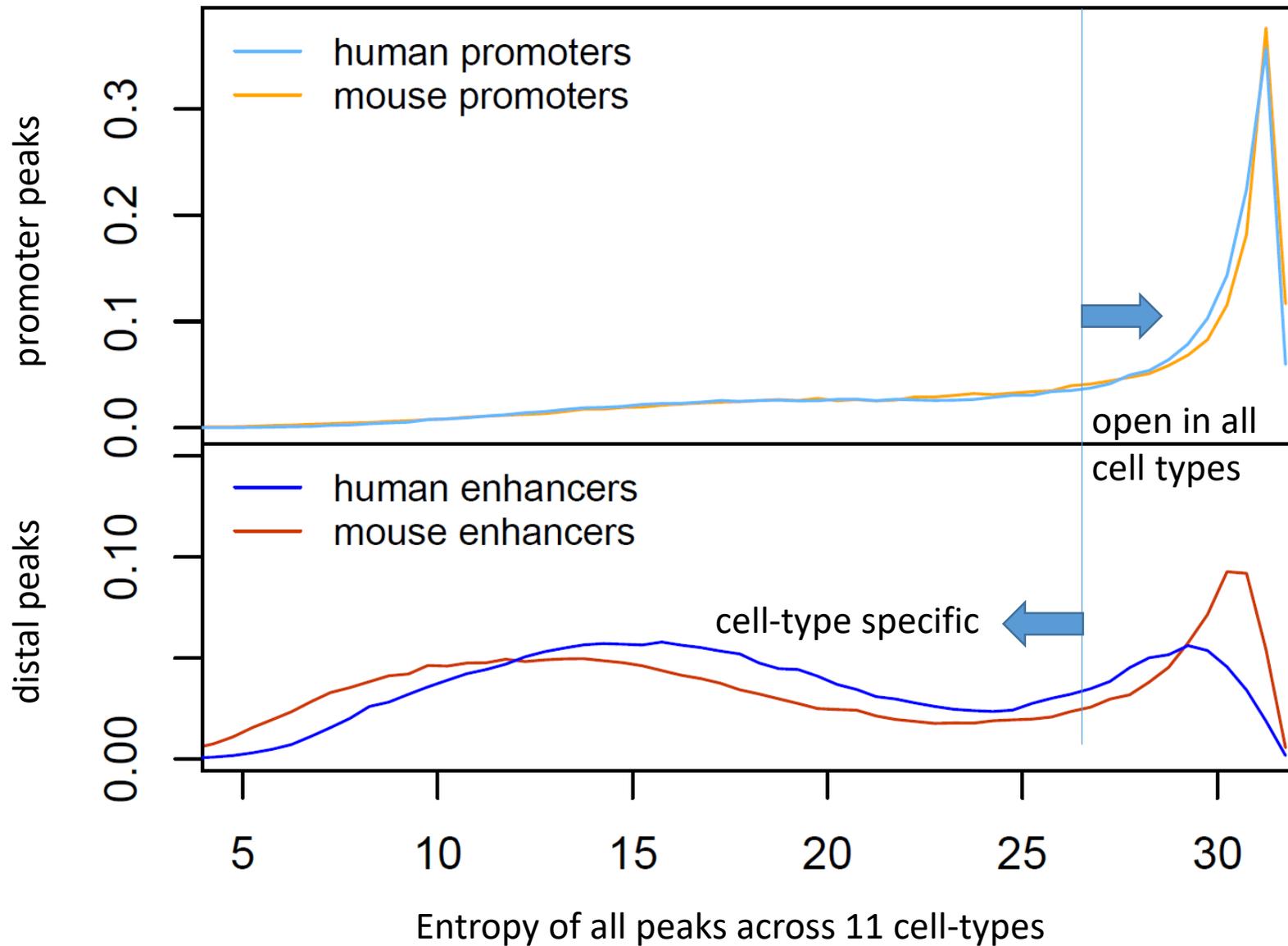
Score Human regions



Score Mouse regions

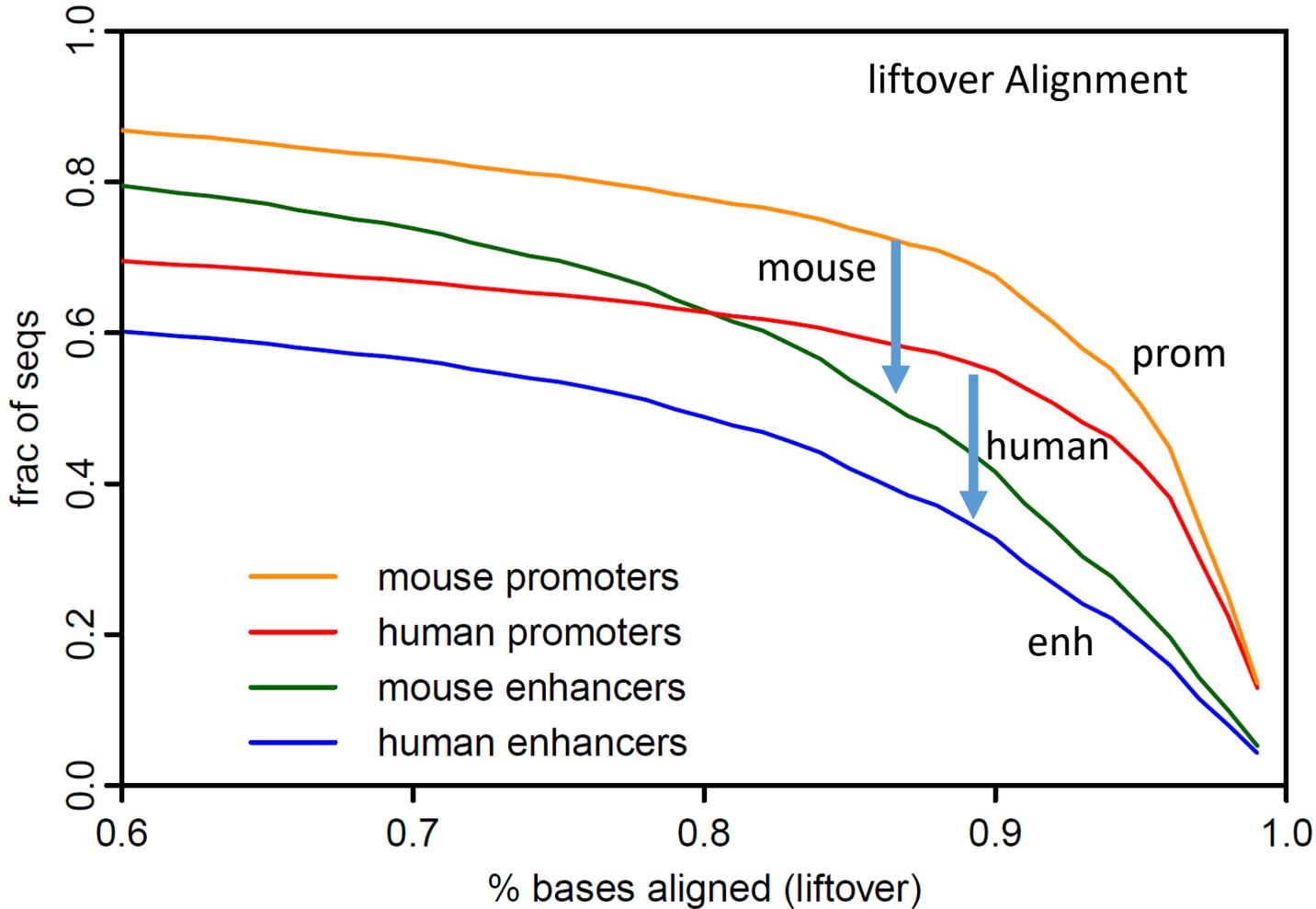


Entropy Distribution Confirms That Promoters Are Generally Open Across All 11 Cell-types



Consistent with
summary statistics of:
Cheng 2014
Vierstra 2014

Although Regulatory Vocabulary of Enh and Prom are Both Conserved,
Enhancers are much less **Alignable** than Promoters



similar conclusions using bnMapper or PhyloP

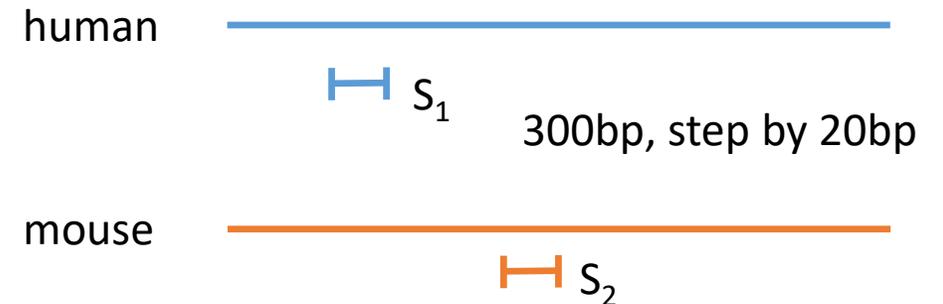
Gapped kmer word composition can detect conservation of enhancers between human and mouse

