

# JClub

Jan 26, 2017

- Wu et al., *Nature Neurosci*, 2016  
Genome-wide, integrative analysis implicates microRNA dysregulation in autism spectrum disorder
- Parikshak et al., *Nature*, 2016  
Genome-wide changes in lncRNA, splicing, and regional gene expression patterns in autism

## Autism Spectrum Disorders

- ASD now includes several conditions that used to be diagnosed separately: autistic disorder, pervasive developmental disorder not otherwise specified (PDD-NOS), Asperger syndrome, and childhood disintegrative disorder
- These disorders involve a wide range of cognitive and behavioral abnormalities: social deficits and communication difficulties, repetitive behaviors and interests, sensory issues, deviance in language development, poor motor skills' and in some cases, cognitive delays.
- About 1 percent of the world population has autism spectrum disorder. (CDC,2014)
- Prevalence in the United States is estimated at 1 in 68 births. (CDC, 2014)
- Prevalence of autism in U.S. children increased by 119.4 percent from 2000 (1 in 150) to 2010 (1 in 68). (CDC, 2014) Autism is the fastest-growing developmental disability. (CDC, 2008)
- Cost of lifelong care can be reduced by 2/3 with early diagnosis and intervention. (Autism Society estimate)
- ASD is almost 5 times more common among boys (1 in 42) than among girls (1 in 189)

# Genome-wide, integrative analysis implicates microRNA dysregulation in autism spectrum disorder

Ye E Wu<sup>1,2</sup>, Neelroop N Parikshak<sup>1,3,4</sup>, T Grant Belgard<sup>1,2,6</sup> & Daniel H Geschwind<sup>1,2,4,5</sup>

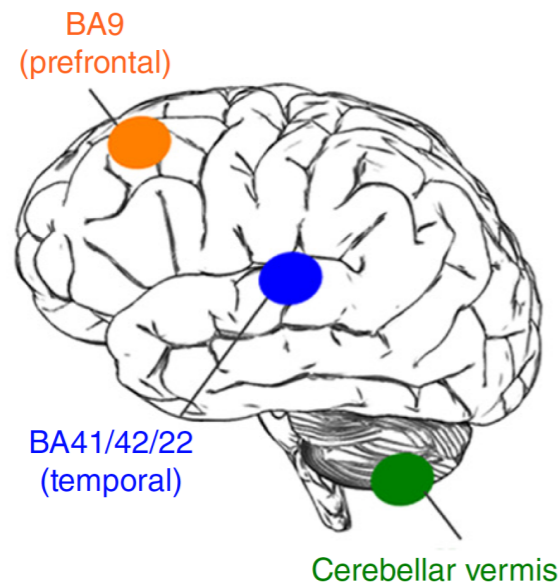
Genetic variants conferring risk for autism spectrum disorder (ASD) have been identified, but the role of post-transcriptional mechanisms in ASD is not well understood. We performed genome-wide microRNA (miRNA) expression profiling in post-mortem brains from individuals with ASD and controls and identified miRNAs and co-regulated modules that were perturbed in ASD. Putative targets of these ASD-affected miRNAs were enriched for genes that have been implicated in ASD risk. We confirmed regulatory relationships between several miRNAs and their putative target mRNAs in primary human neural progenitors. These include hsa-miR-21-3p, a miRNA of unknown CNS function that is upregulated in ASD and that targets neuronal genes downregulated in ASD, and hsa\_can\_1002-m, a previously unknown, primate-specific miRNA that is downregulated in ASD and that regulates the epidermal growth factor receptor and fibroblast growth factor receptor signaling pathways involved in neural development and immune function. Our findings support a role for miRNA dysregulation in ASD pathophysiology and provide a rich data set and framework for future analyses of miRNAs in neuropsychiatric diseases.

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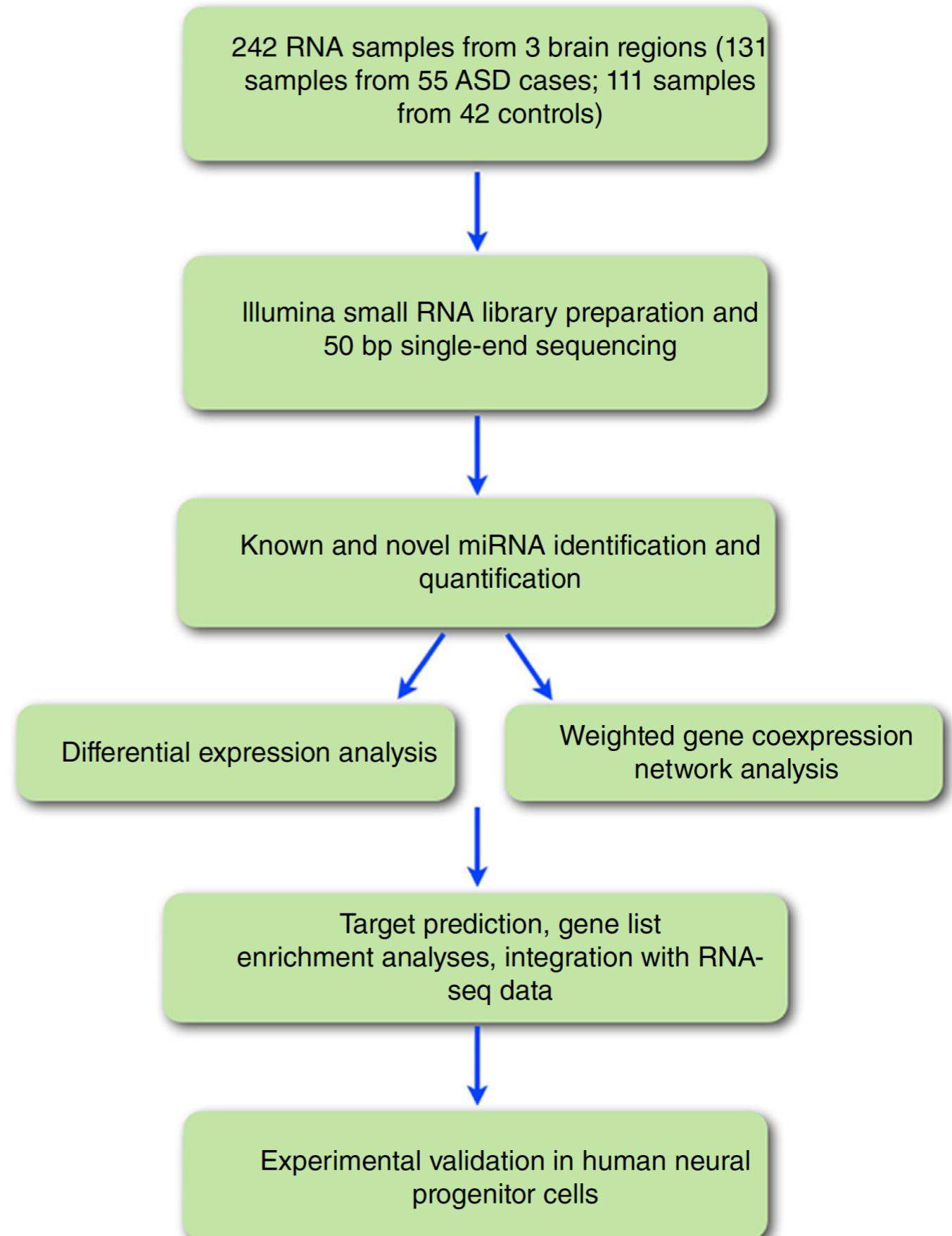
Received 16 January; accepted 29 July; published online 29 August 2016; doi:10.1038/nn.4373

## Flow chart of the overall approach

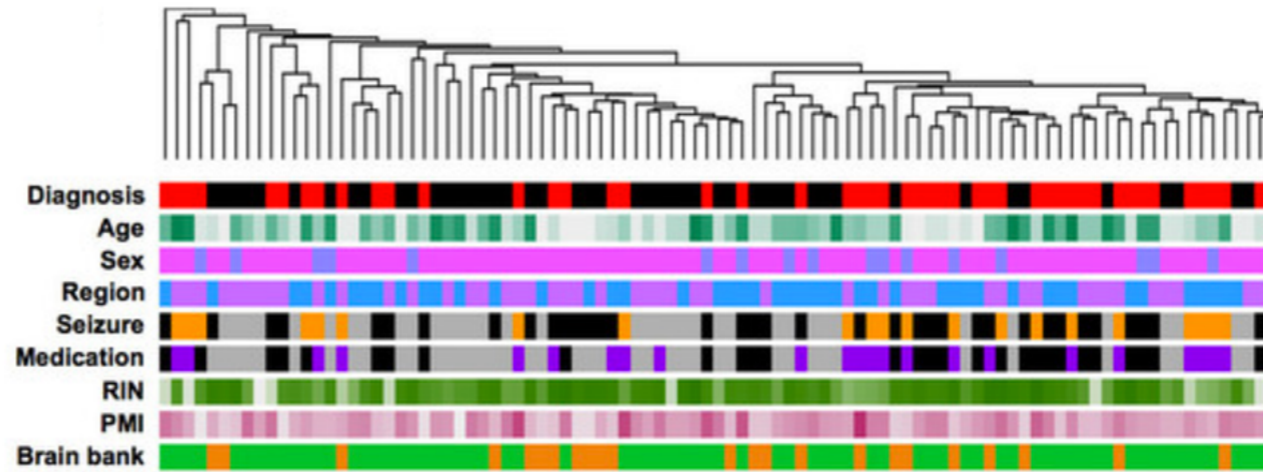
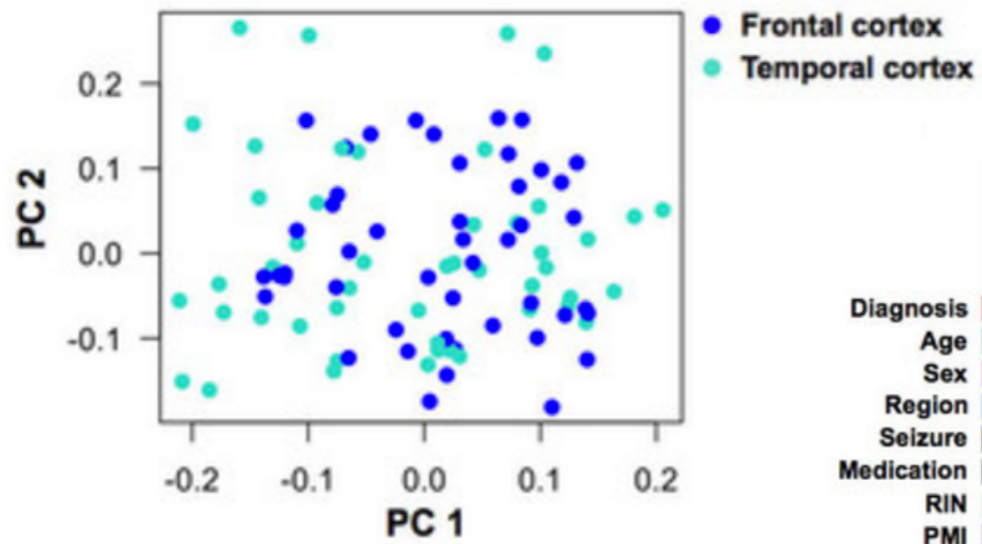
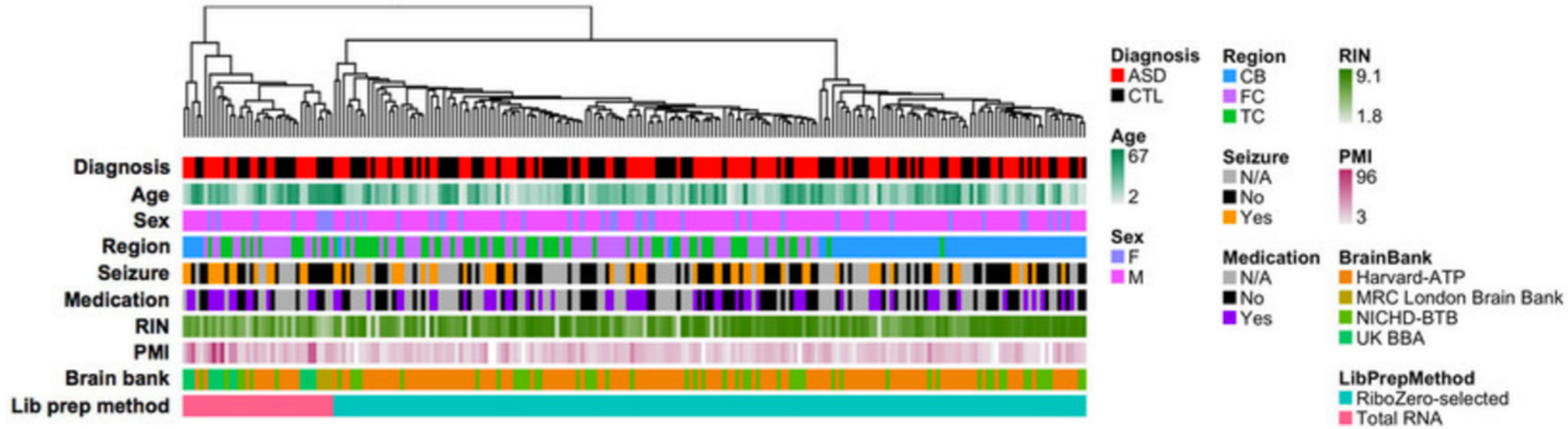
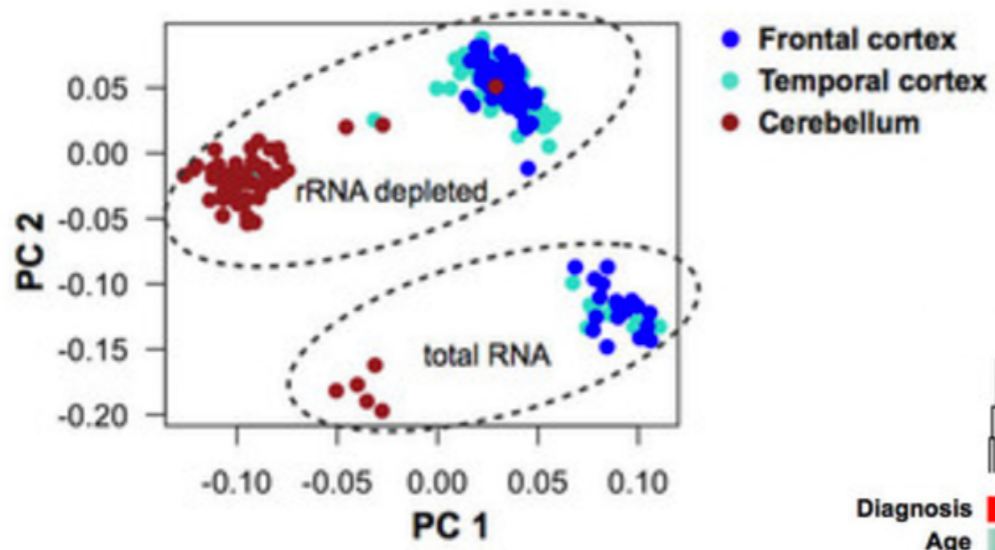


up to 3 brain regions (frontal cortex, temporal cortex, cerebellar vermis) implicated with ASD from **242** post-mortem brain samples

identified and quantified **699** miRNAs (552 documented in *miRBase* and 147 unknown)

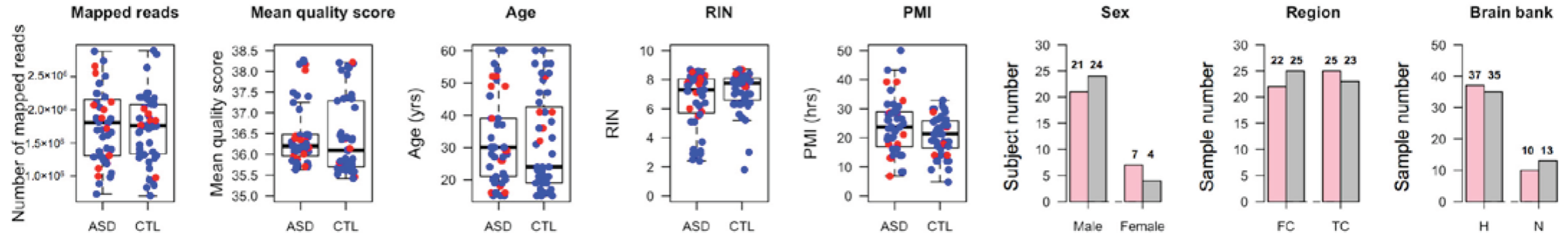


# PCA and hierarchical sample clustering

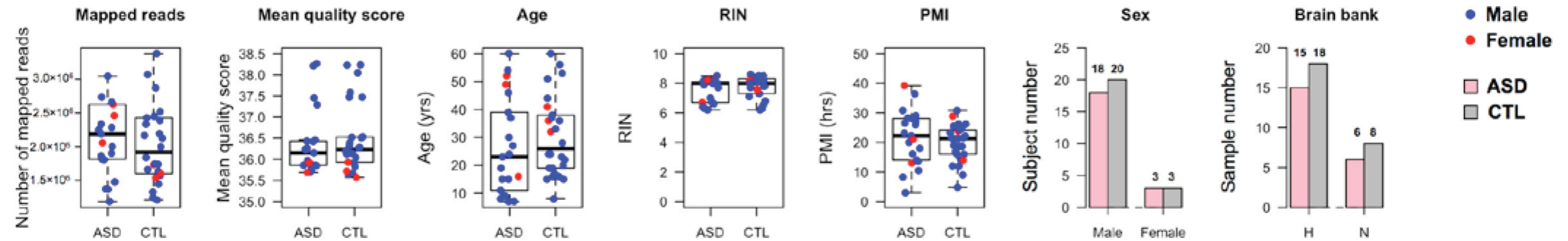


# Characteristics of samples used in analyses

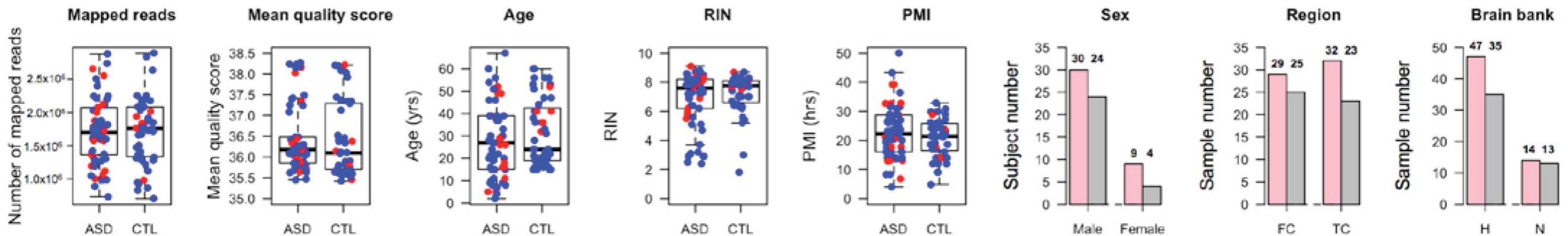
## 95 cortex samples for DGE analysis



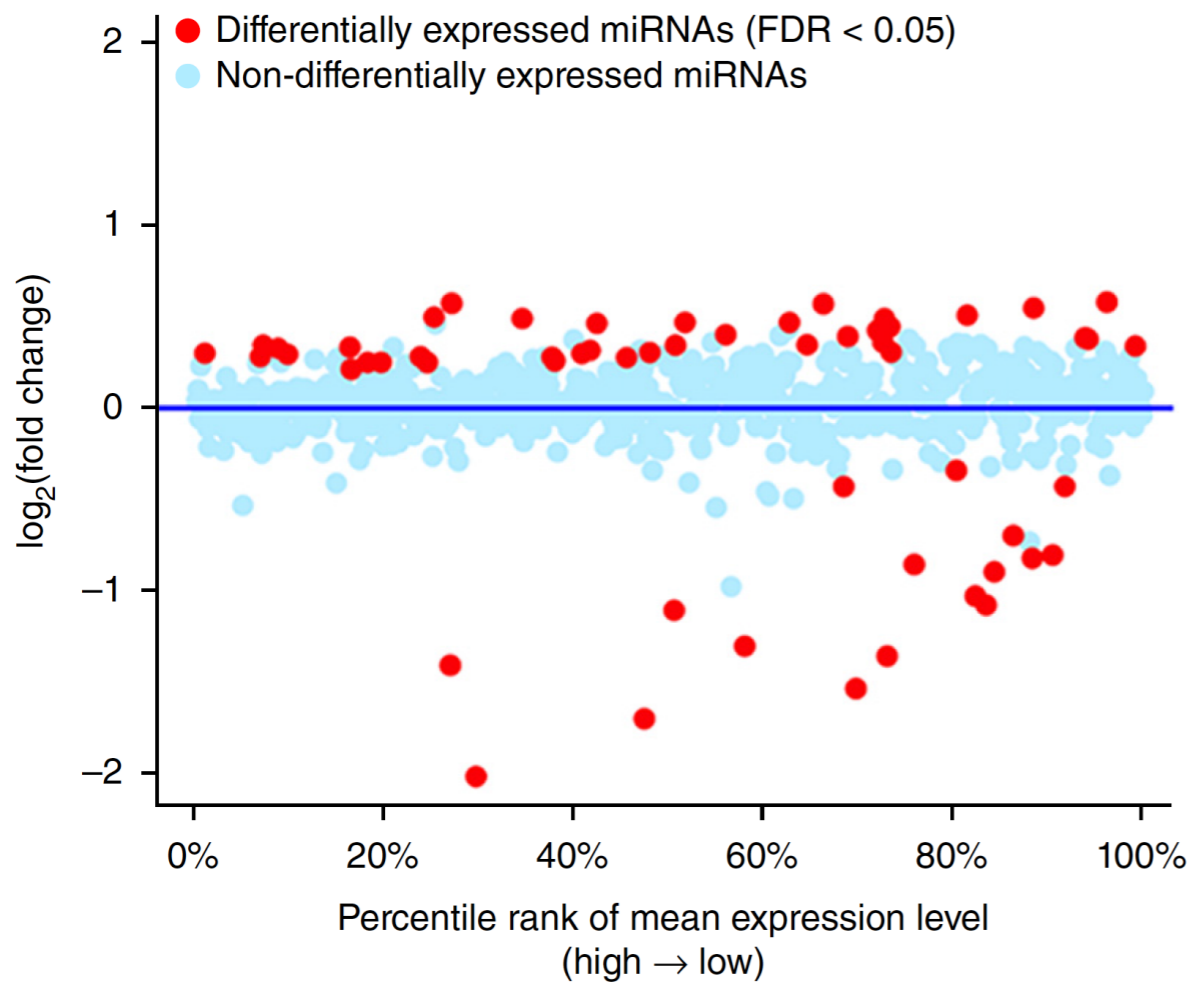
## 47 cerebellum samples for DGE analysis