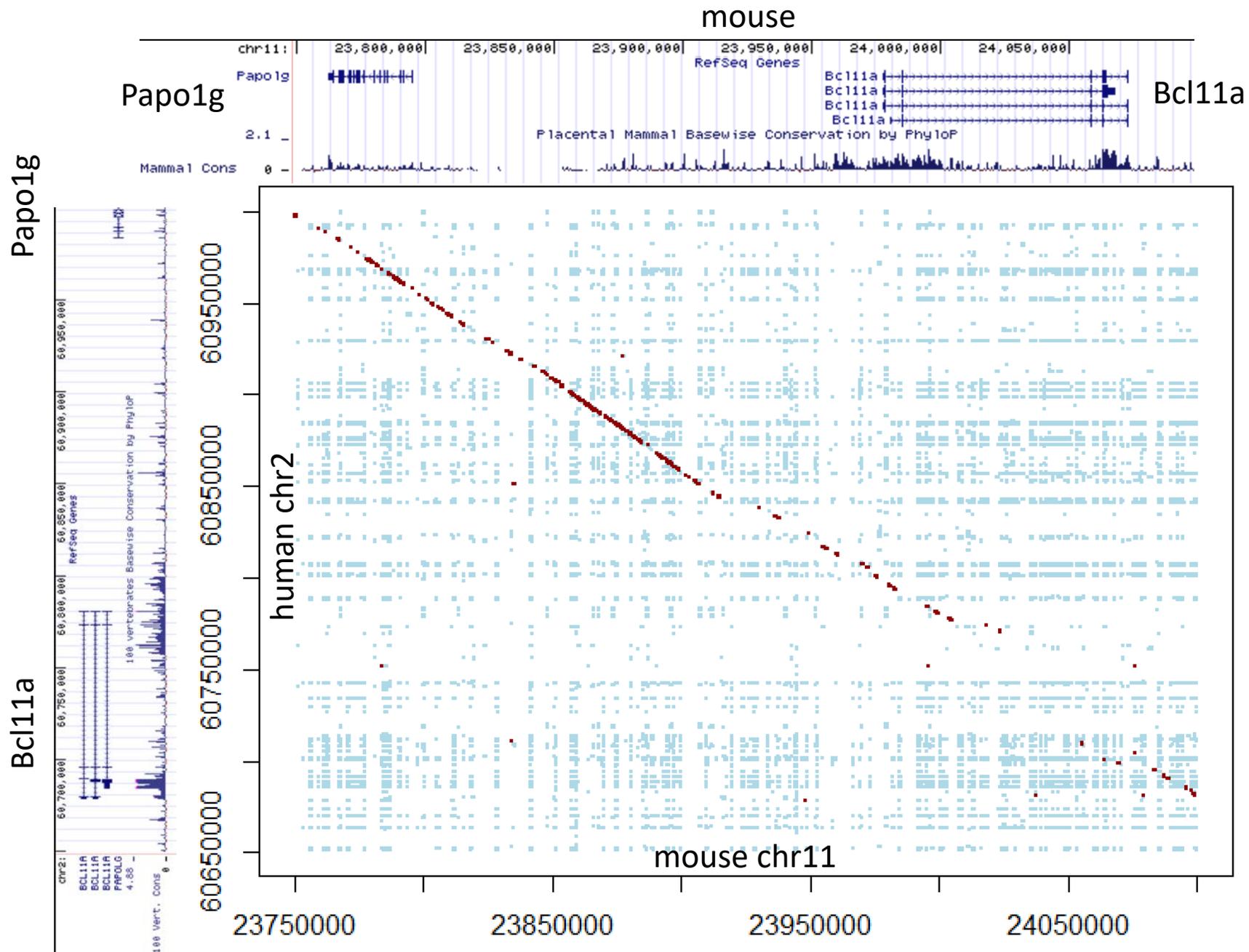
But sequence alignment cannot

Use gapped kmer kernel to detect conservation:

$$K(S_1, S_2) = \frac{\langle f^{S_1}, f^{S_2} \rangle}{\|f^{S_1}\| \|f^{S_2}\|}$$

dot product of normalized gapped kmer count vectors for two sequences S_1 S_2





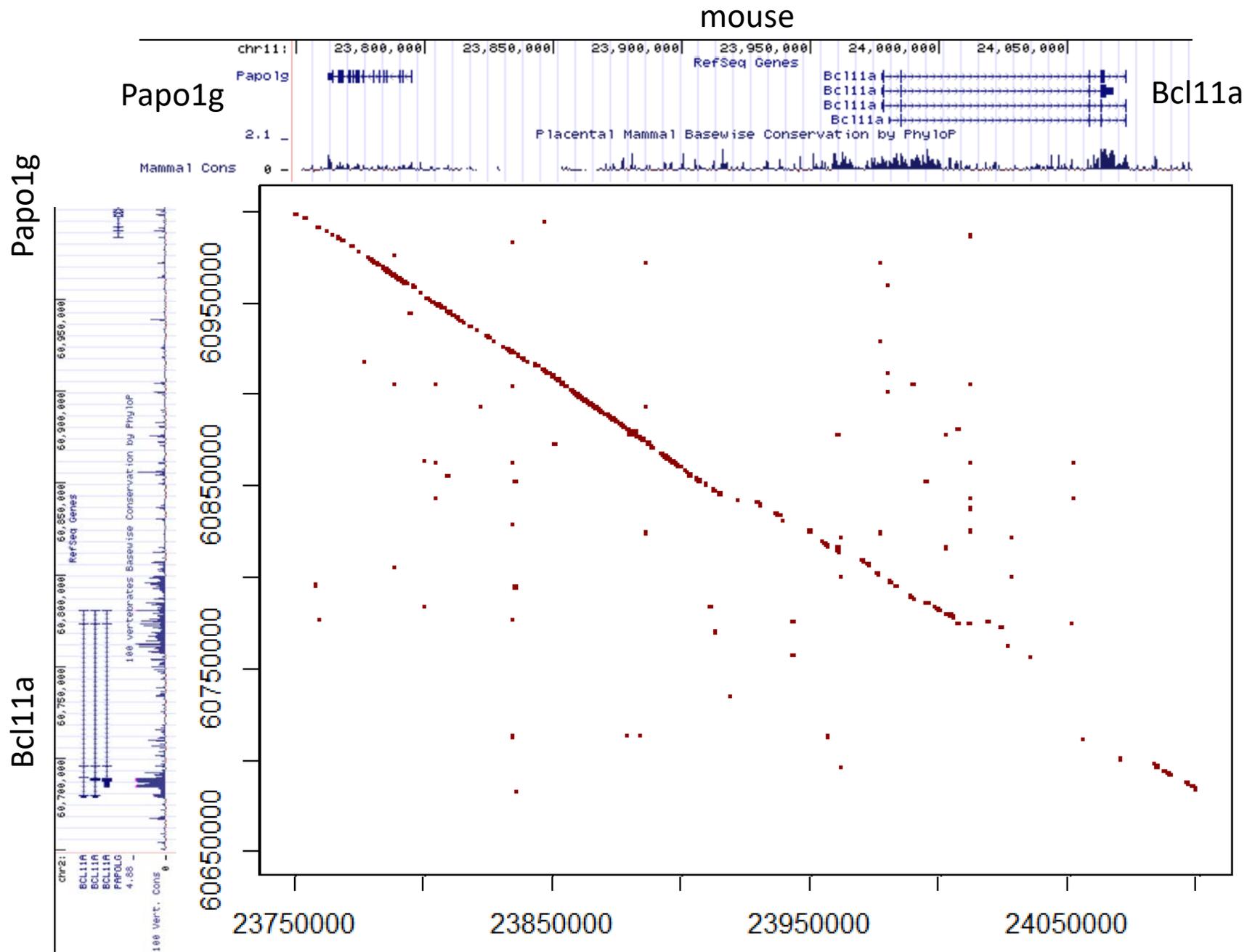
K>0.2

- ≤20 hits
- >20 hits:
SINE
LINE

350kb

>300,000,000 comparisons of two 300bp seqs