## 109 cortex samples for WGCNA

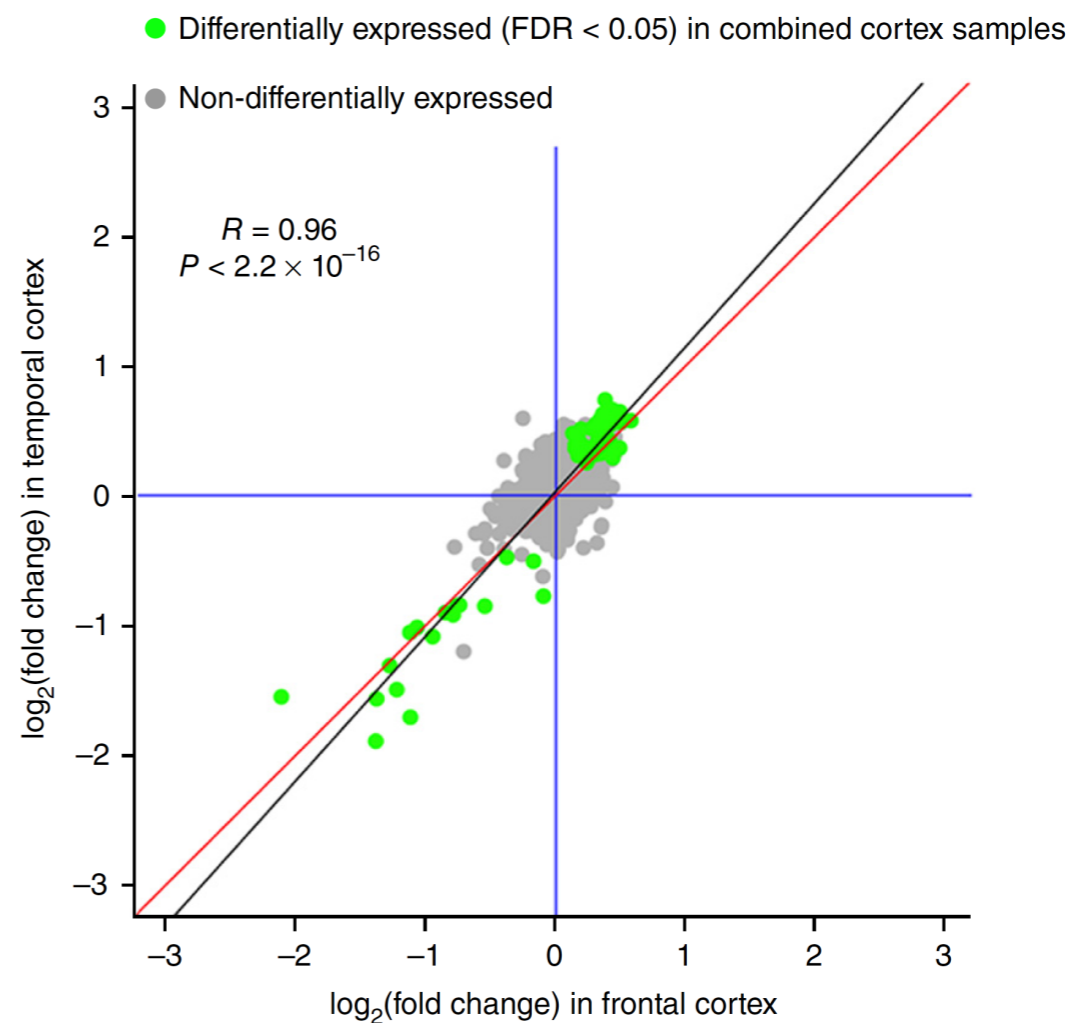


## Differential expression analyses



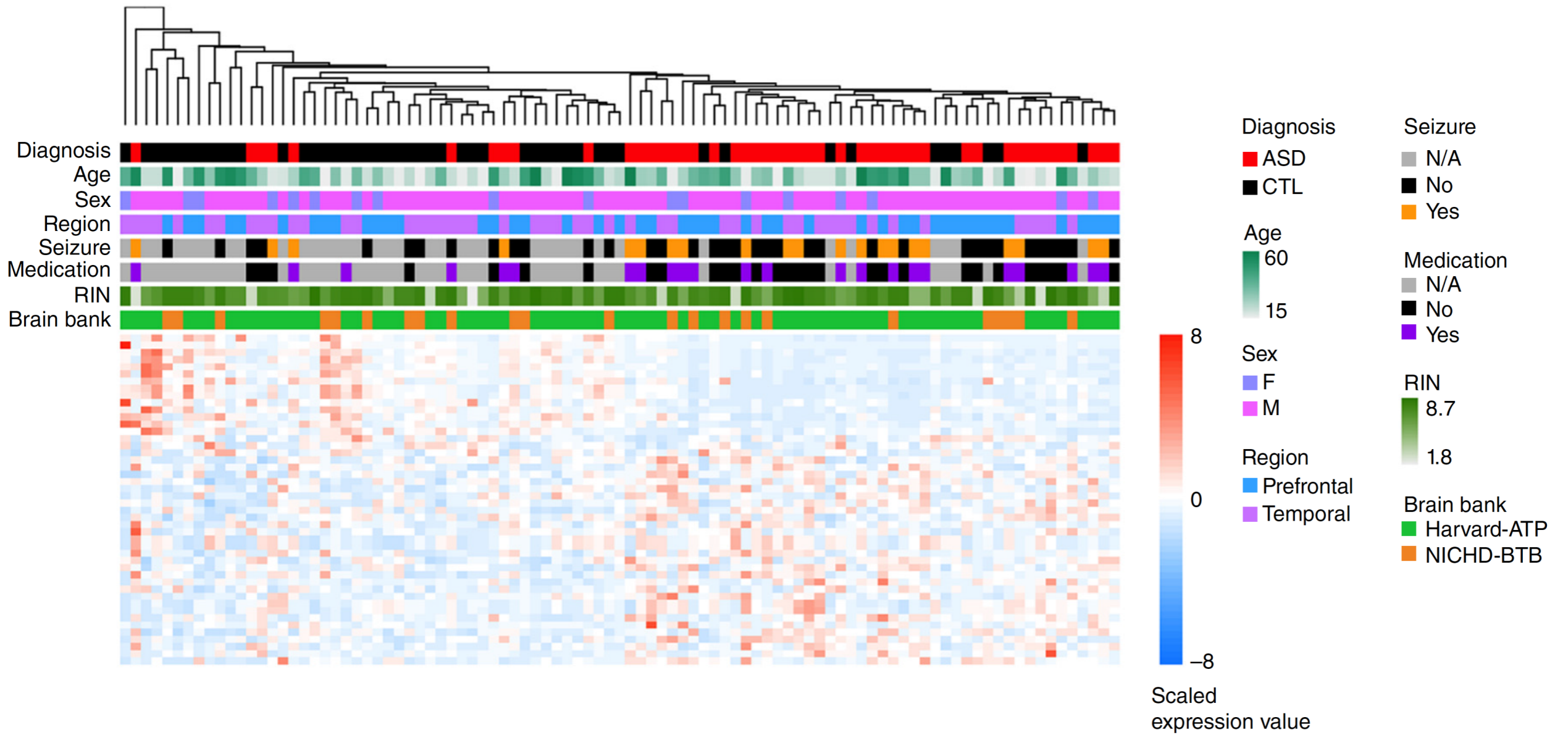
Concordance between FC and TC fold changes

**17** down-regulated and **41** up-regulated miRNA in ASD cortex compared to control



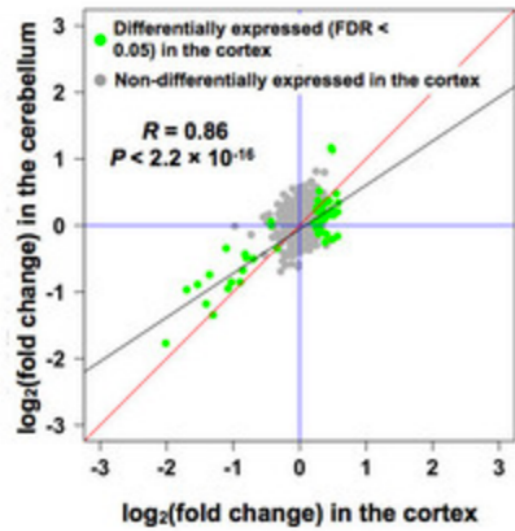
# Hierarchical clustering of 95 cortex samples based on top DE miRNAs (FDR<0.05; abs(log2FC) > 0.3)

FDR<0.05; abs(log2FC) > 0.3

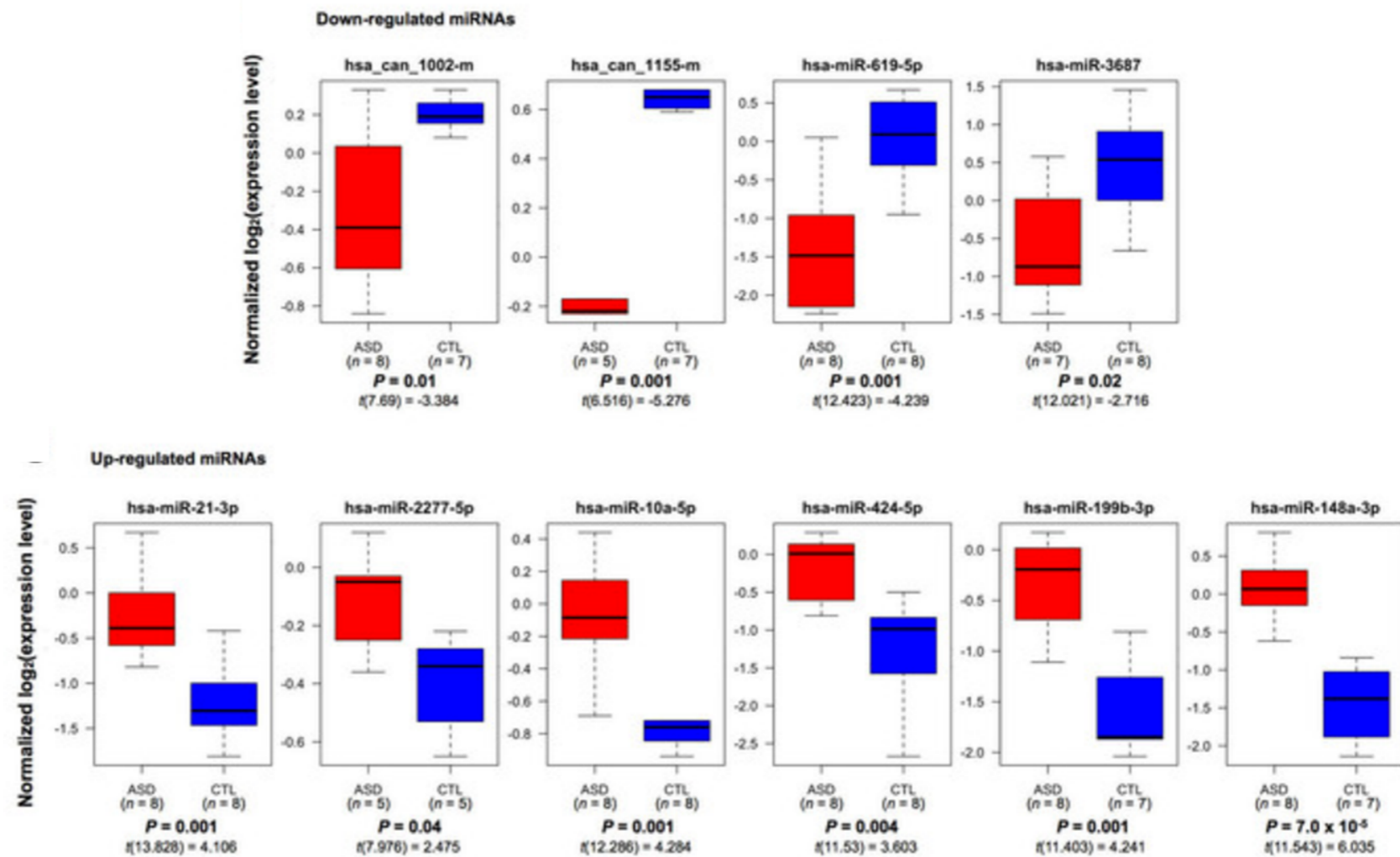




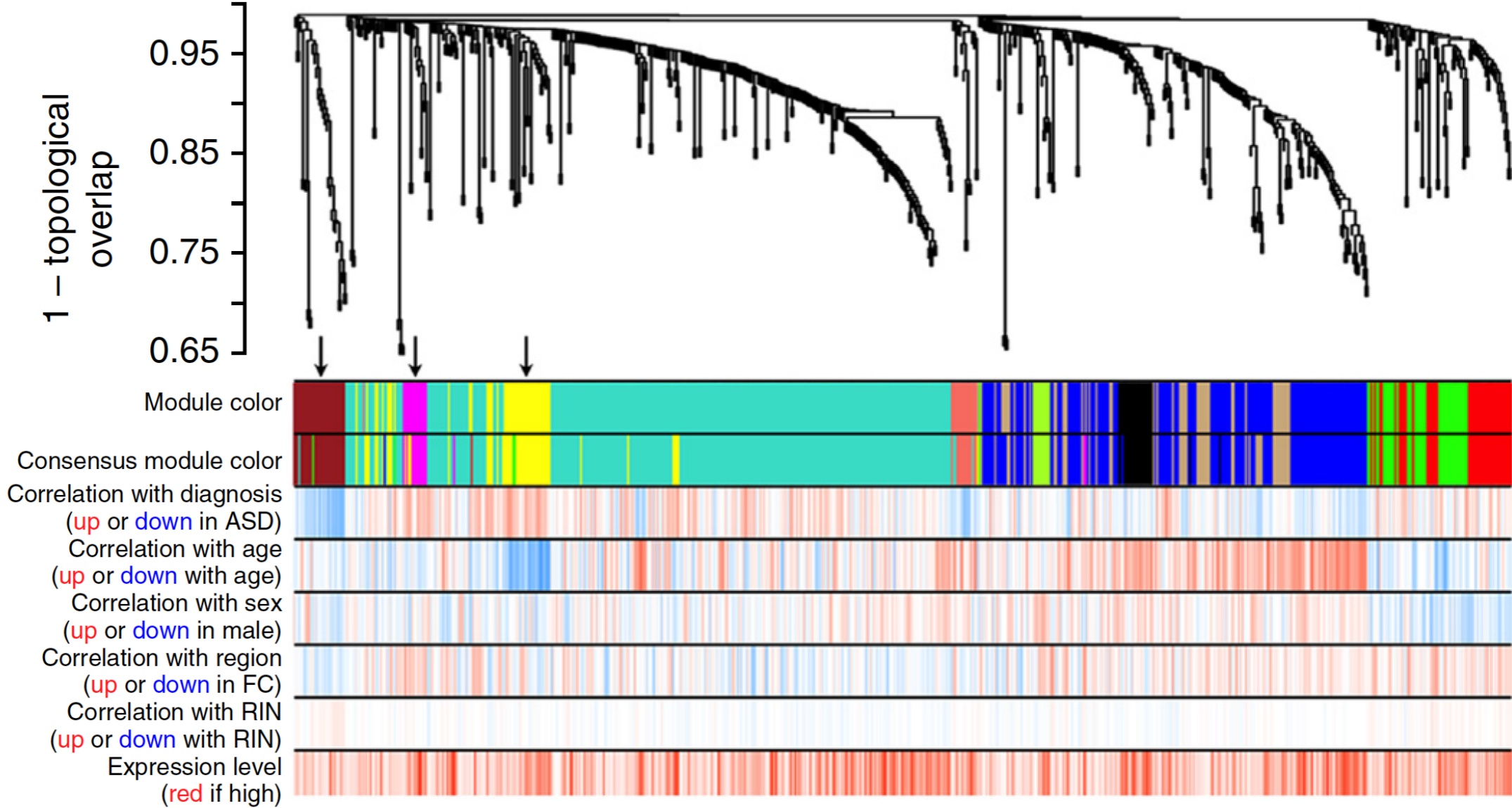
16 are miRNAs differentially expressed (at FDR<5%) in both cortex and cerebellum