diagonal = syntenic blocks of similar gapped kmer composition

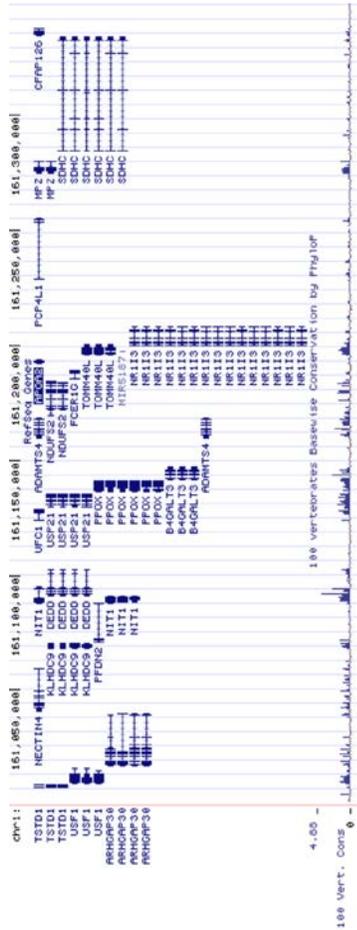


Better to remove repeats with PCA

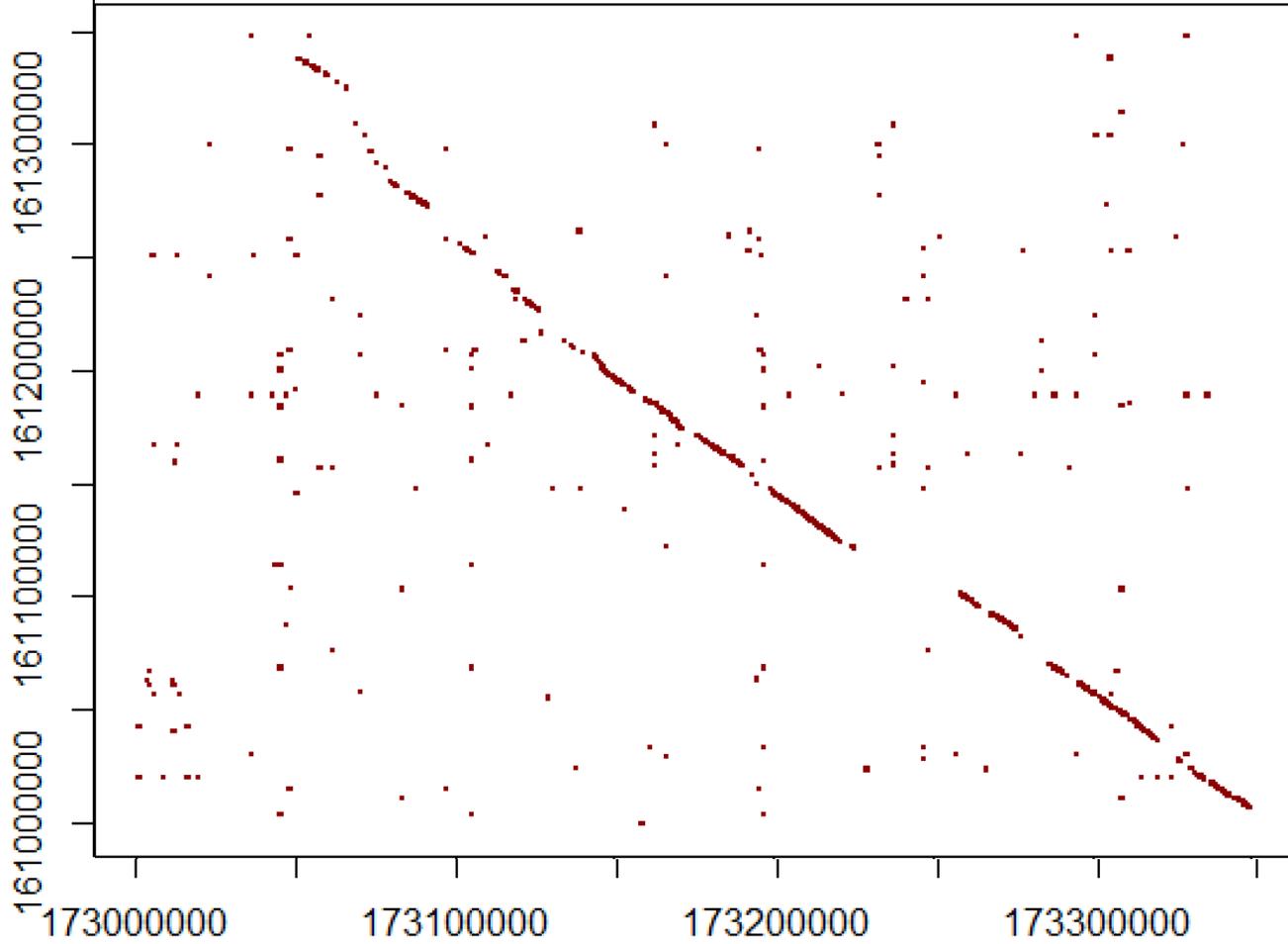
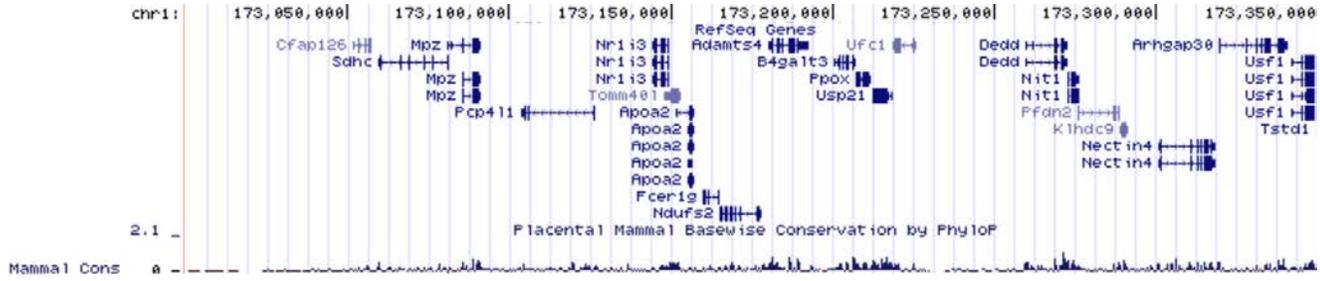
350kb