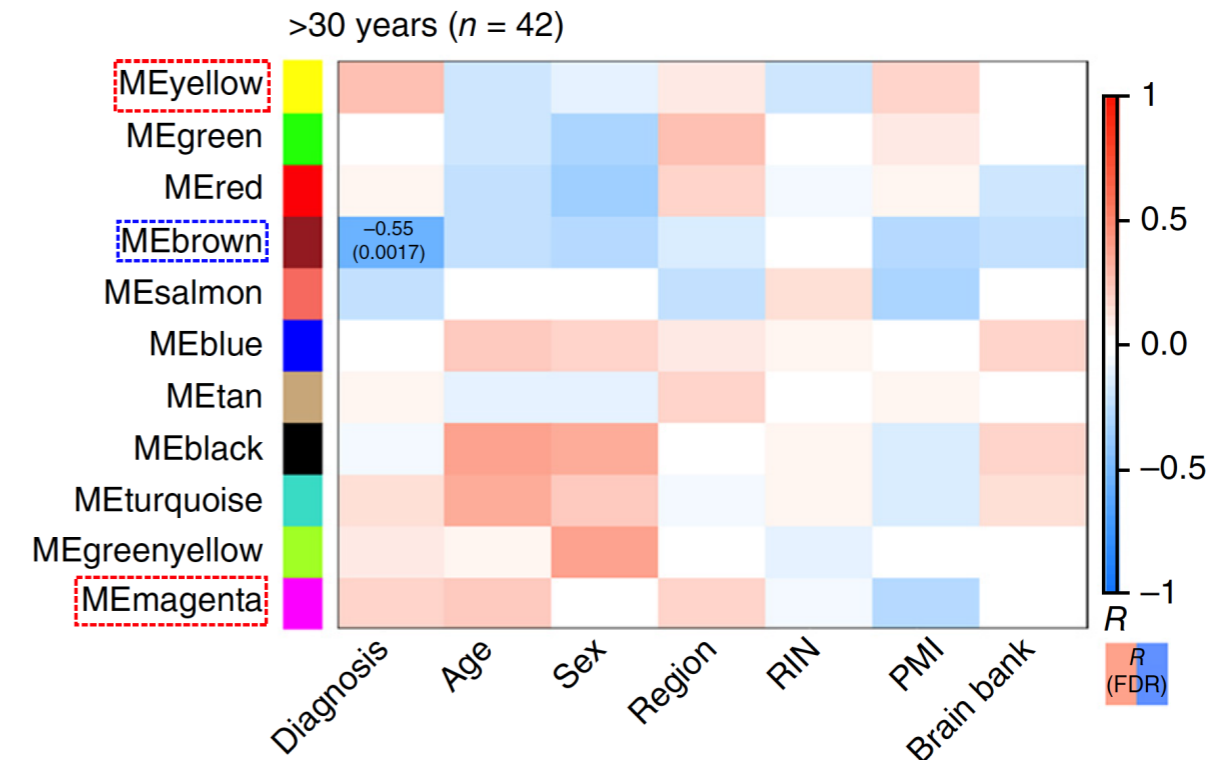
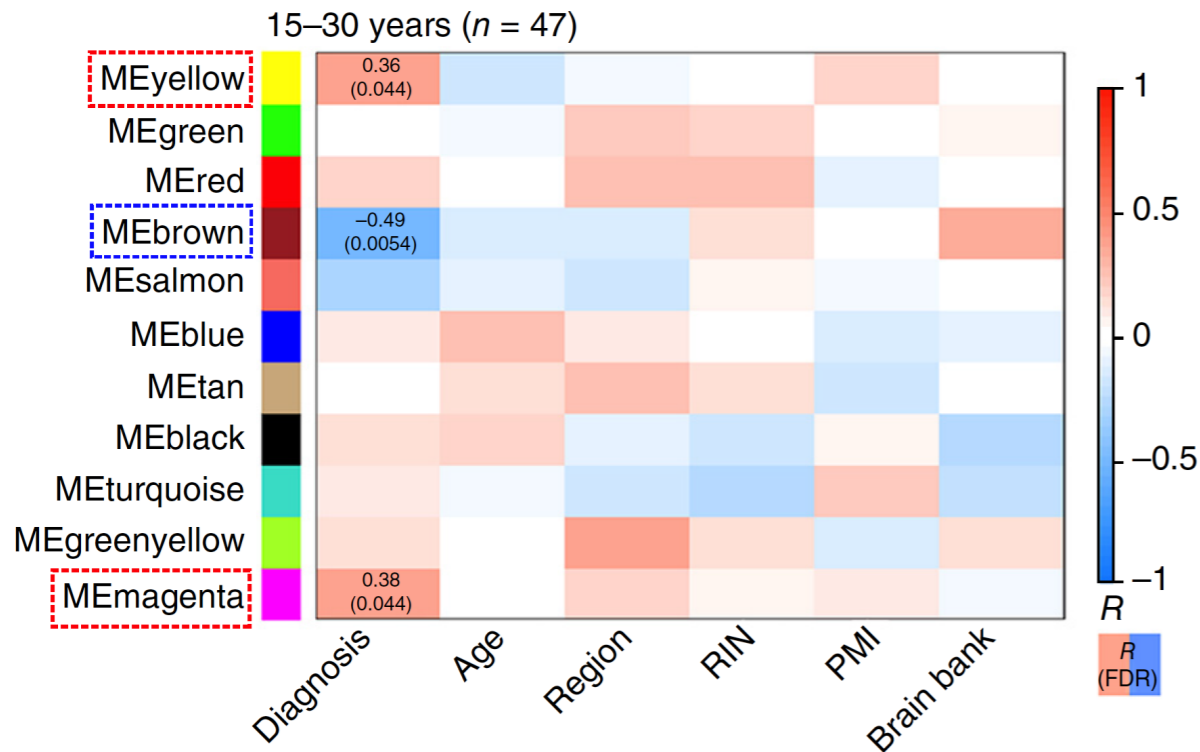
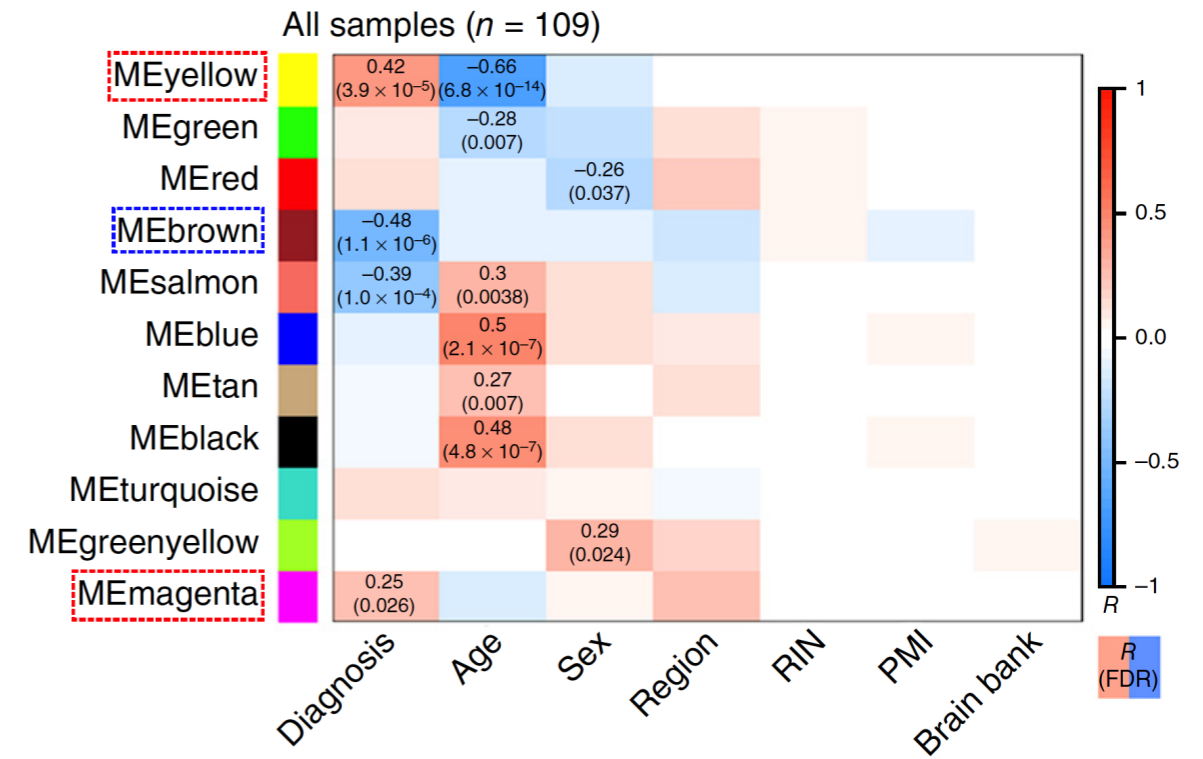
qRT-PCR validation of 10 differentially expressed miRNAs (n=5-8)



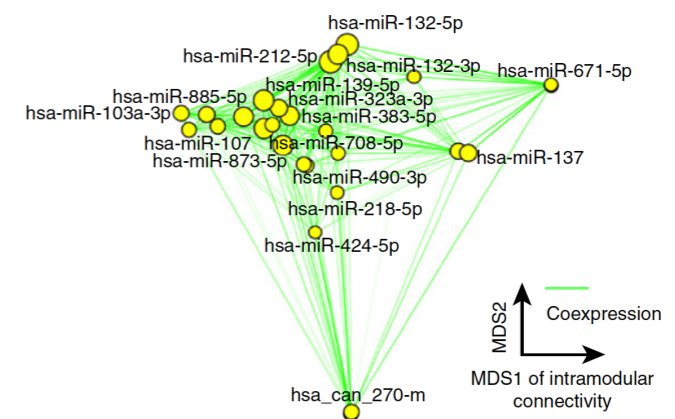
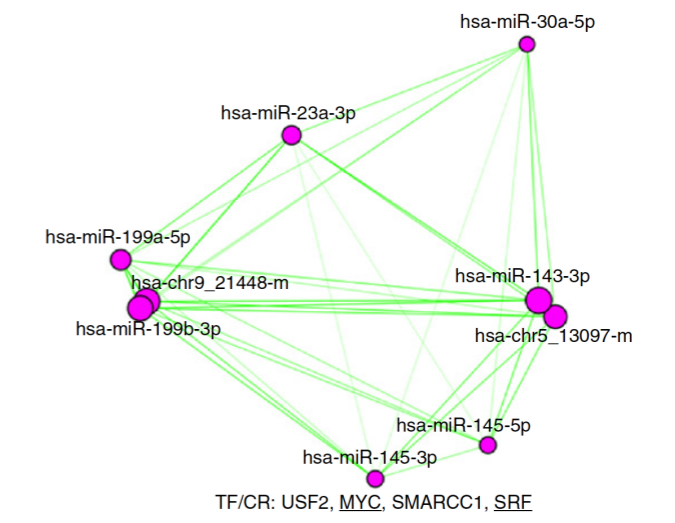
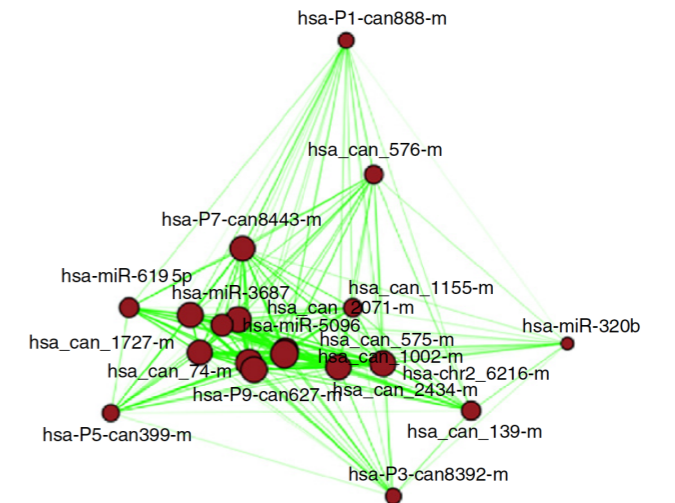
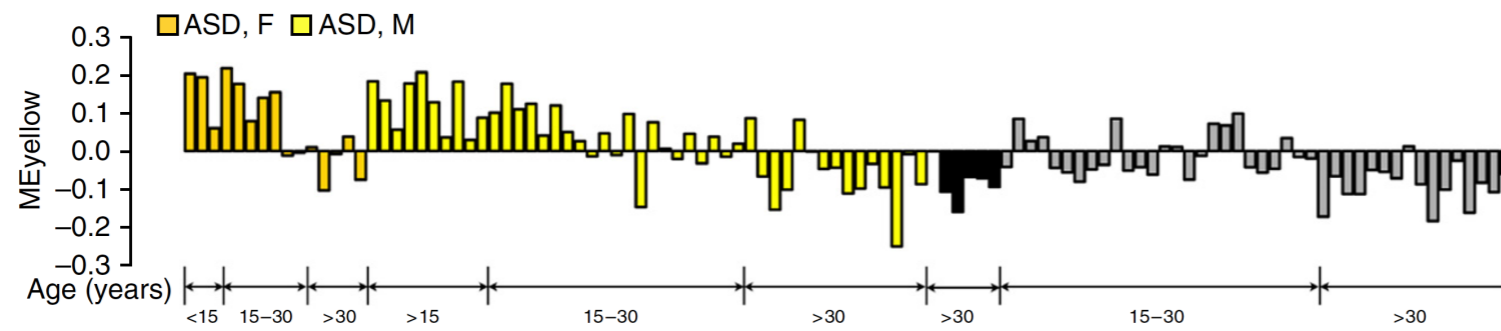
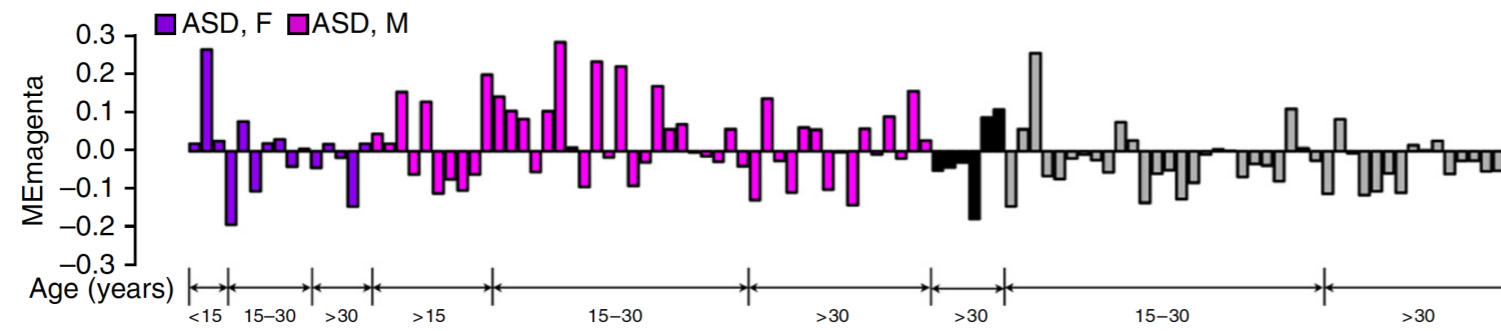
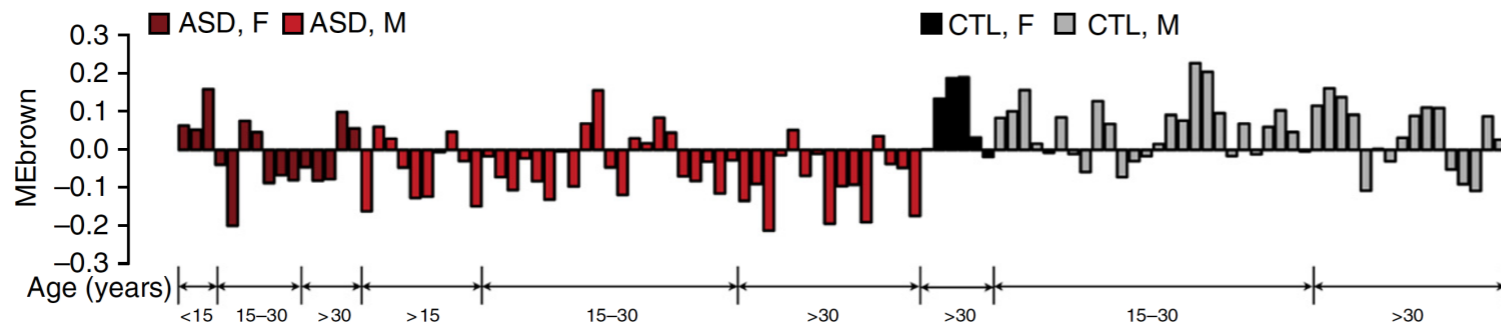
# Weighted Gene Coexpression Network Analysis (WGCNA)



## Pearson correlation coefficients with diagnosis and other potential confounders and covariates



# Scaled module eigengene values and network plots



MDS2 ↑  
MDS1 of intramodular connectivity →

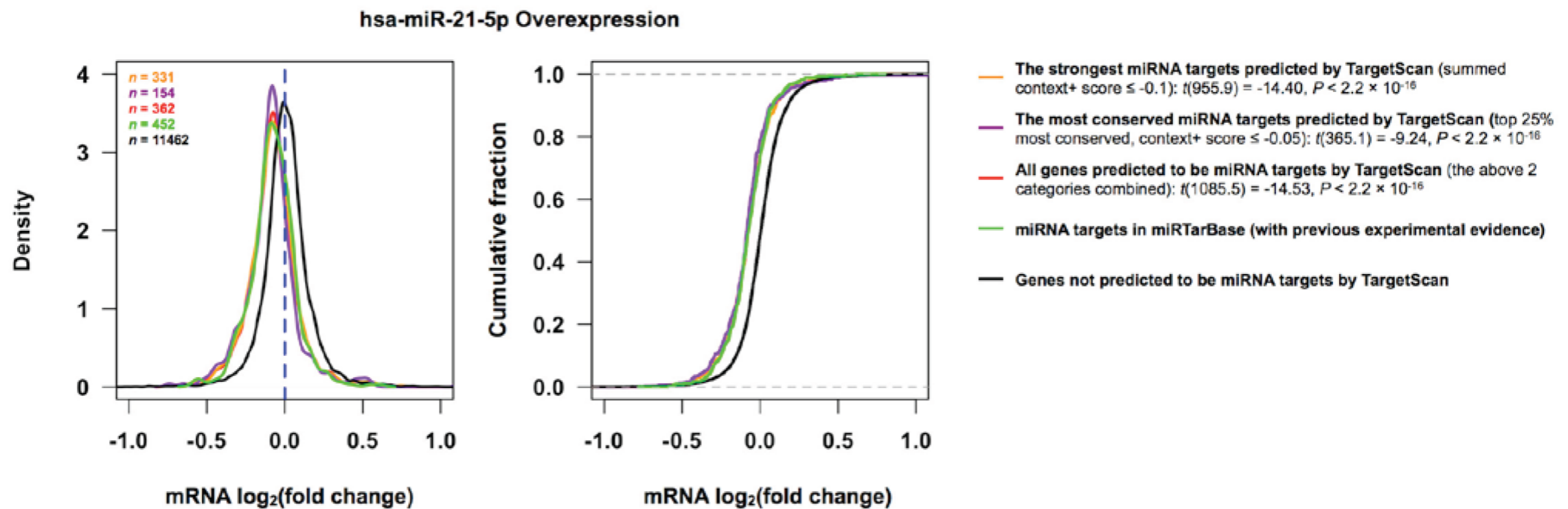
— Coexpression

## mRNA targets of top DE miRNAs and hub miRNAs in ASD-related modules

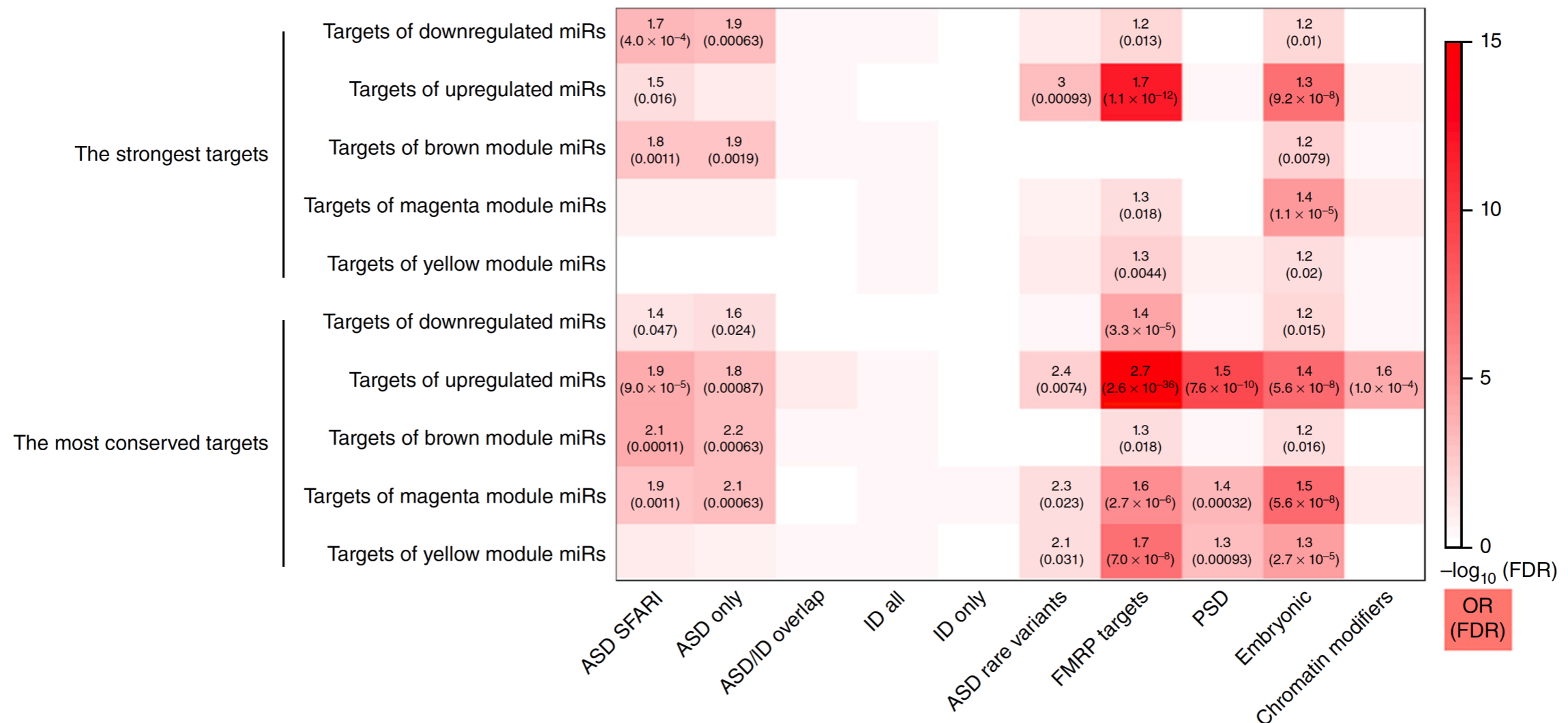
top targets that are expressed in the temporal and frontal cortex:

- targets with the highest predicted targeting efficacy and shared by two or more miRNA
- the most conserved target sites, which are more likely to have conserved physiological roles

experimentally validated hsa-miR-21-5p in human neural progenitor cells (hNPCs):



## Enrichment of ASD risk genes among the top targets of ASD-affected miRNAs and miRNA modules



**ASD SFARI:** set of ASD risk genes from the Simons Foundation Autism Research Initiative (SFARI) AutDB database

**ASD only:** **ASD SFARI** genes not overlapping with **ID all** genes

**ASD/ID overlap:** **ASD SFARI** genes overlapping with **ID all** genes

**ID all:** genes implicated in monogenic form of intellectual disability

**ID only:** **ID all** genes not overlapping with **ASD SFARI** genes

**ASD rare variants:** ASD risk genes implicated by rare variants

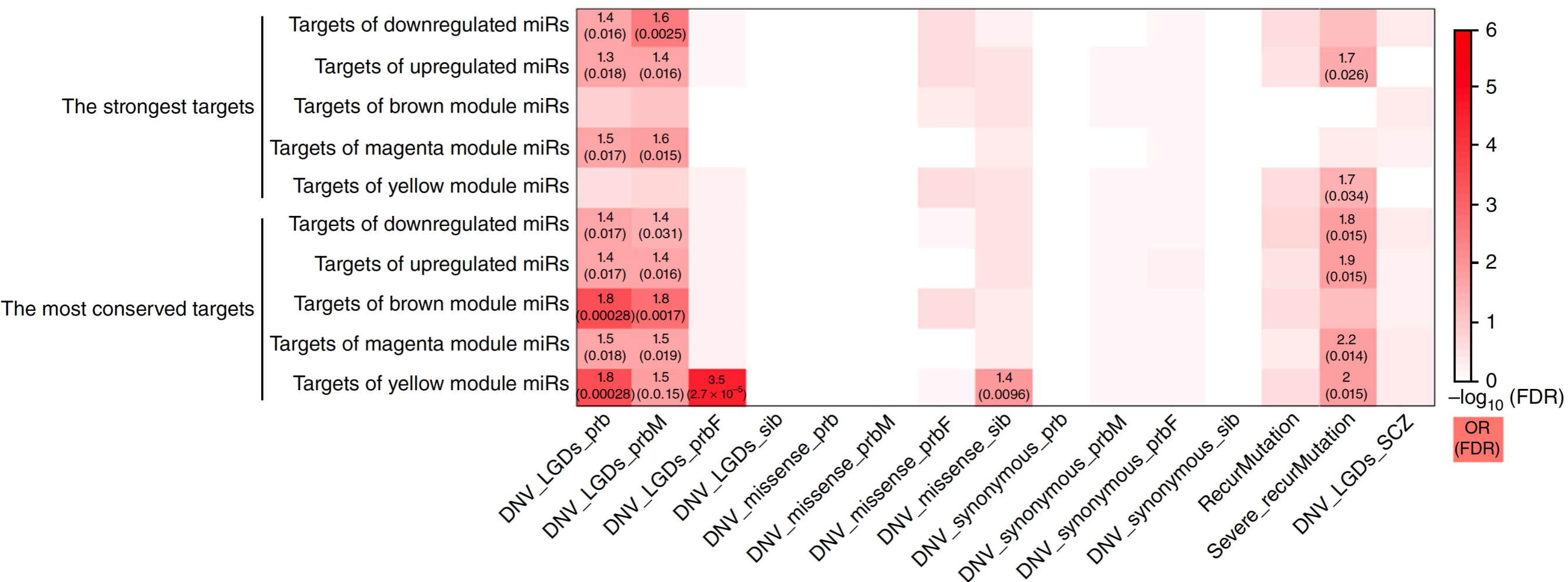
**FMRP targets:** fragile X mental retardation protein transcripts

**PSD:** genes encoding postsynaptic density

**Embryonic:** genes expressed preferentially during embryonic brain development

**Chromatin modifiers:** genes encoding chromatin modifiers

## Enrichment of genes affected by *de novo* ASD variants



**DNV:** *de novo* variants in ASD

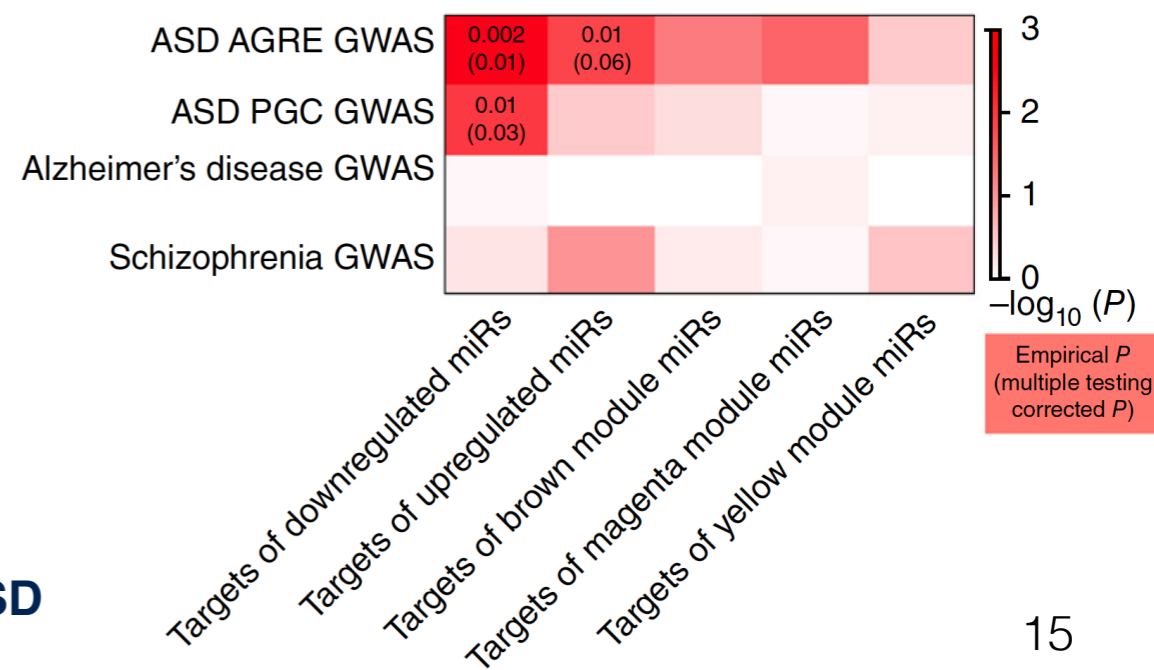
**LGD:** 'likely-gene-disrupting' (including nonsense, splice-site and frame shift)

**prb, prbM, prbF:** probants: all, male, female

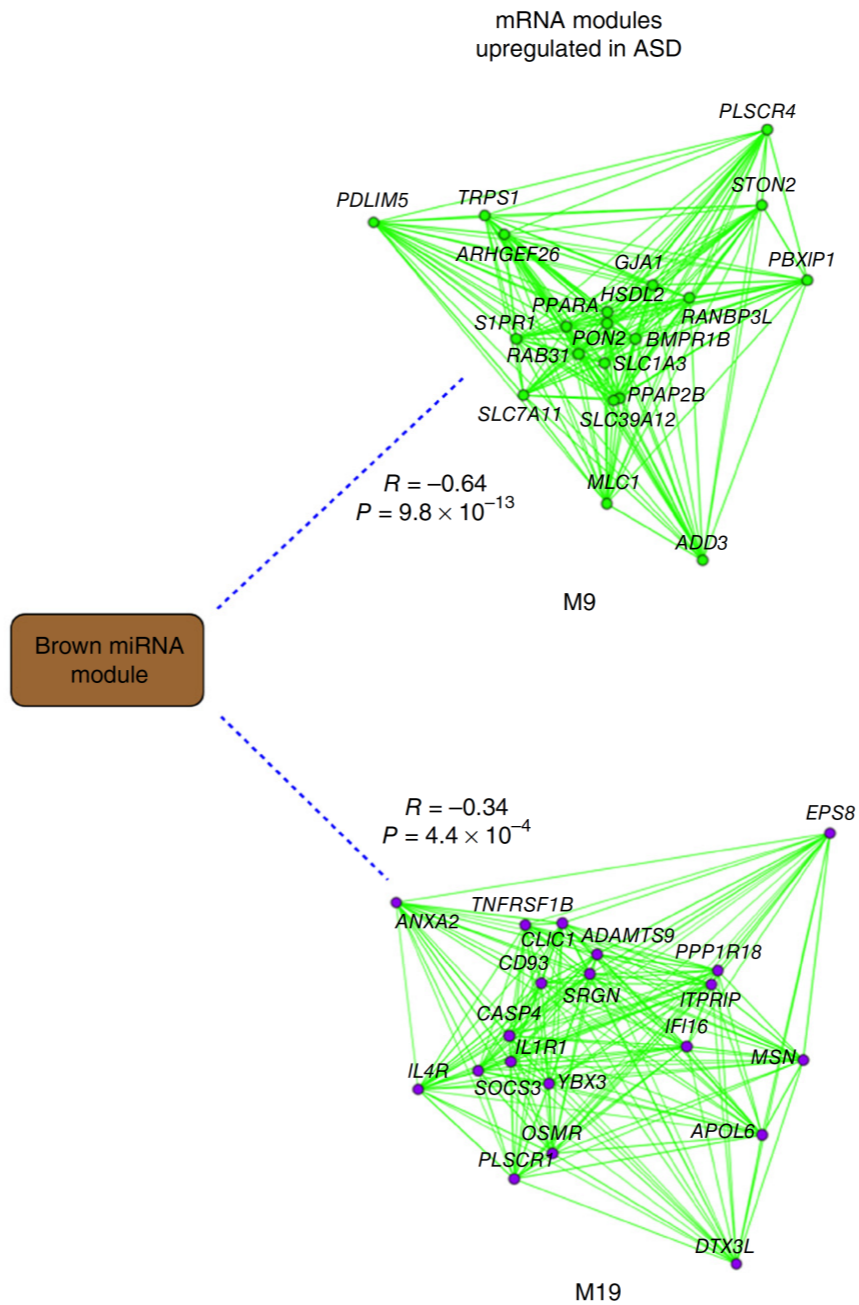
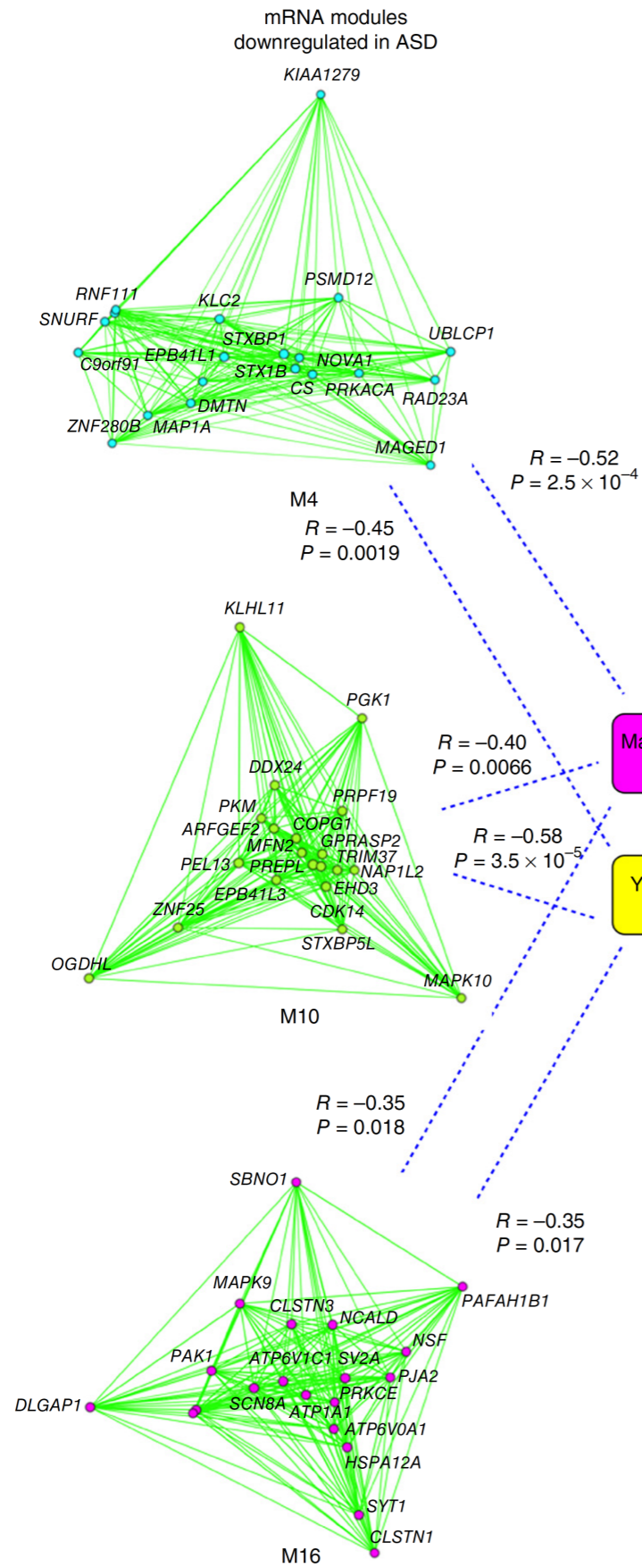
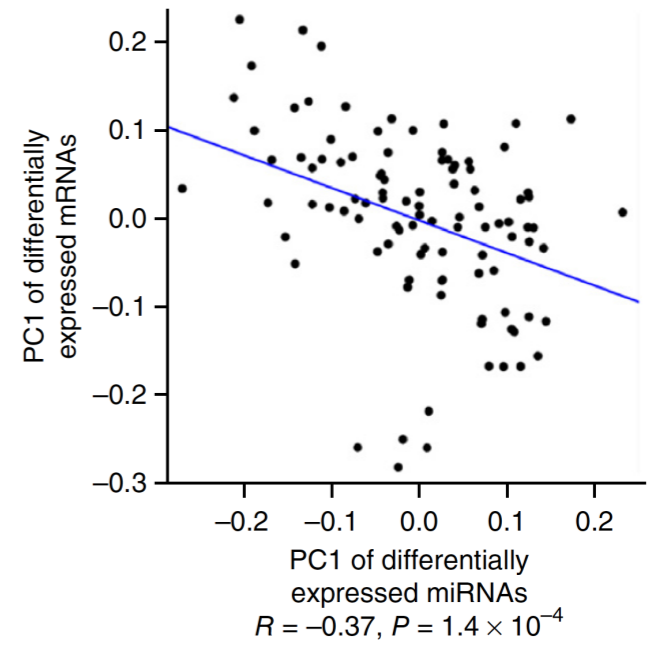
**sib:** siblings