diagonal = syntenic blocks of similar gapped kmer composition

human Apoa2



mouse Apoa2



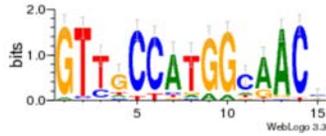
diagonal = syntenic blocks of similar gapped kmer composition

Mouse gkmSVM regulatory vocabulary interprets human GWAS Schizophrenia SNP

Schizophrenia SNP has large deltaSVM when gkm-SVM trained on

- adult mouse dentate gyrus ATAC-seq (Song Nature 2017)
- or midbrain DA neurons ATAC-seq (McCallion Lab)

clearly disrupts RFX BS:



rs1498232

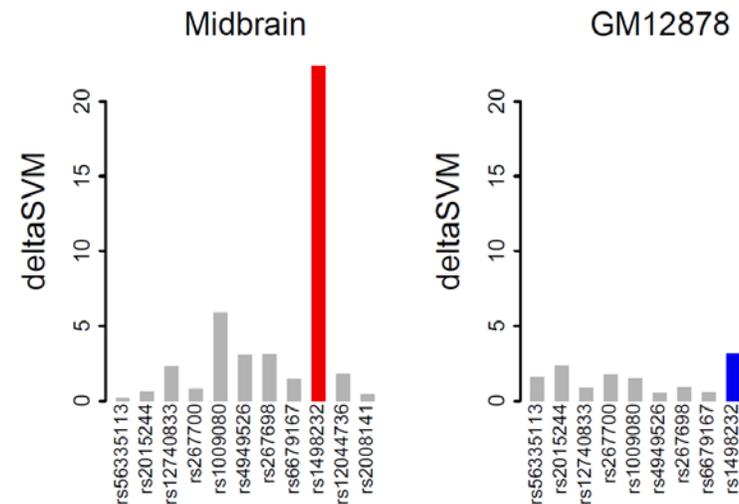
Ref allele: CCGTTTCCATGGCAACCAG 0.53

Alt allele: CCGTTTCCACGGCAACCAG 0.47

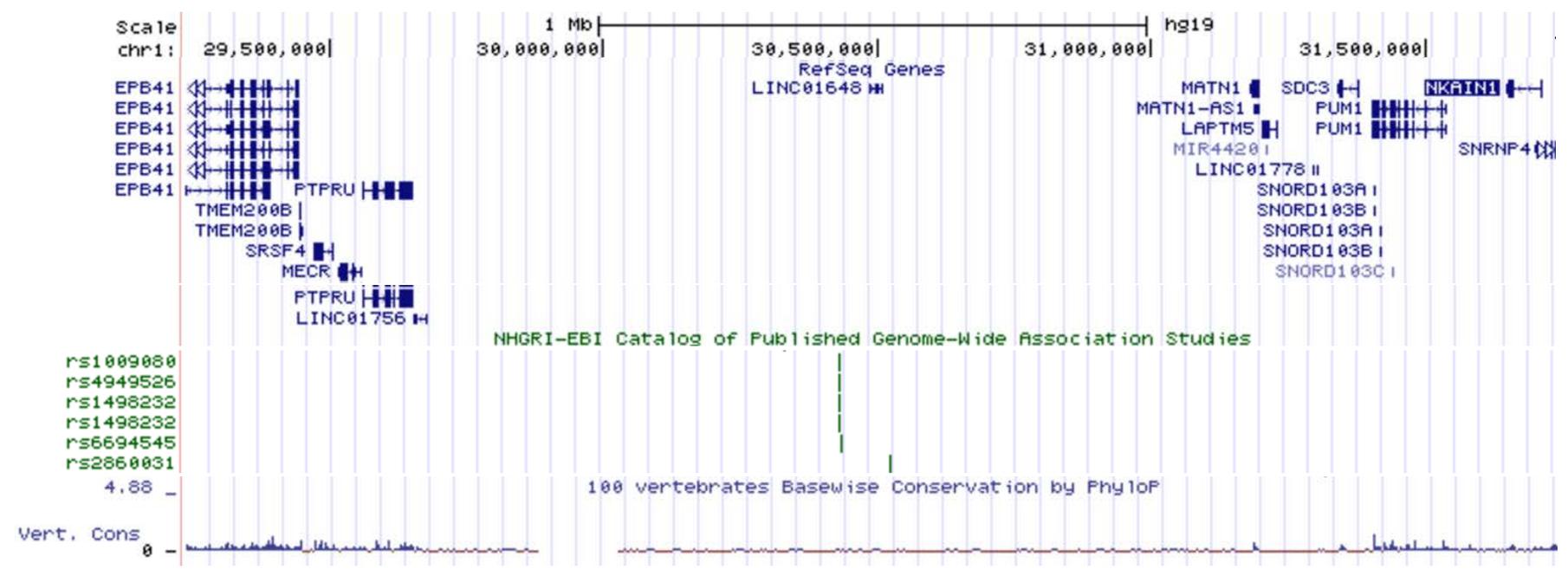
Ref 10-mer	wt	Alt 10-mer	wt	Diff
CCGTTTCCAT	1.27	CCGTTTCCAC	0.25	1.02
CGTTTCCATG	2.07	CGTTTCCACG	0.75	1.32
GTTTCCATGG	6.08	GTTTCCACGG	2.20	3.89
TTTCCATGGC	2.86	TTTCCACGGC	1.01	1.85
TTCATGGCA	2.15	TTCACGGCA	0.76	1.39
TCCATGGCAA	3.66	TCCACGGCAA	1.32	2.35
CCATGGCAAC	7.93	CCACGGCAAC	2.84	5.09
CATGGCAACC	4.31	CACGGCAACC	1.61	2.70
ATGGCAACCA	2.45	ACGGCAACCA	0.90	1.55
TGGCAACCAG	1.96	CGGCAACCAG	0.77	1.19