**SCZ:** individuals with schizophrenia

## Enrichment of overlap with LD-based independent genomic regions associated with ASD

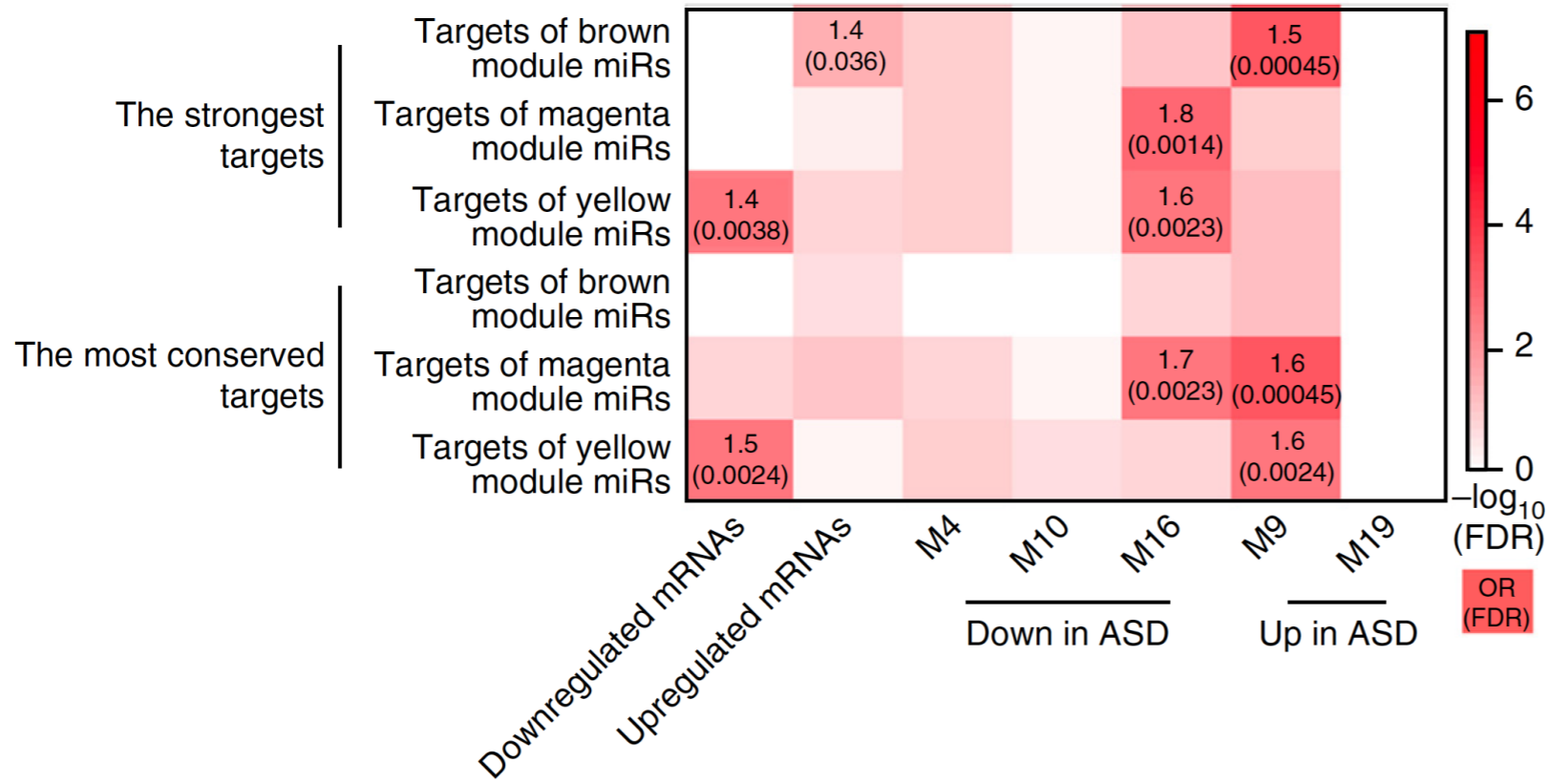


# Relationship b/w miRNA and mRNA (101 matching samples in 'unpublished observations') expression changes



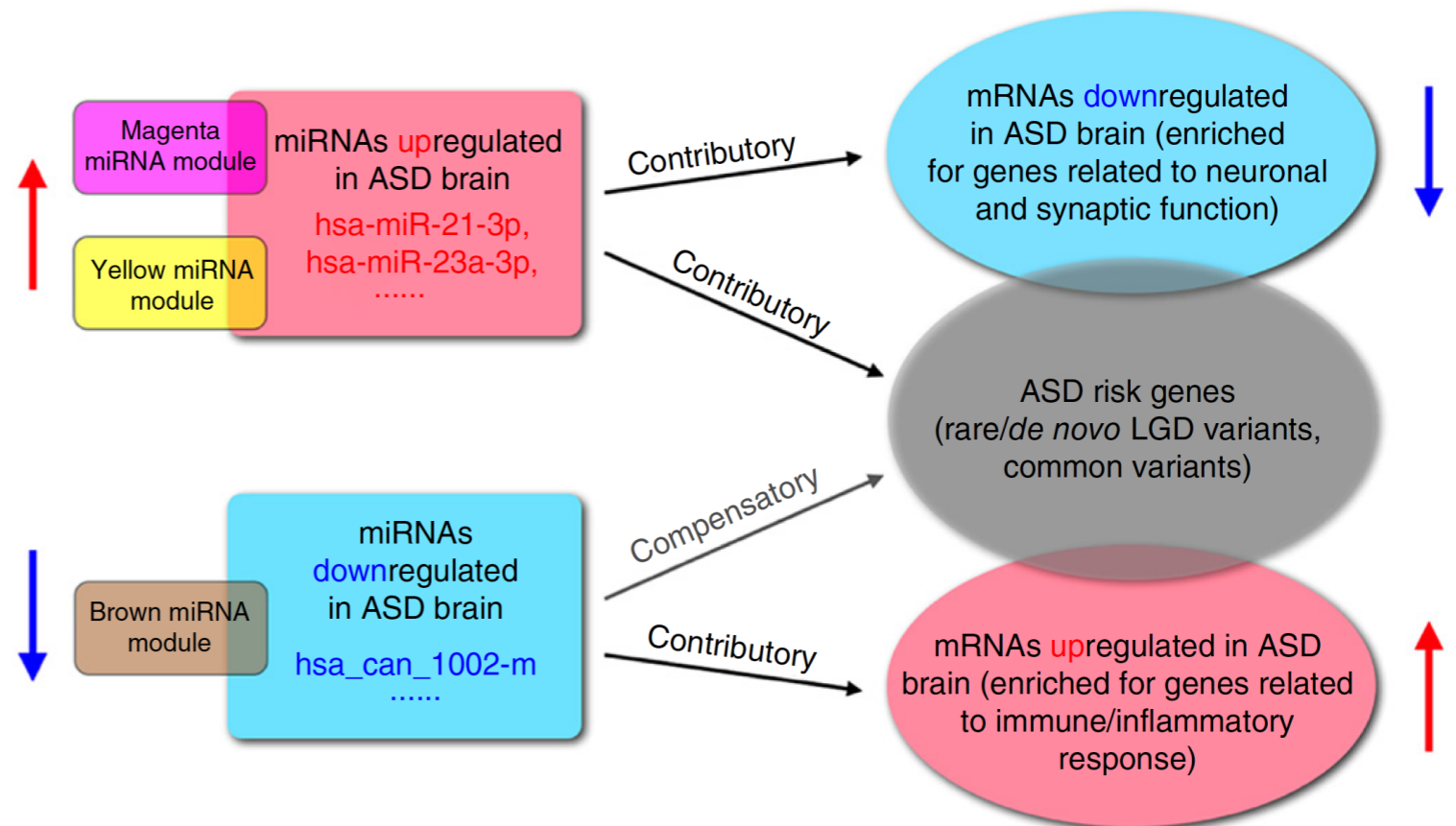


# Enrichment for ASD-affected mRNAs and mRNA modules in the top targets of ASD-affected mRNAs

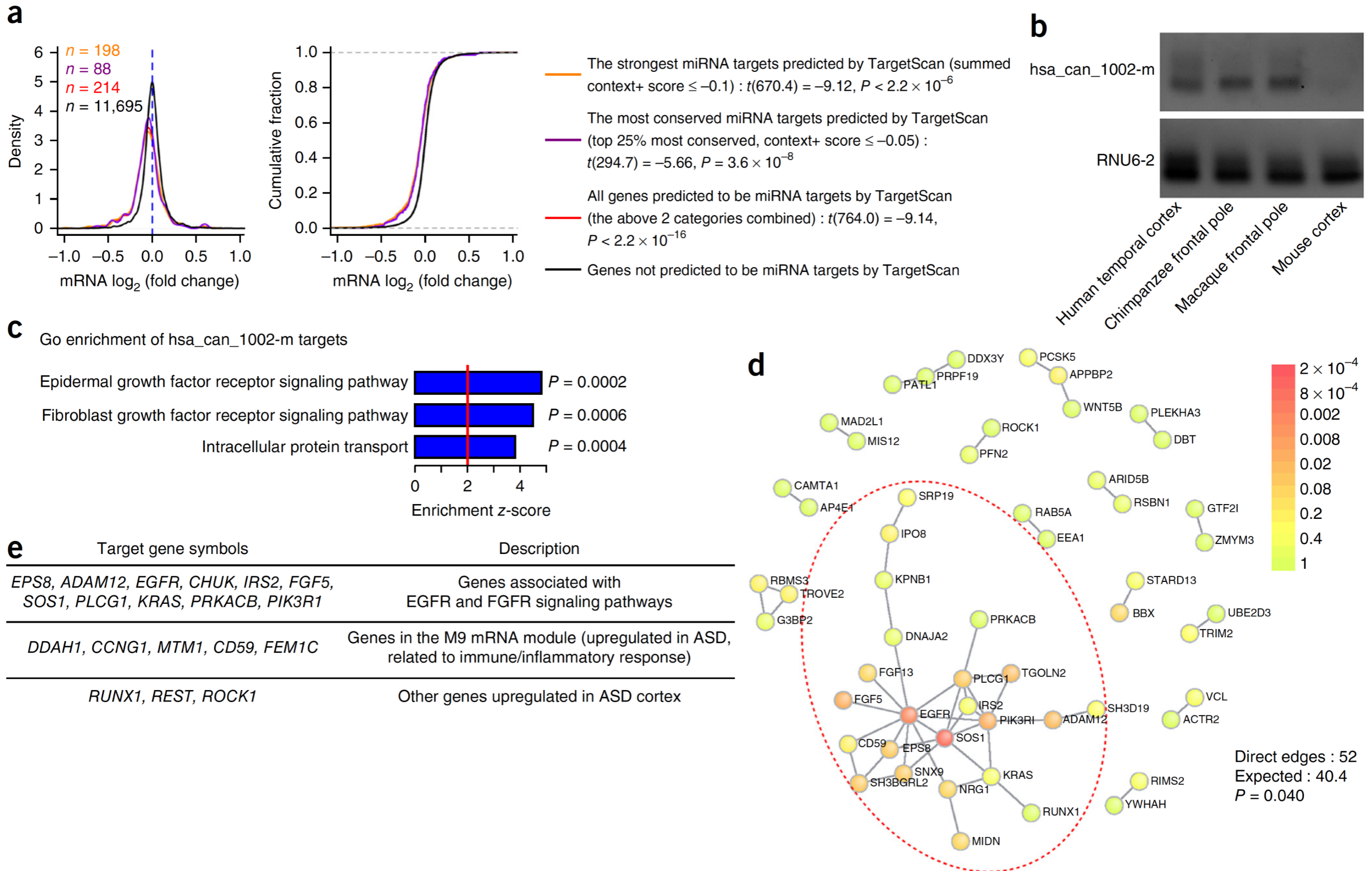


“The upregulated miRNAs and miRNA modules may have a contributory role by repressing ASD risk genes and neuronal/synaptic genes downregulated in ASD.

The downregulated miRNAs and miRNA module may contribute to the upregulation of immune/inflammatory genes in ASD, but might also have a compensatory role given the enrichment of their targets for rare protein-disrupting and common genetic variants associated with ASD”



# Experimental validation of hsa\_can\_1002-m



# Genome-wide changes in lncRNA, splicing, and regional gene expression patterns in autism

Neelroop N. Parikshak<sup>1,2\*</sup>, Vivek Swarup<sup>1,2\*</sup>, T. Grant Belgard<sup>1,2\*†</sup>, Manuel Irimia<sup>3,4</sup>, Gokul Ramaswami<sup>1,2</sup>, Michael J. Gandal<sup>1,2</sup>, Christopher Hartl<sup>1,2</sup>, Virpi Leppä<sup>1</sup>, Luis de la Torre Ubieta<sup>1,2</sup>, Jerry Huang<sup>1,2</sup>, Jennifer K. Lowe<sup>1</sup>, Benjamin J. Blencowe<sup>5,6</sup>, Steve Horvath<sup>7,8</sup> & Daniel H. Geschwind<sup>1,2,7</sup>

**Autism spectrum disorder (ASD) involves substantial genetic contributions. These contributions are profoundly heterogeneous but may converge on common pathways that are not yet well understood<sup>1–3</sup>. Here, through post-mortem genome-wide transcriptome analysis of the largest cohort of samples analysed so far, to our knowledge<sup>4–7</sup>, we interrogate the noncoding transcriptome, alternative splicing, and upstream molecular regulators to broaden our understanding of molecular convergence in ASD. Our analysis reveals ASD-associated dysregulation of primate-specific long noncoding RNAs (lncRNAs), downregulation of the alternative splicing of activity-dependent neuron-specific exons, and attenuation of normal differences in gene expression between the frontal and temporal lobes. Our data suggest that SOX5, a transcription factor involved in neuron fate specification, contributes to this reduction in regional differences. We further demonstrate that a genetically defined subtype of ASD, chromosome 15q11.2-13.1 duplication syndrome (dup15q), shares the core transcriptomic signature observed in idiopathic ASD. Co-expression network analysis reveals that individuals with ASD show age-related changes in the trajectory of microglial and synaptic function over the first two decades, and suggests that genetic risk for ASD may influence changes in regional cortical gene expression. Our findings illustrate how diverse genetic perturbations can lead to phenotypic convergence at multiple biological levels in a complex neuropsychiatric disorder.**

status (Extended Data Fig. 2a) and confirmed the technical quality of our data with qRT-PCR (Extended Data Fig. 2b, c). We next evaluated enrichment of the gene sets for pathways and cell types (Extended Data Fig. 2d, e), and found that the downregulated set was enriched in genes expressed in neurons and involved in neuronal pathways, including *PVALB* and *SYT2*, which are highly expressed in interneurons; by contrast, the upregulated gene set was enriched in genes expressed in microglia and astrocytes<sup>8</sup>.

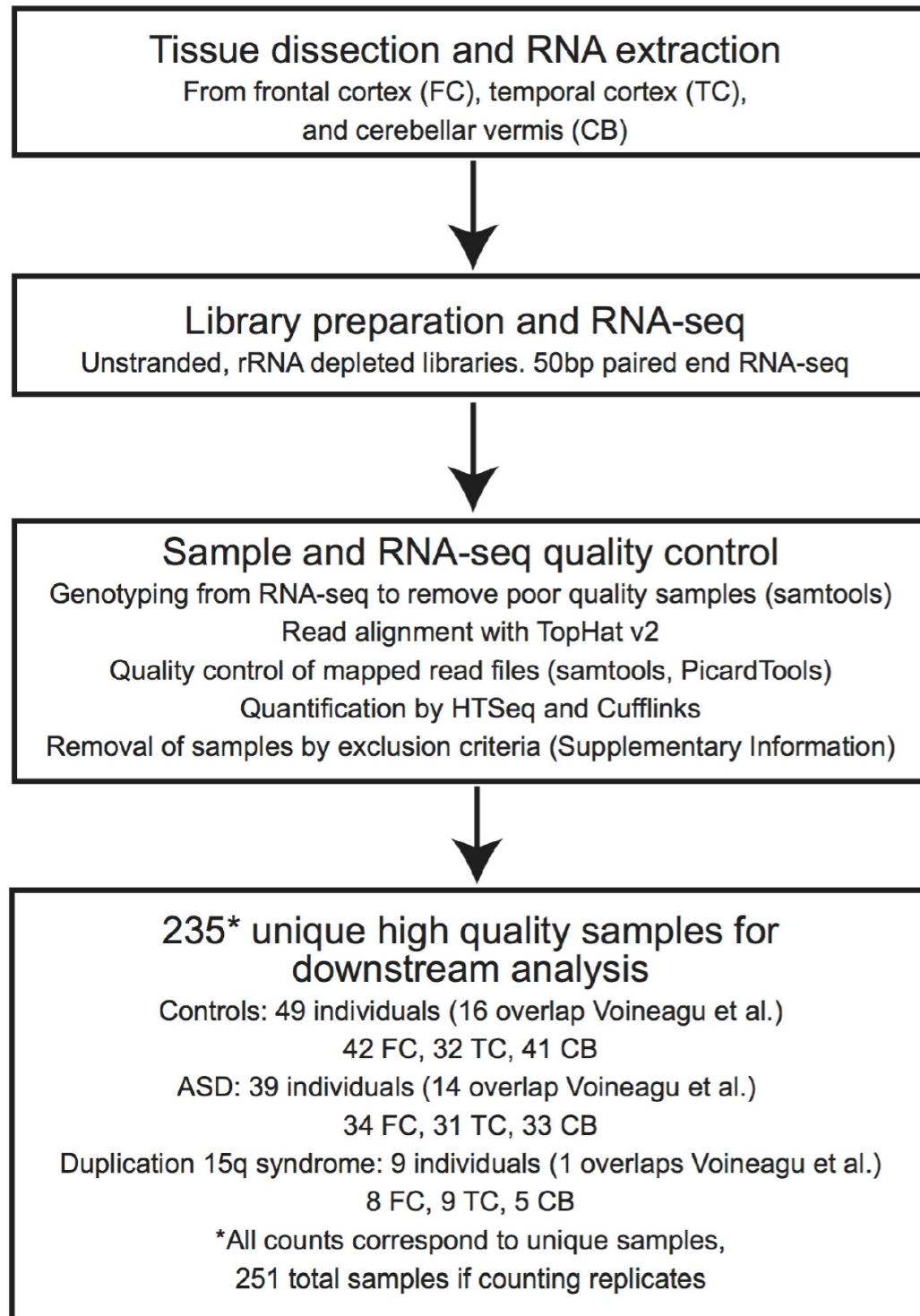
Although there was no significant DGE in the cerebellum (FDR < 0.05, *P* distributions in Fig. 1b), similar to observations in a smaller cohort<sup>8</sup>, there was a replication signal in the cerebellum and overall concordance between ASD-related fold changes in the cortex and cerebellum (Extended Data Fig. 2f–h). The lack of significant DGE in the cerebellum is explained by the fact that changes in expression were consistently stronger in the cortex than in the cerebellum (Extended Data Fig. 2h), which suggests that the cortex is more selectively vulnerable to these transcriptomic alterations. We also compared our results to an RNA-seq study of protein coding genes in the occipital cortex of individuals with ASD and control subjects<sup>4</sup>. Despite significant technical differences that reduce power to detect DGE, and profiling of different brain regions in that study, there was a weak but significant correlation in fold changes, which was due mostly to upregulated genes in both studies (*P* = 0.038, Extended Data Fig. 2i, j).

We next explored lncRNAs, most of which have little functional annotation, and identified 60 lncRNAs in the DGE set (FDR < 0.05,

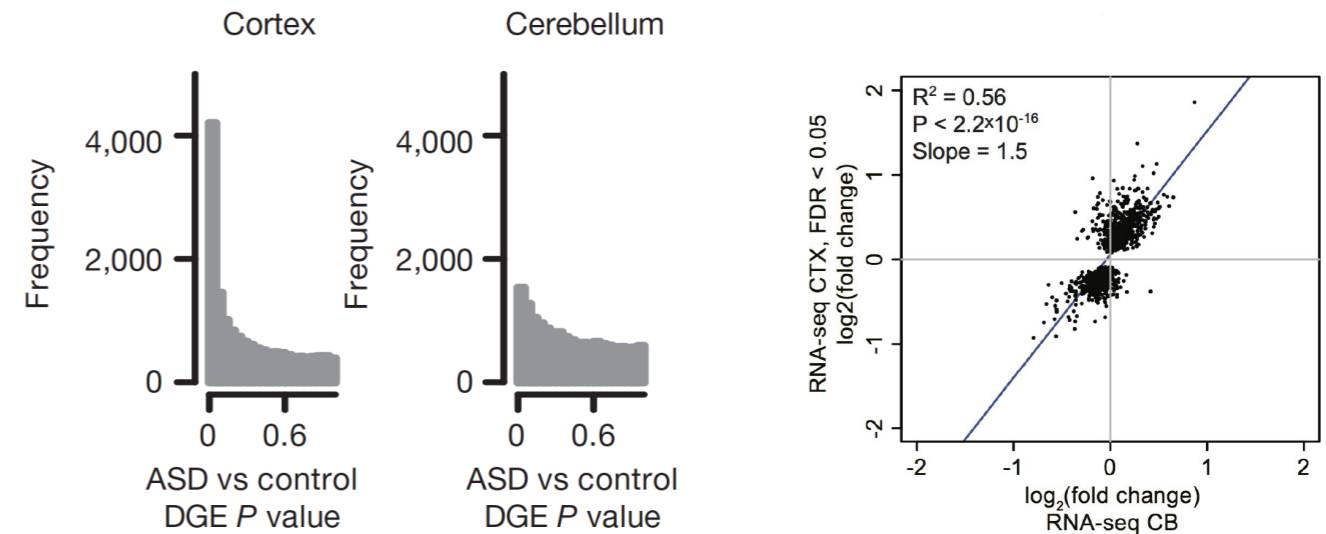
<sup>1</sup>Center for Autism Research and Treatment and Program in Neurobehavioral Genetics, Semel Institute, David Geffen School of Medicine, University of California, Los Angeles, Los Angeles, California 90095, USA. <sup>2</sup>Department of Neurology, David Geffen School of Medicine, University of California Los Angeles, 695 Charles E. Young Drive South, Los Angeles, California 90095, USA. <sup>3</sup>Centre for Genomic Regulation, Barcelona Institute of Science and Technology (BIST), 88 Dr. Aiguader, Barcelona 08003, Spain. <sup>4</sup>Universitat Pompeu Fabra (UPF), Barcelona, Spain. <sup>5</sup>Donnelly Centre, University of Toronto, 160 College Street, Toronto, ON M5S 3E1, Canada. <sup>6</sup>Department of Molecular Genetics, University of Toronto, 1 King's College Circle, Toronto, ON M5S 1A8, Canada. <sup>7</sup>Department of Human Genetics, David Geffen School of Medicine, University of California, Los Angeles, California, USA. <sup>8</sup>Department of Biostatistics, David Geffen School of Medicine, University of California, Los Angeles, California, USA. †Present address: Verge Genomics, 42A Dore Street, San Francisco, California 94103, USA.

\*These authors contributed equally to this work.