deltaSVM=-22.35

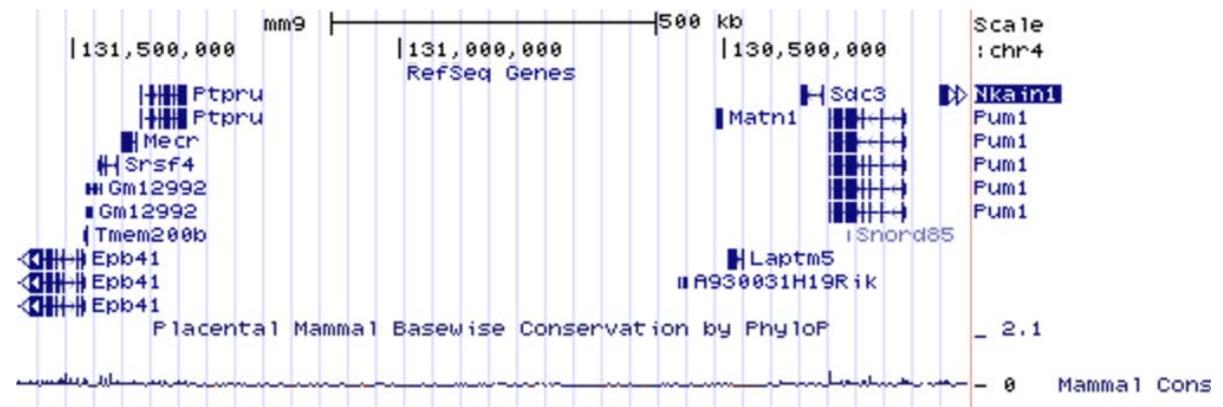
CCATGGCAAC	7.927474
CCATGGAAAC	6.0849012
CCATAGCAAC	5.8148892
CCATGGTAAC	5.515136
CATGGCAACC	4.3127716
CATGGCAACA	4.298156
CTATGGCAAC	4.1949162
CCTTGGCAAC	3.8884124
CCCTGGCAAC	3.7884638
CCATGGGAAC	3.7854512
CCATGACAAC	3.7819106
CTATGGAAAC	3.729401
TCCATGGCAA	3.6644538
...	