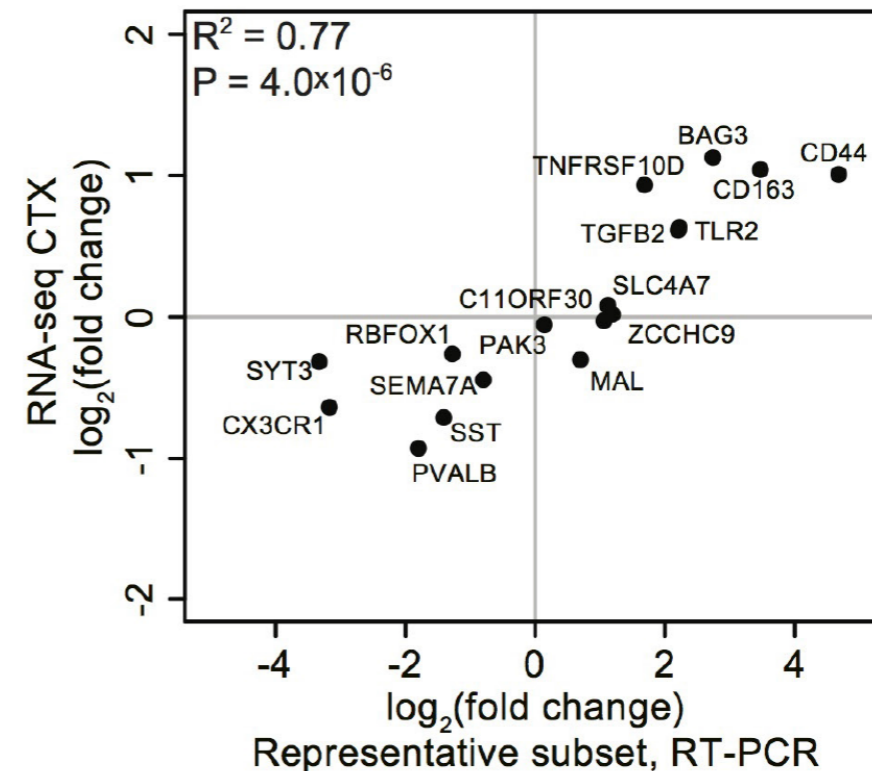
## RNA-seq workflow



## DE for entire transcriptome and qRT-PCR validation

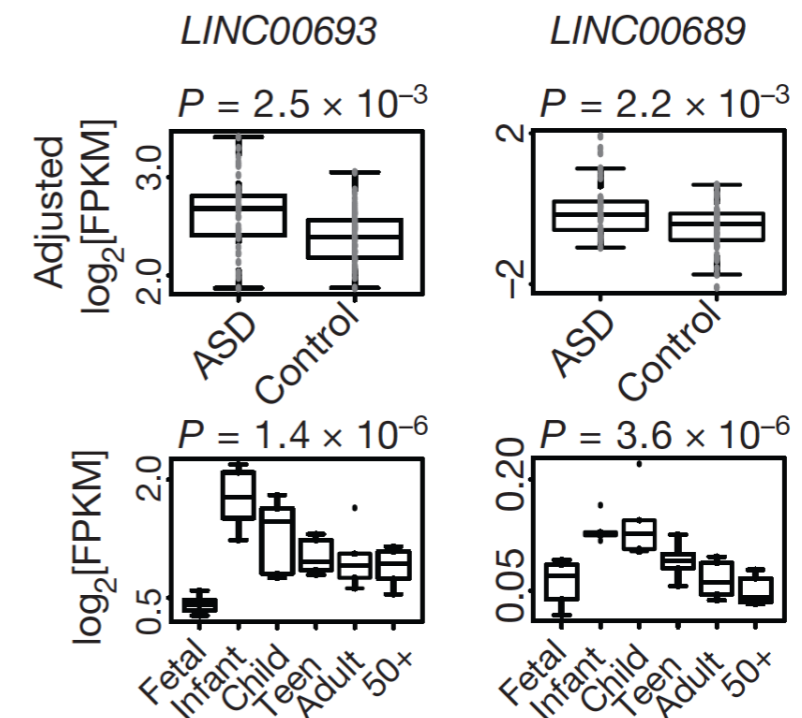
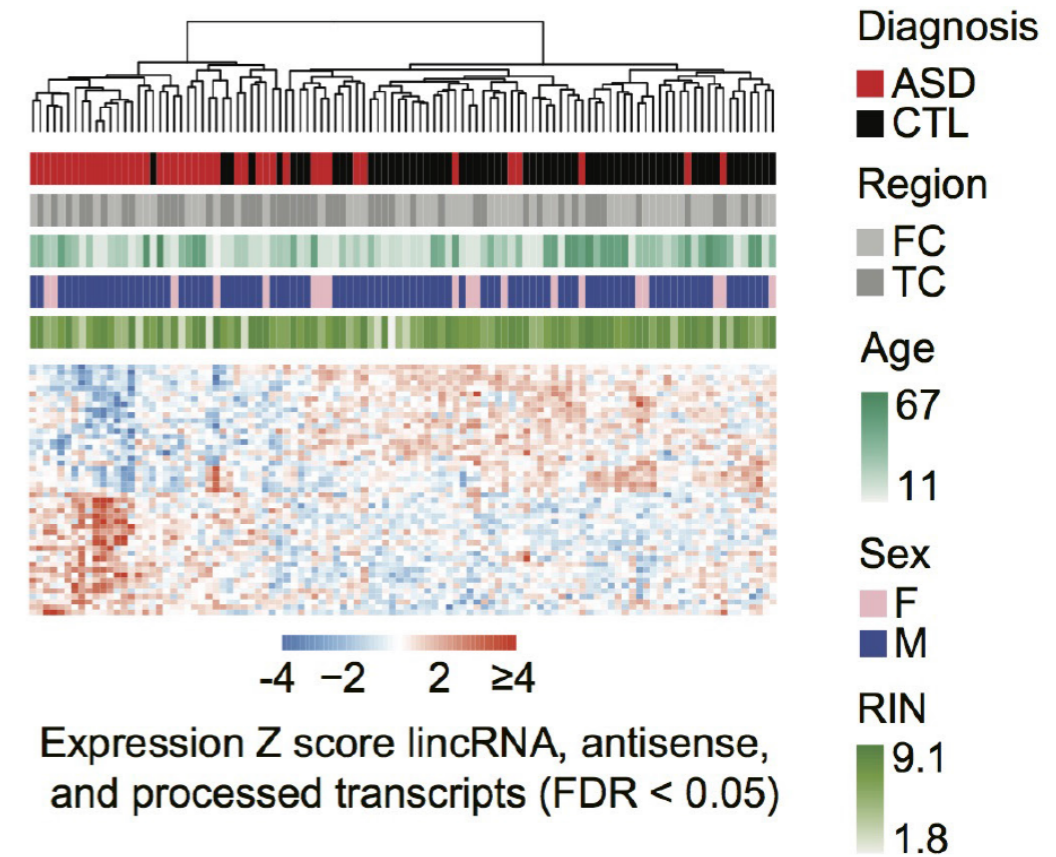
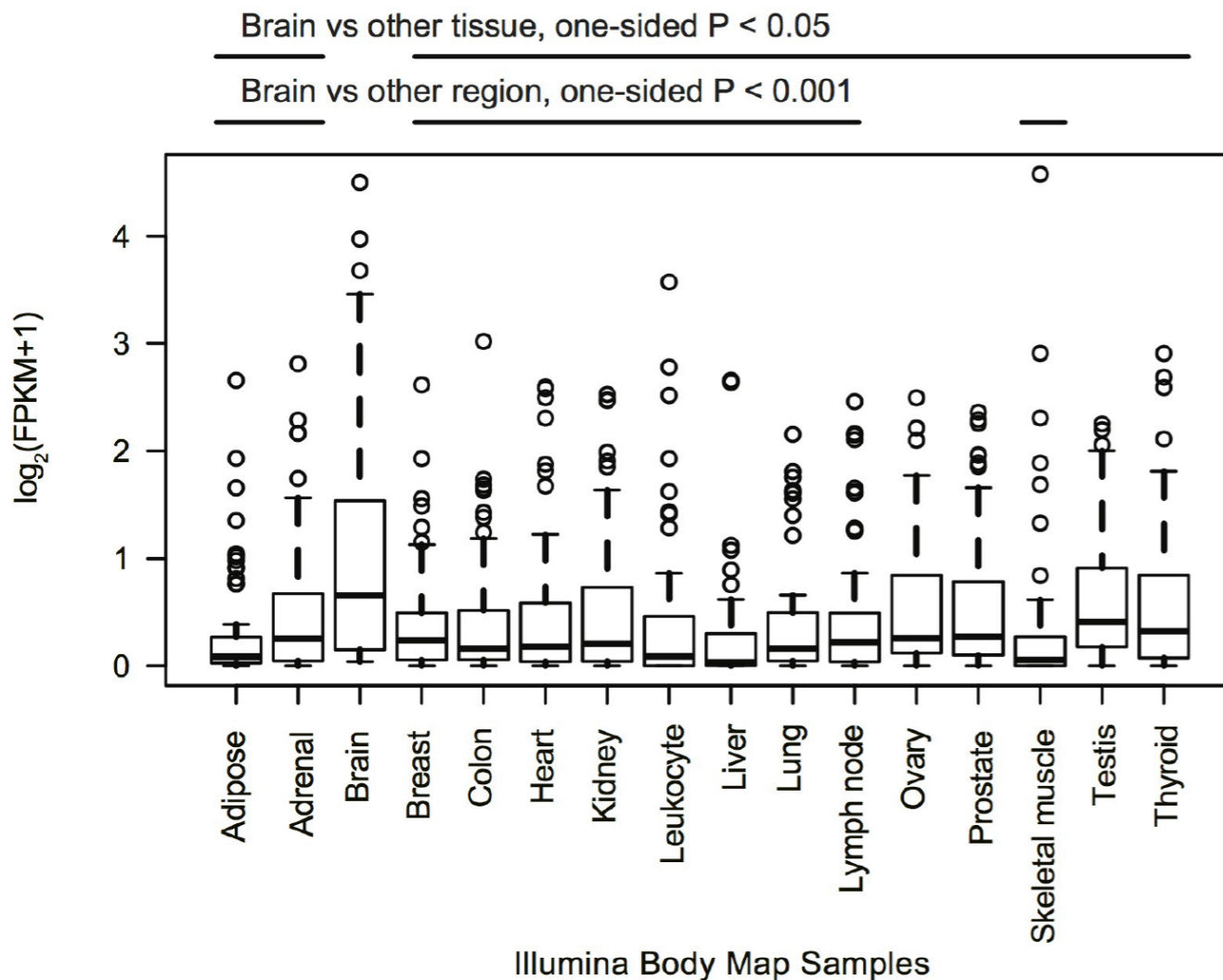


**584** genes showed increased and **558** - decreased expression in idiopathic ASD cortex



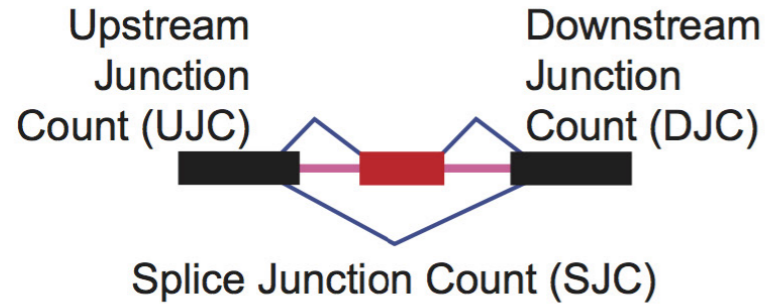
## 60 lincRNA in the DGE set: hierarchical clustering and enrichment in brain relative to other tissues

most exhibit primate-specific expression patterns in brain  
 20 interact with miRNA-protein complexes  
 9 - with FMRP  
 thus, predicted to affect protein expression through miRNA and FMRP interactions



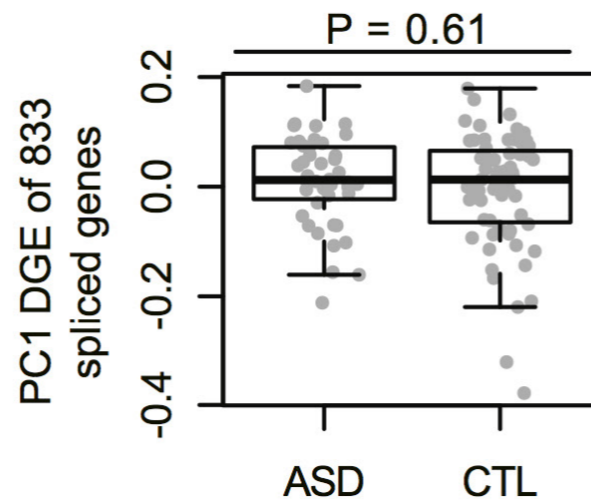
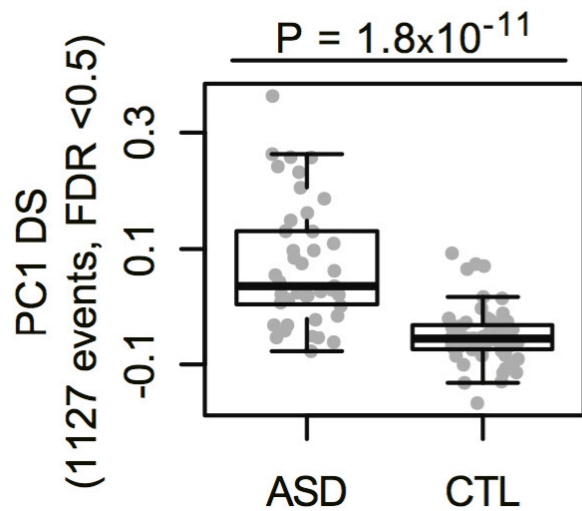
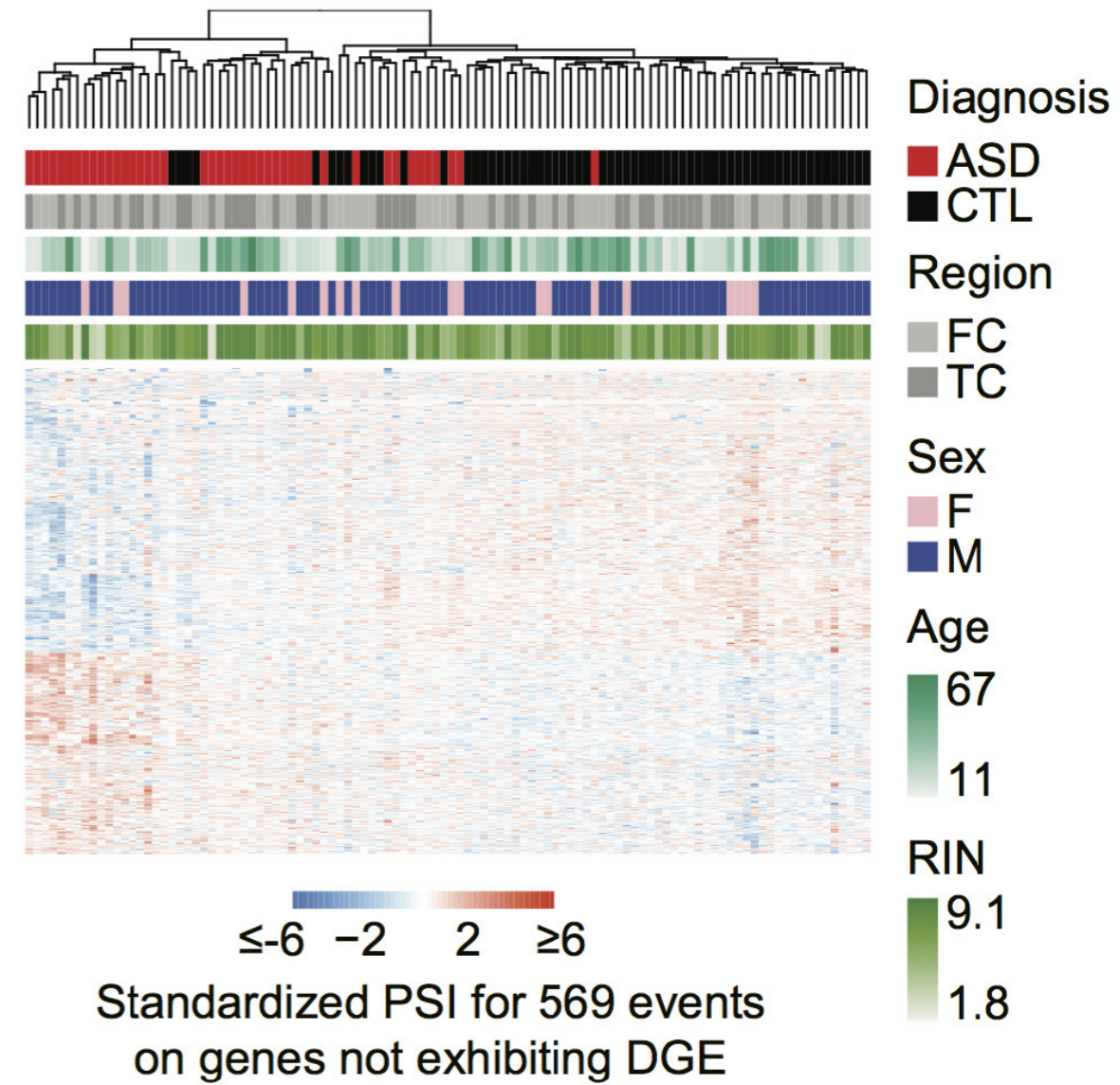
# Splicing differential analysis and validation

Calculation of Percent Spliced In (PSI)



$$PSI (\psi) = \frac{(UJC + DJC)/2}{(UJC + DJC)/2 + SJC} \times 100$$

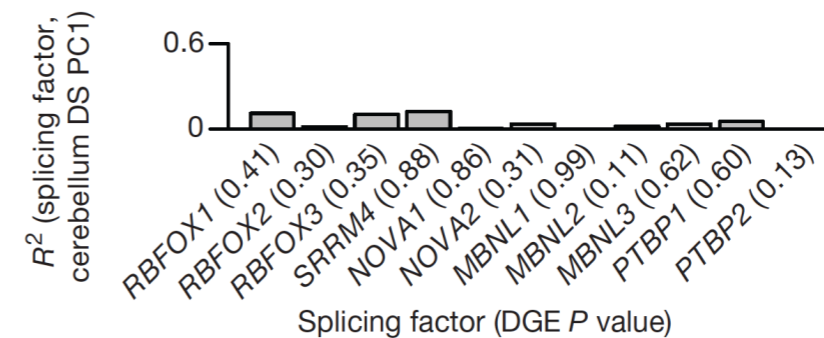
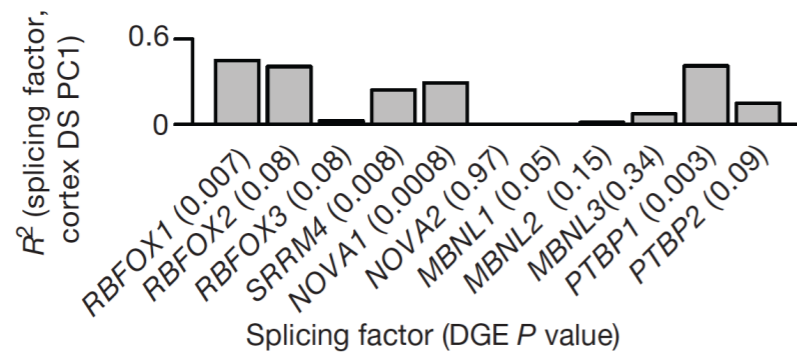
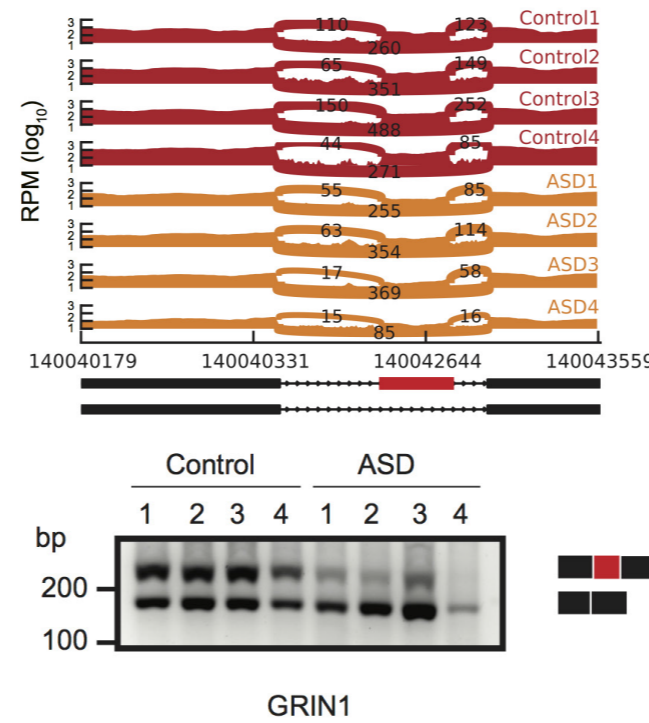
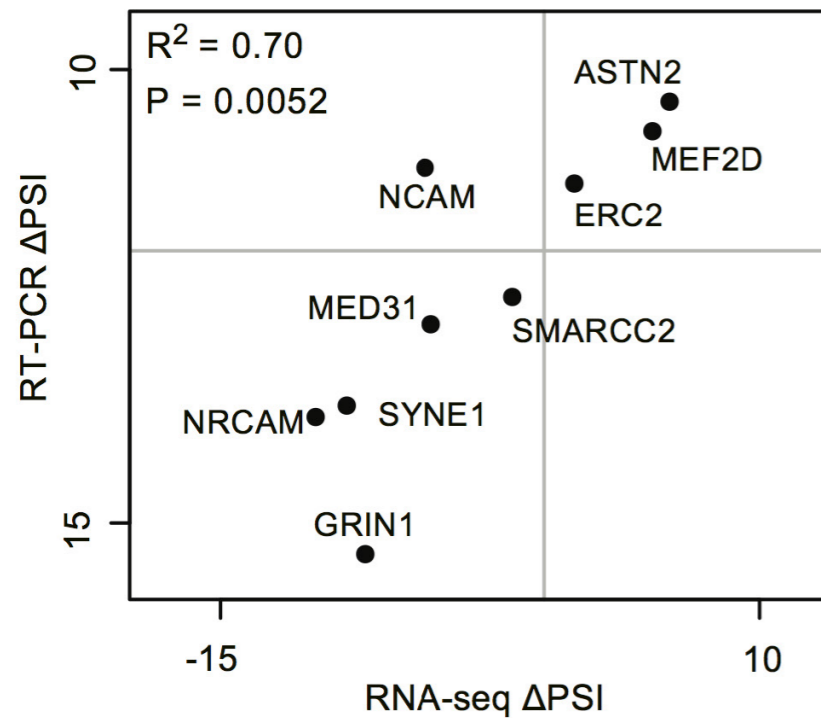
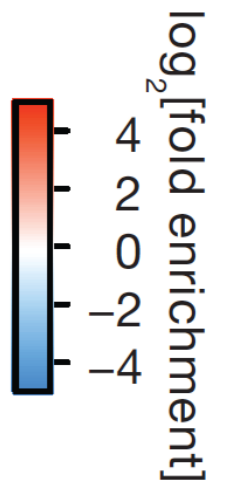
**1127** differential splicing events in **833** genes in cortex