human locus



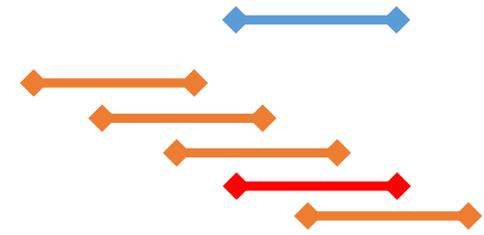
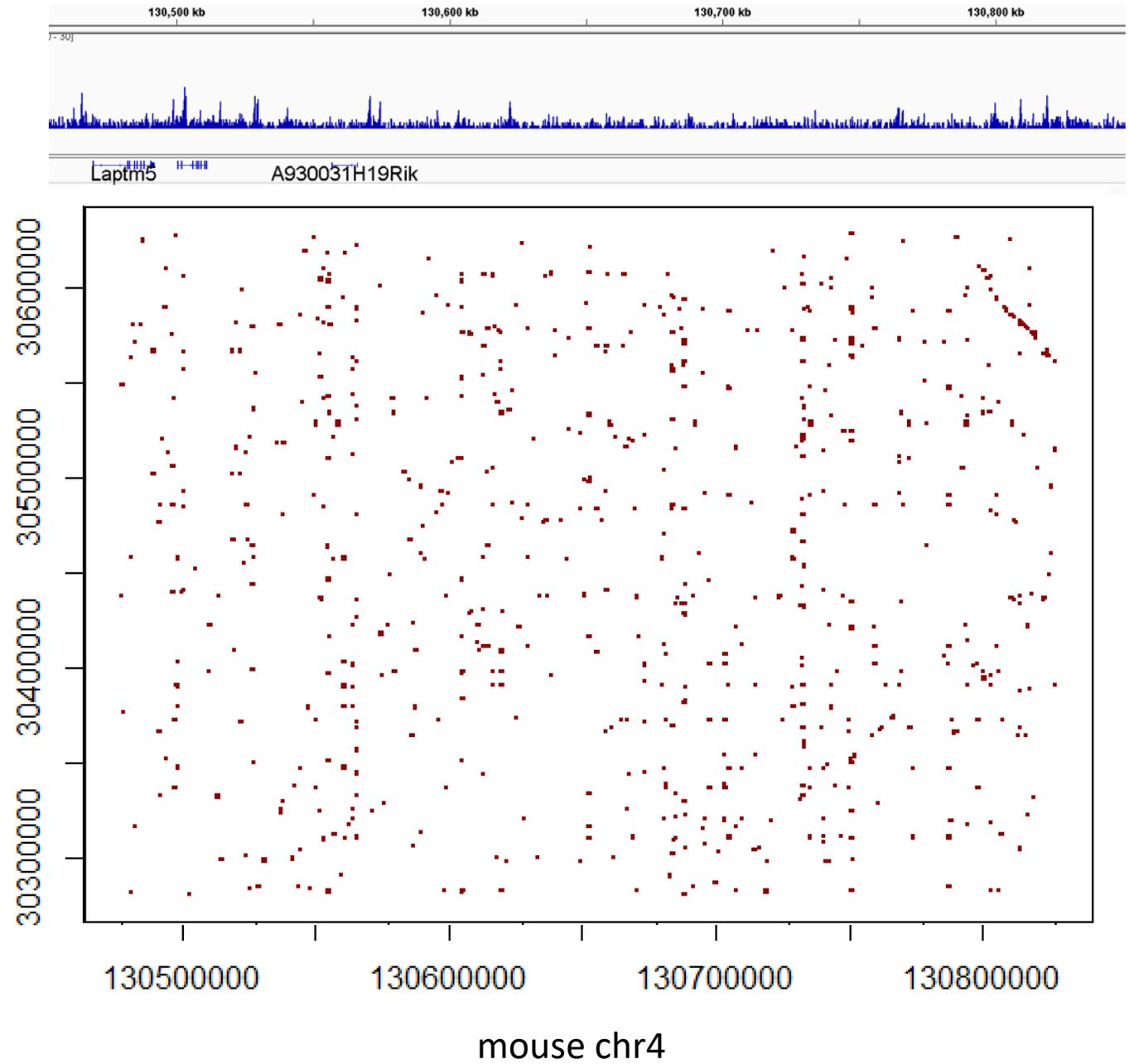
mouse locus



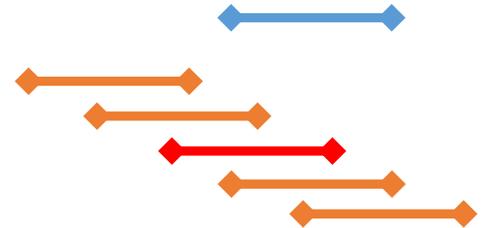
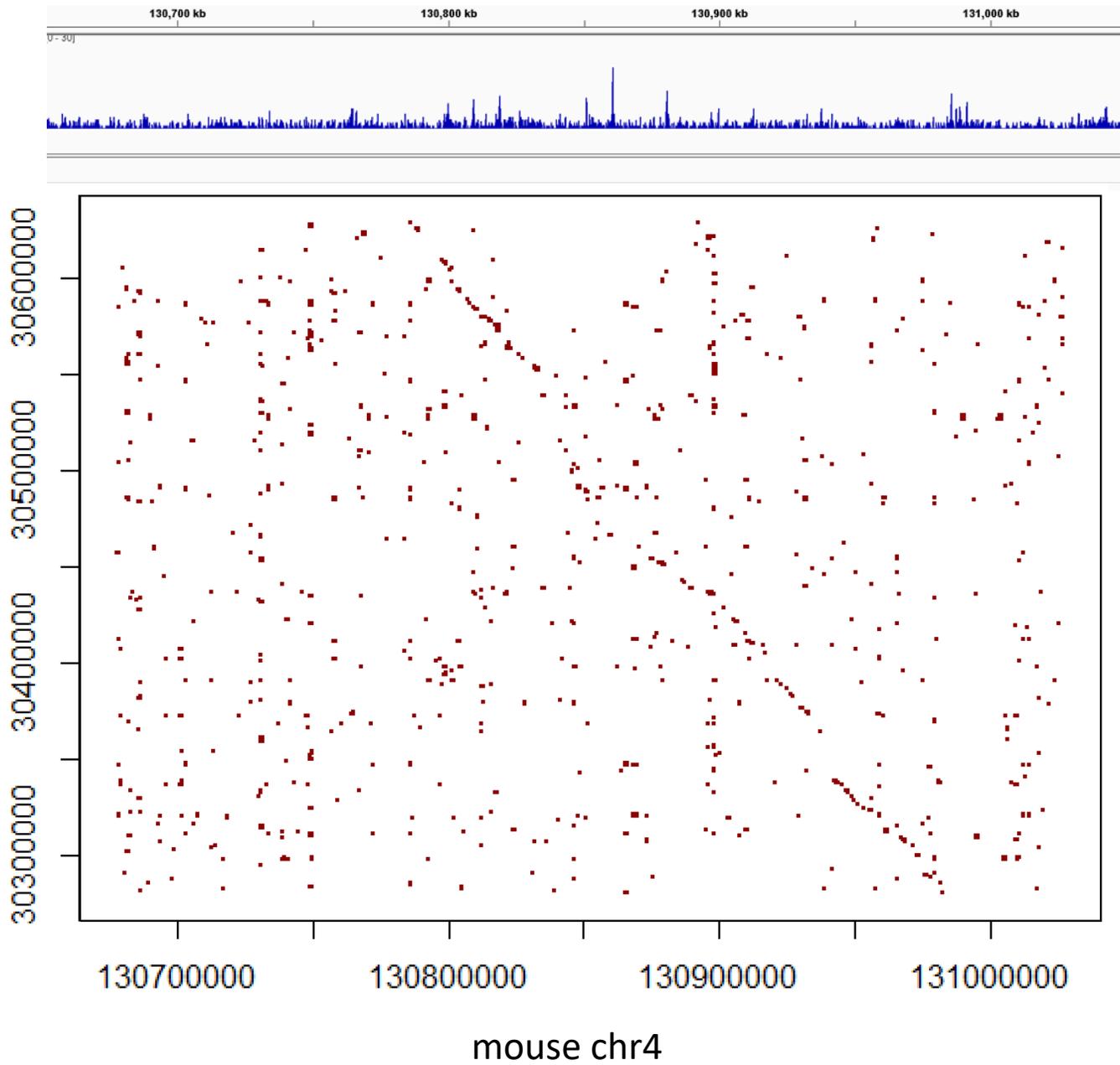
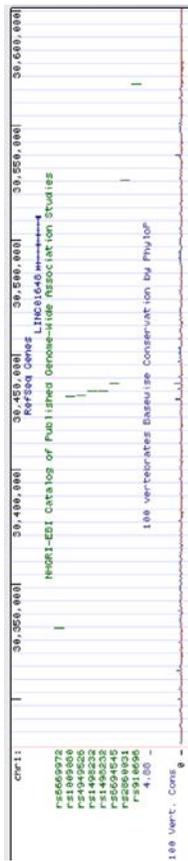
adult mouse
dentate gyrus ATAC-seq