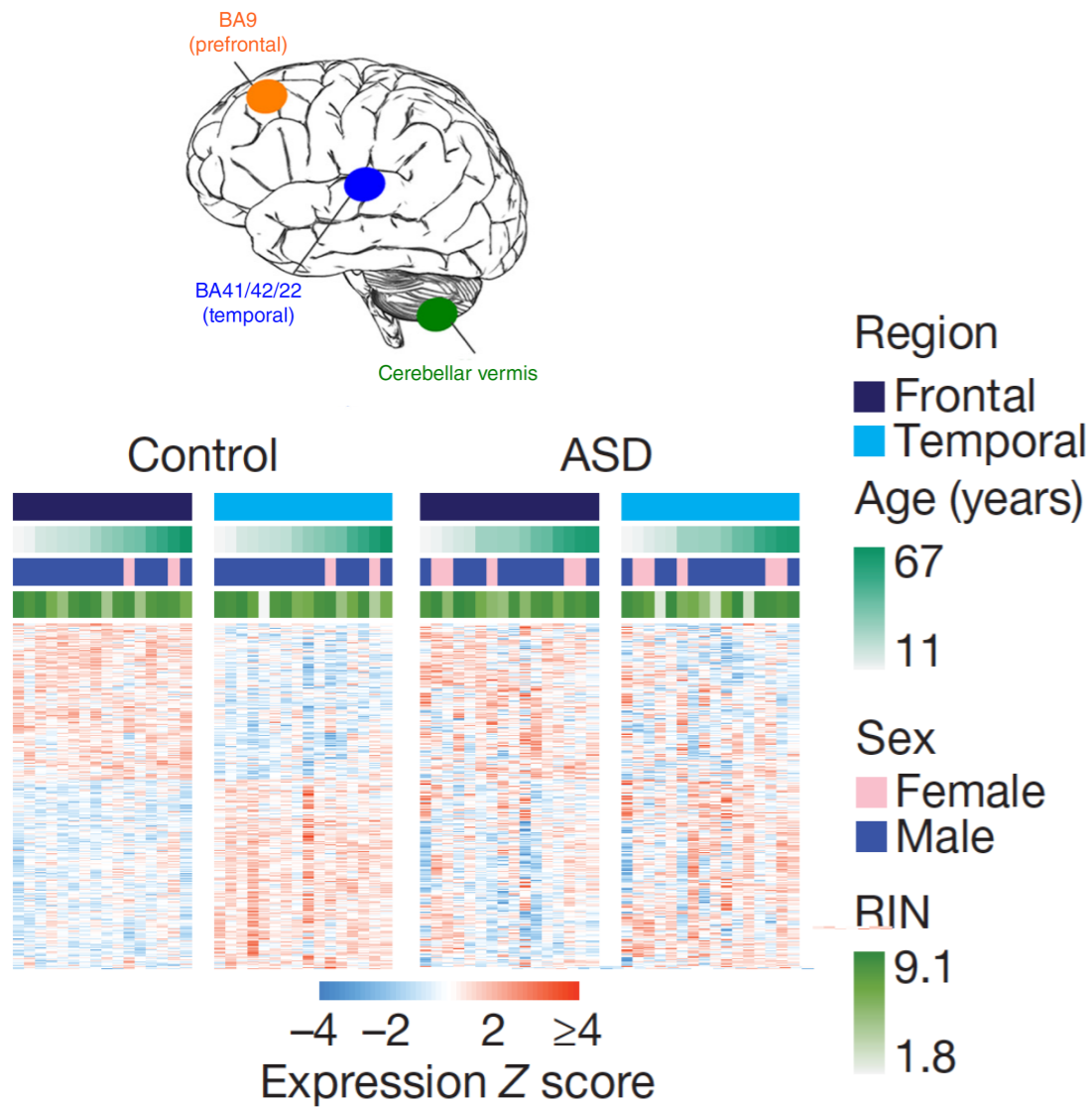
# Splicing differential analysis and validation

Fold enrichment  
(*P* value)

All events	2.9 ( $2.8 \times 10^{-5}$ )	1.4 (0.48)	2 (0.14)	1.1 (0.8)
Inclusion in ASD	0.33 (0.37)	3.2 (0.14)	1 (1)	1.5 (0.39)
Exclusion in ASD	4.1 ( $1.8 \times 10^{-7}$ )	0.65 (1)	2.4 (0.072)	0.97 (1)
	Neurons	Astrocytes	Oligo- dendrocytes	Microglia

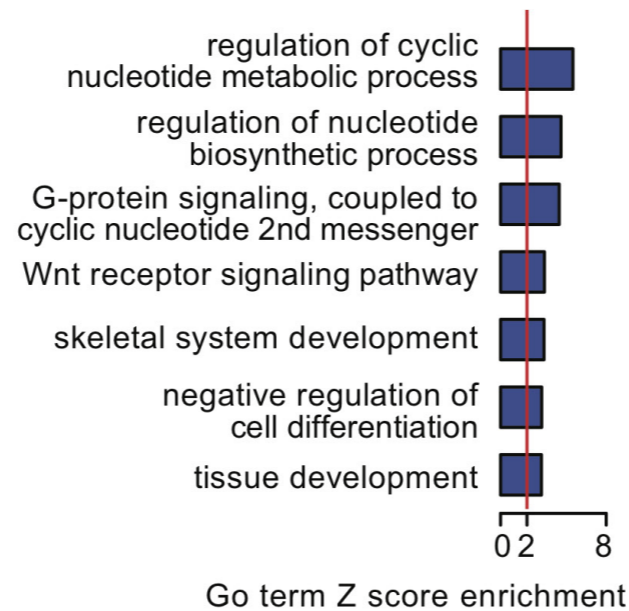


# Attenuation of cortical patterning in ASD



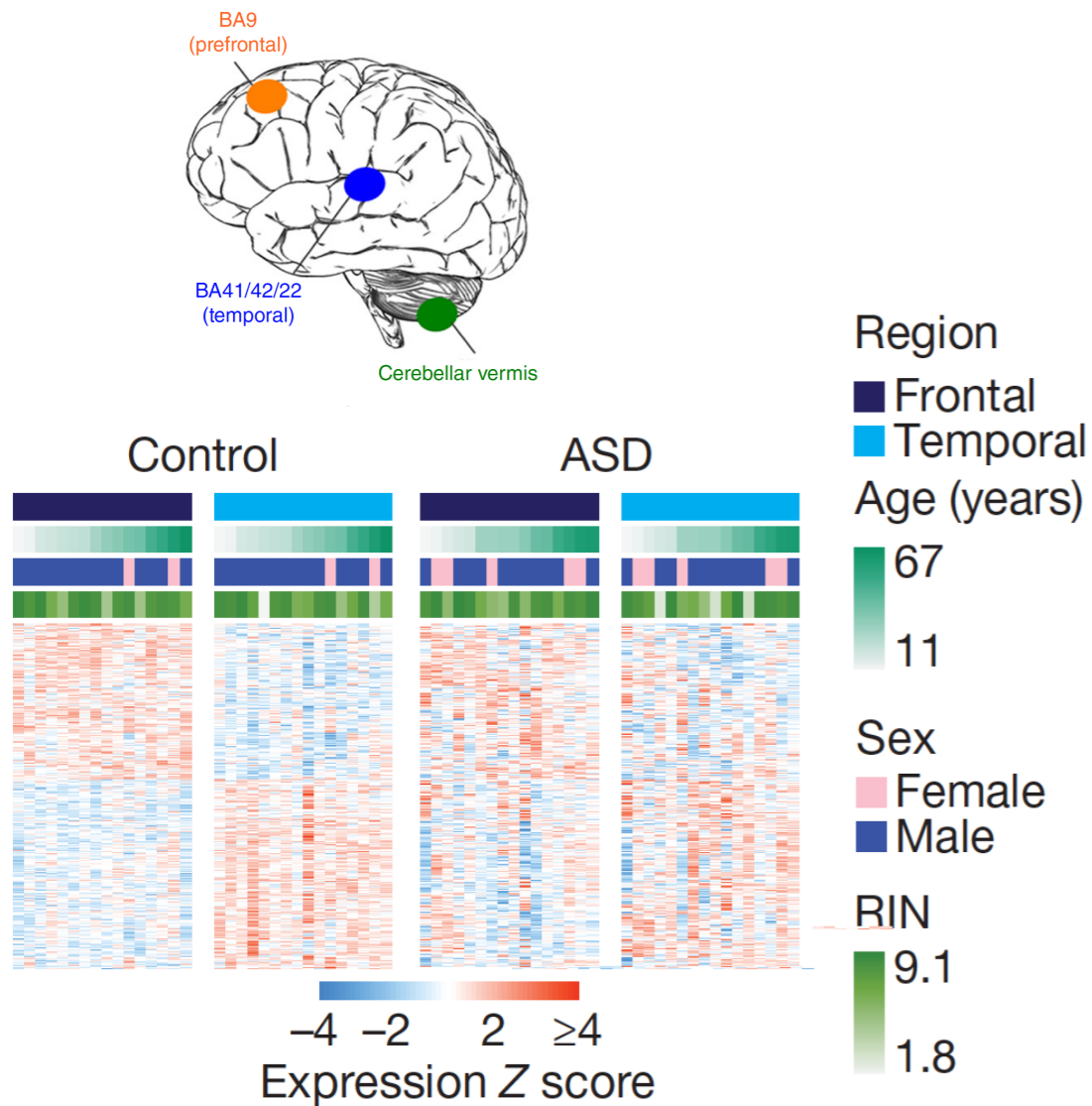
Number of genes DE b/w FC and TC:  
**551** in control samples  
 only **51** in ASD samples

**Attenuated cortical patterning (ACP)** set -  
**523** genes that show DE in control, but not ASD



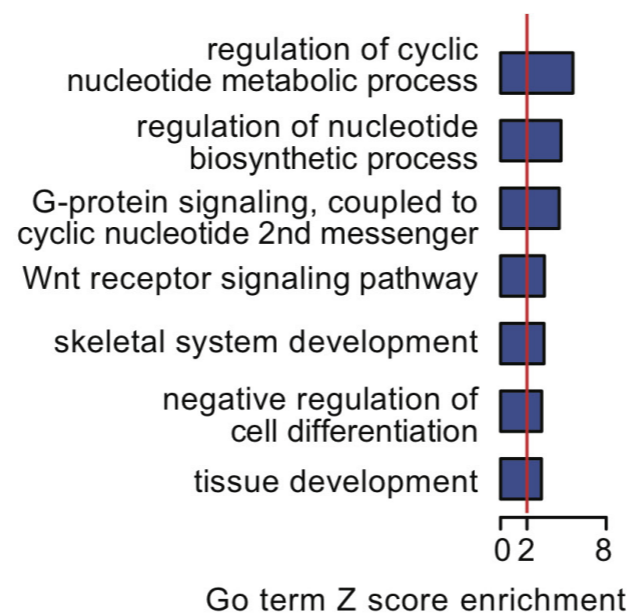


# Attenuation of cortical patterning in ASD

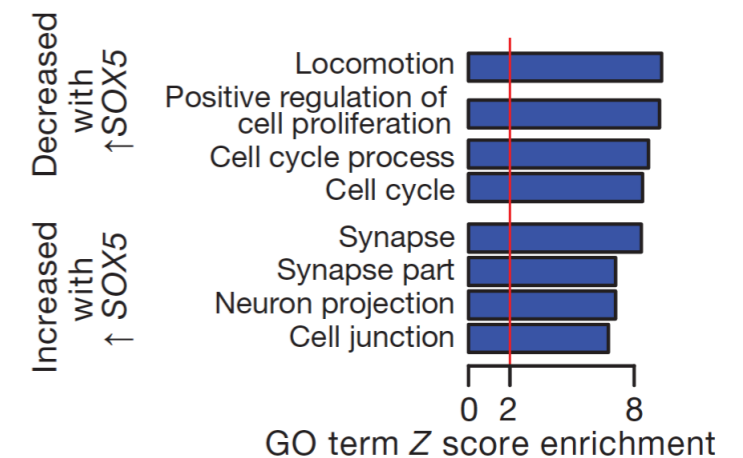
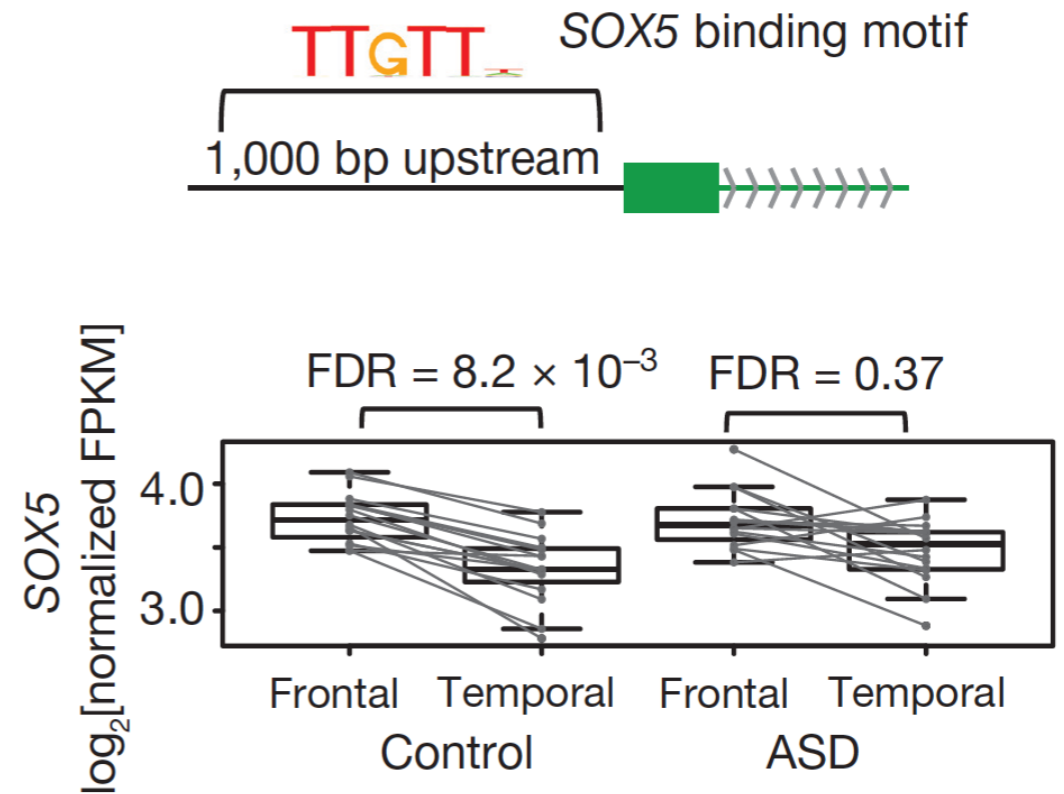


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**Attenuated cortical patterning (ACP)** set -  
**523** genes that show DE in control, but not ASD

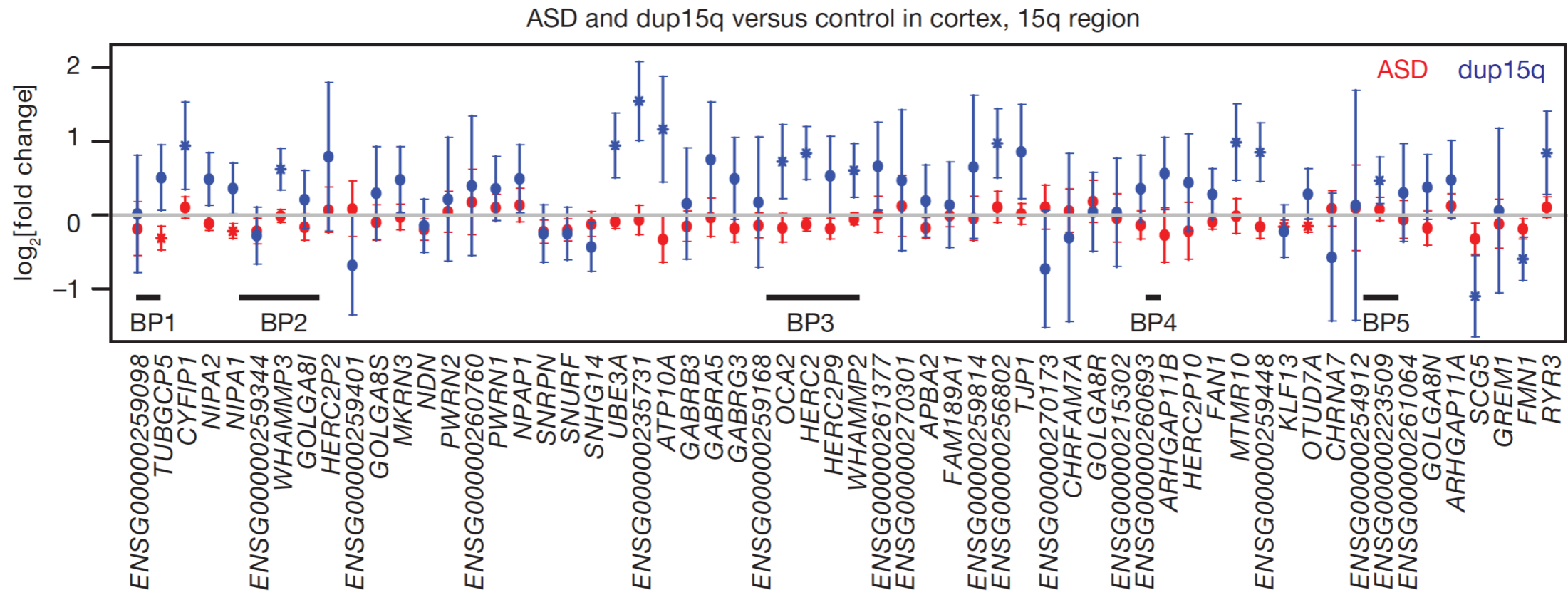


# TF motif enrichment



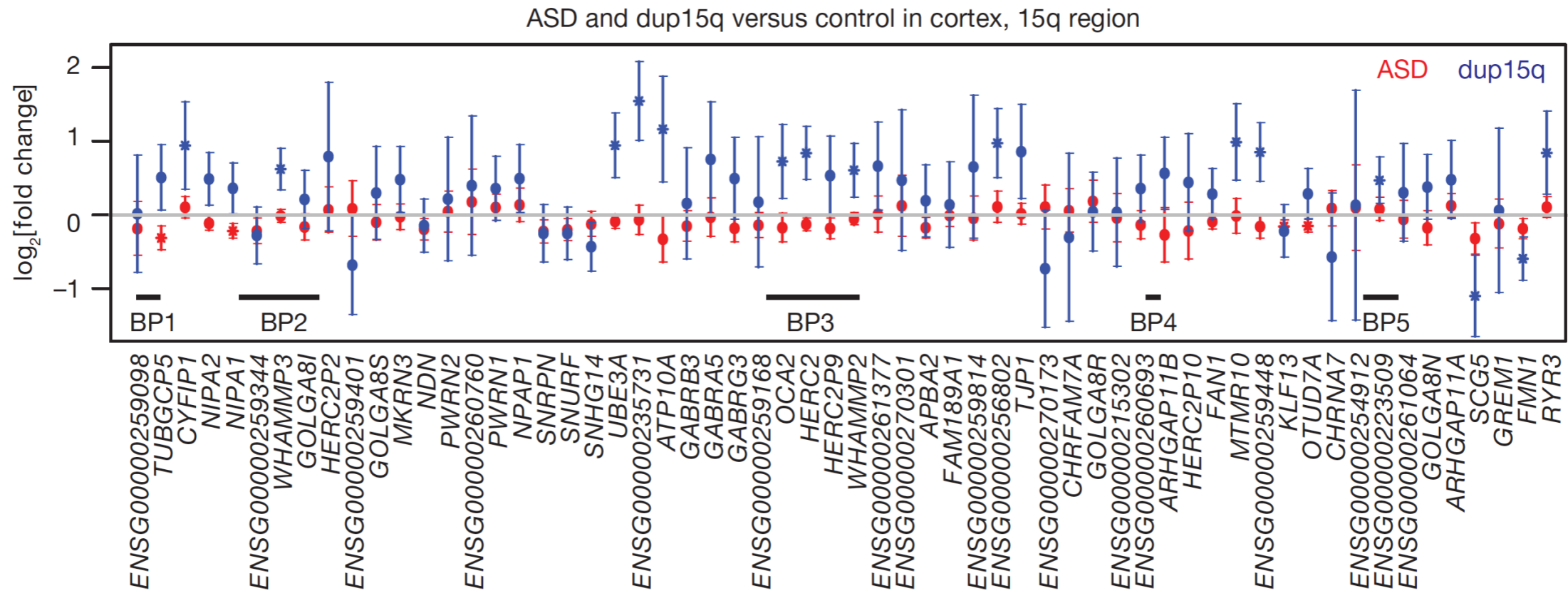
# Duplication 15q syndrome recapitulates transcriptomic changes in idiopathic ASD