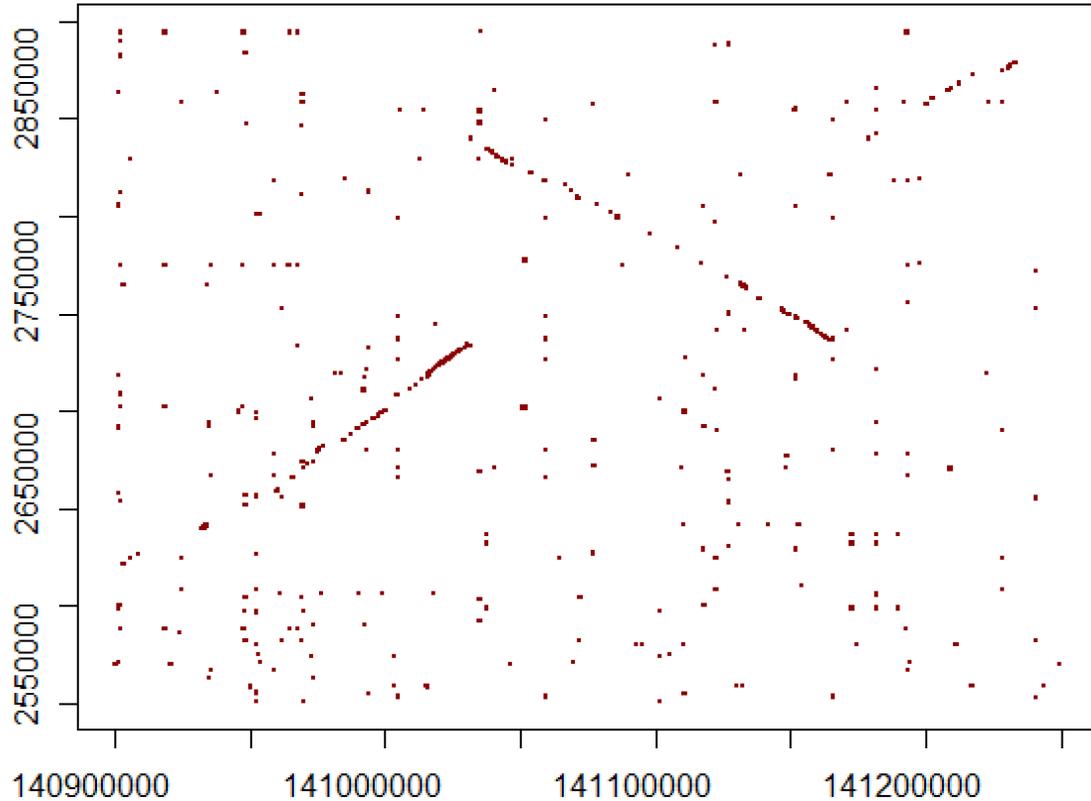
human Schizophrenia locus



human Schizophrenia locus



gkm-SVM detects inversion in gna12 locus



Summary:

- Cell-specific enhancers and promoters are predictable from gapped kmer sequence features
- Regulatory vocabulary of cell types is conserved between human and mouse for **both enhancers and promoters**
- **Enhancer** regulatory vocabulary is cell-specific
- **Promoter** regulatory vocabulary more cell-type independent
- **gkm-SVM** kernel can detect syntenic blocks of conserved gapped-kmer composition, independent of cell type

Future directions:

- Use segment detection to map conserved chains
- Generalize across multiple species
- Develop heuristic algorithm to apply genome wide