**a**

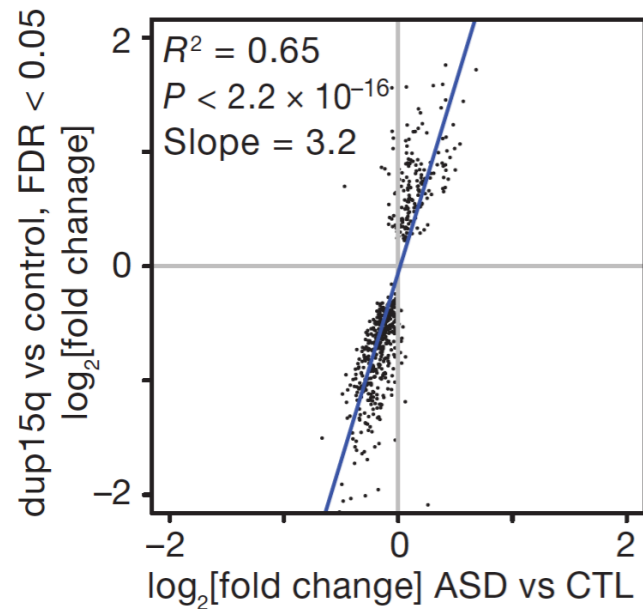


# Duplication 15q syndrome recapitulates transcriptomic changes in idiopathic ASD

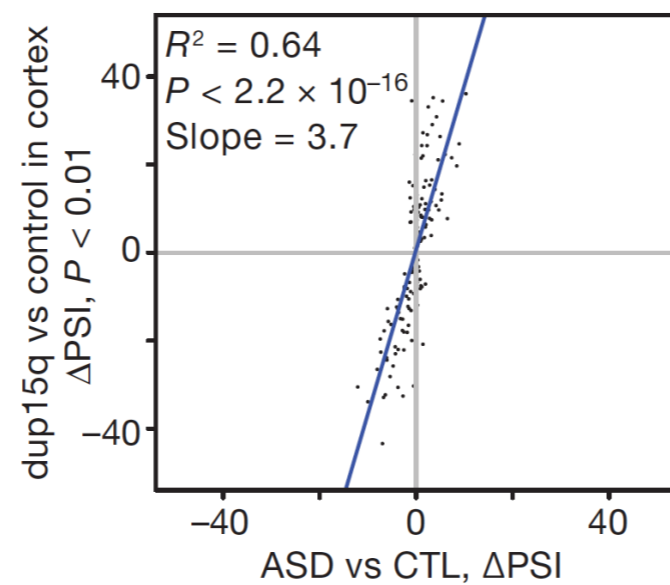
**a**



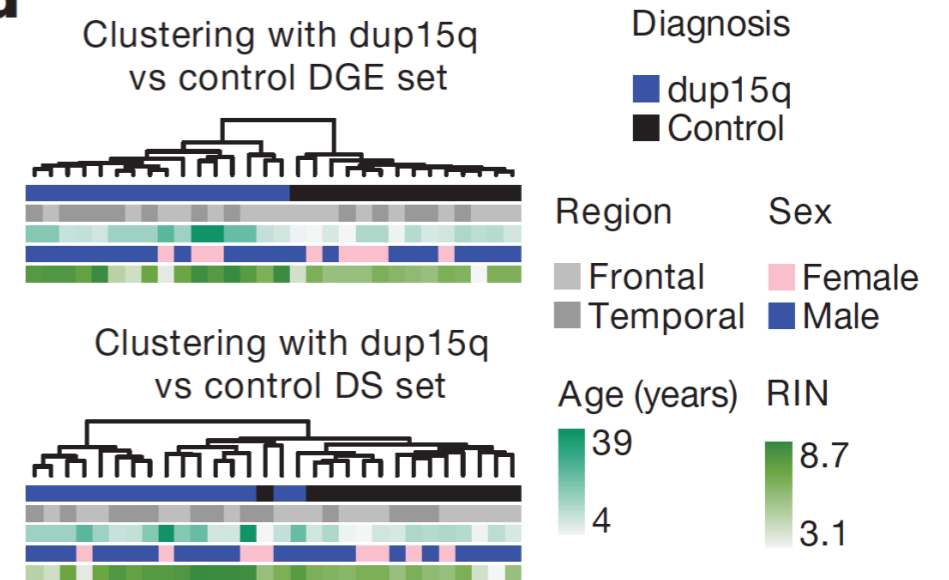
**b**



**c**

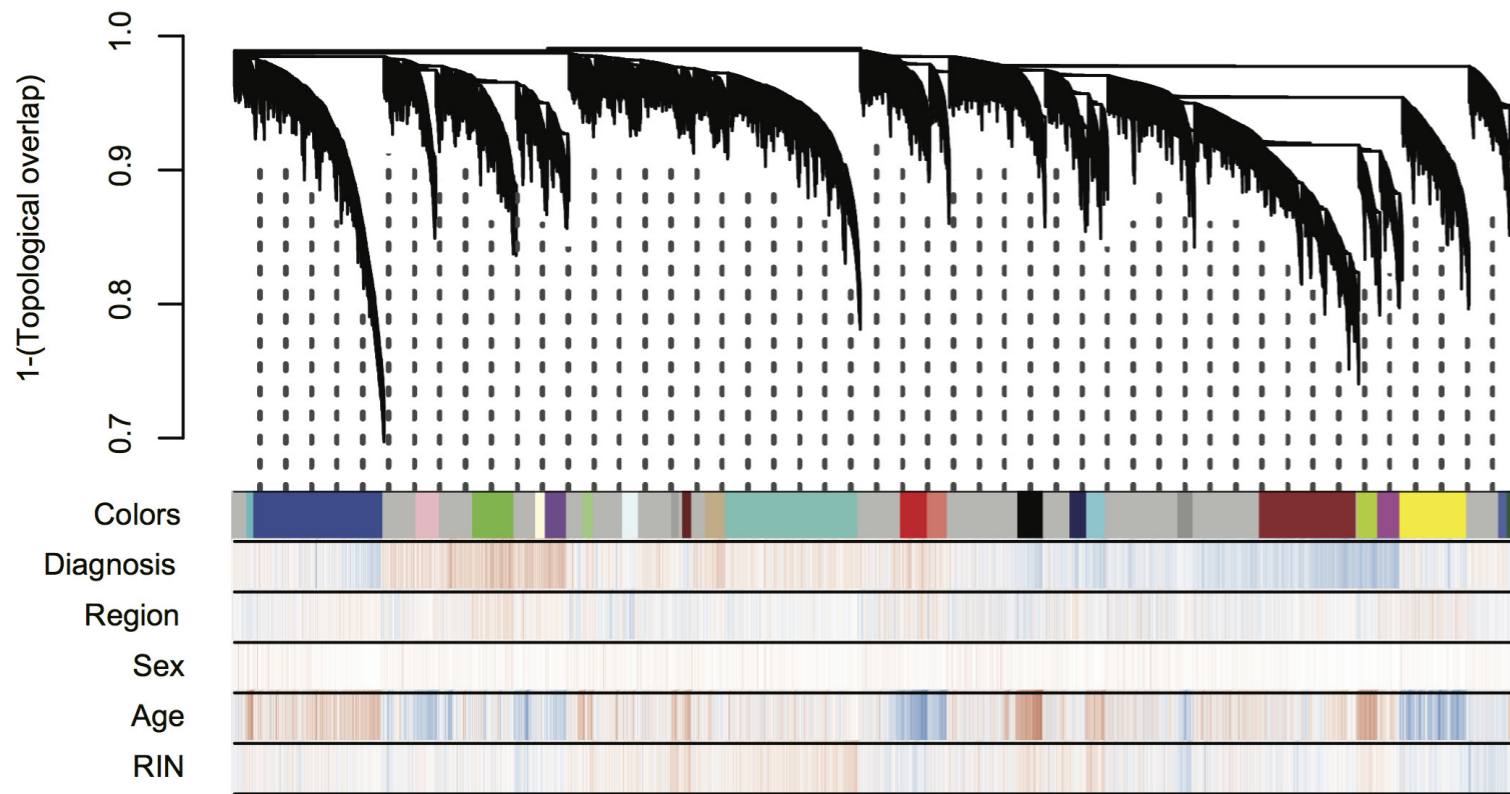


**d**

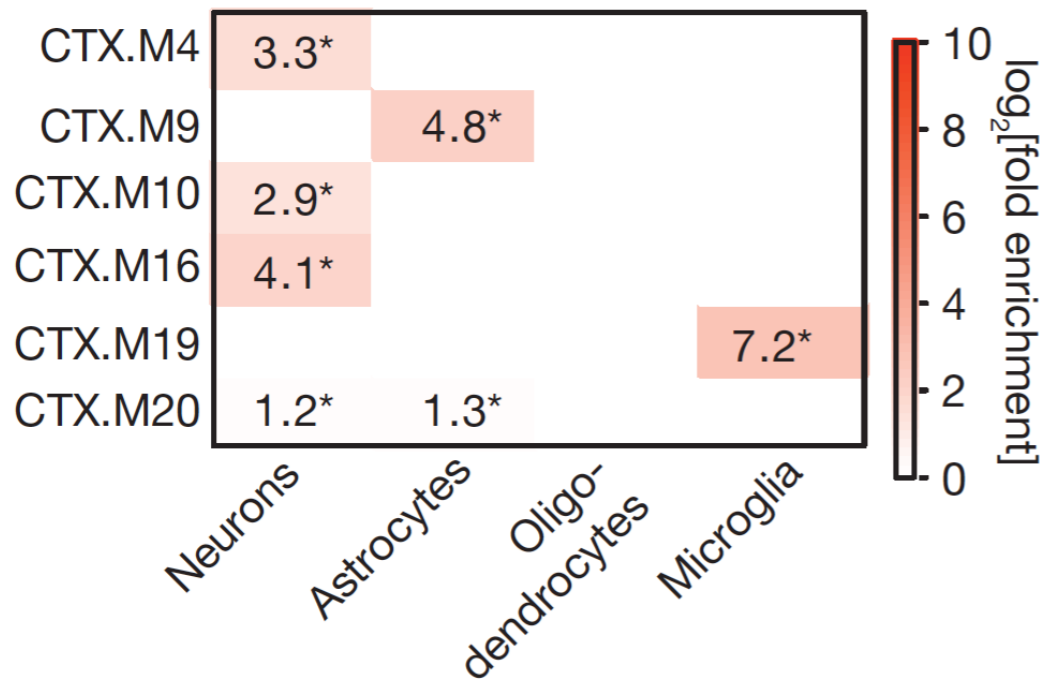


# Co-expression network analysis

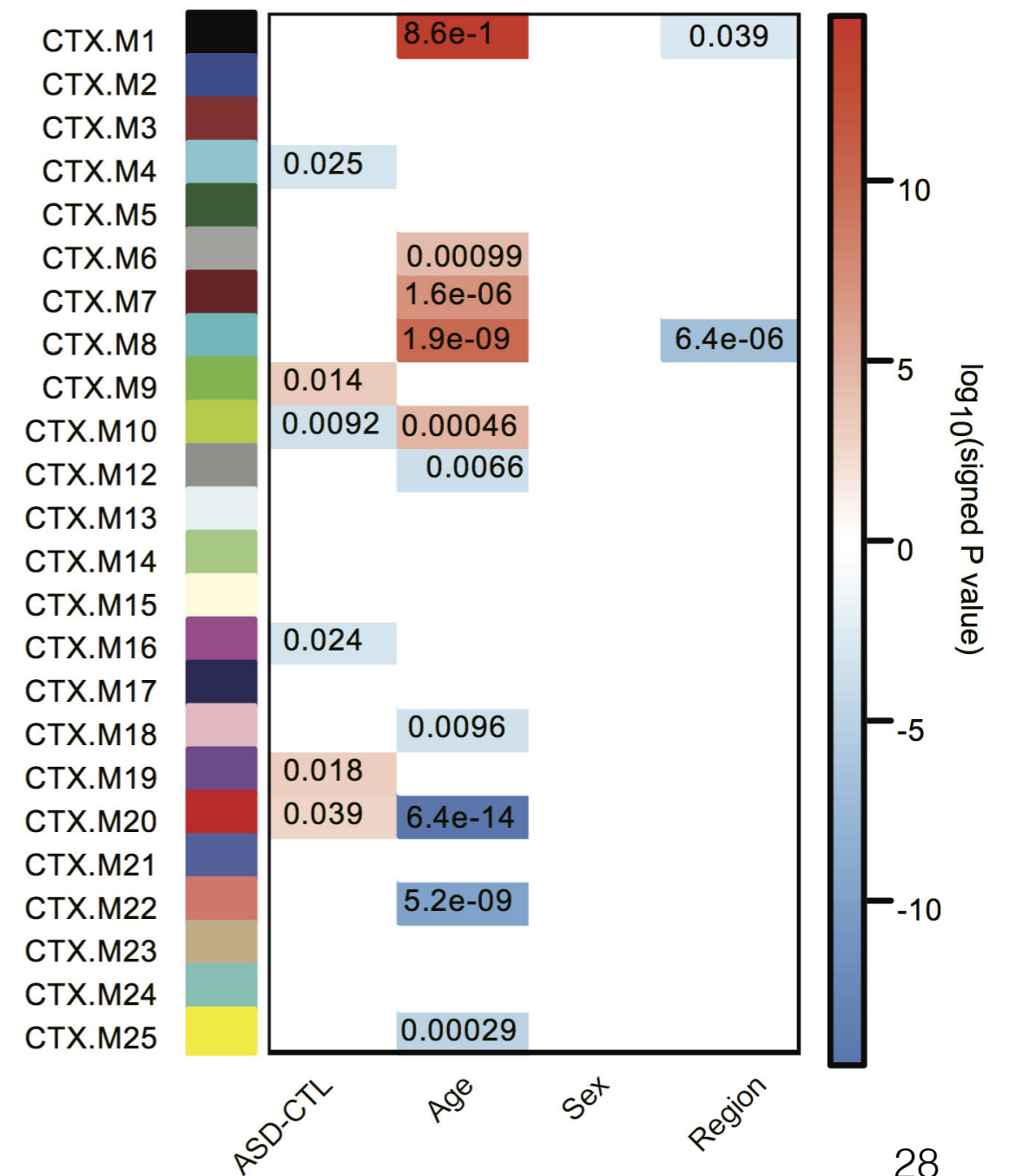
Co-expression network dendrogram for CTX



Fold enrichment  
(\*FDR < 0.05)

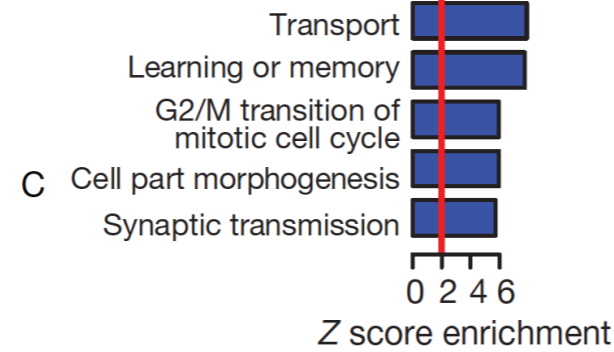
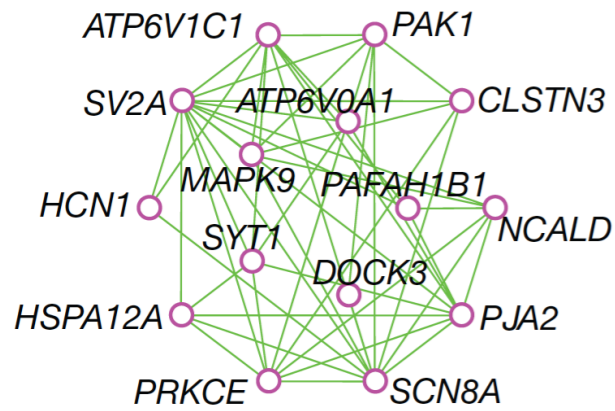


Module eigengene associations with diagnosis and covariates

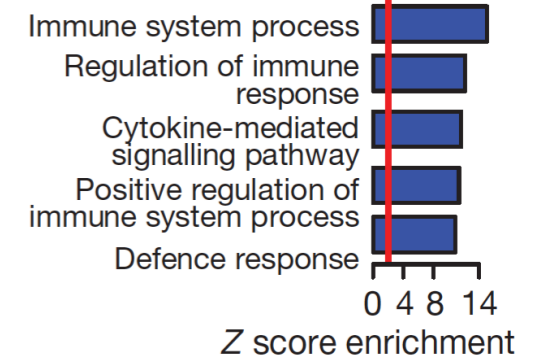
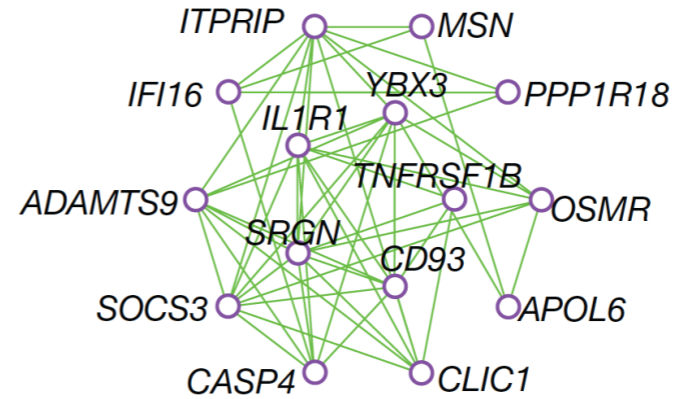


# Co-expression network analysis

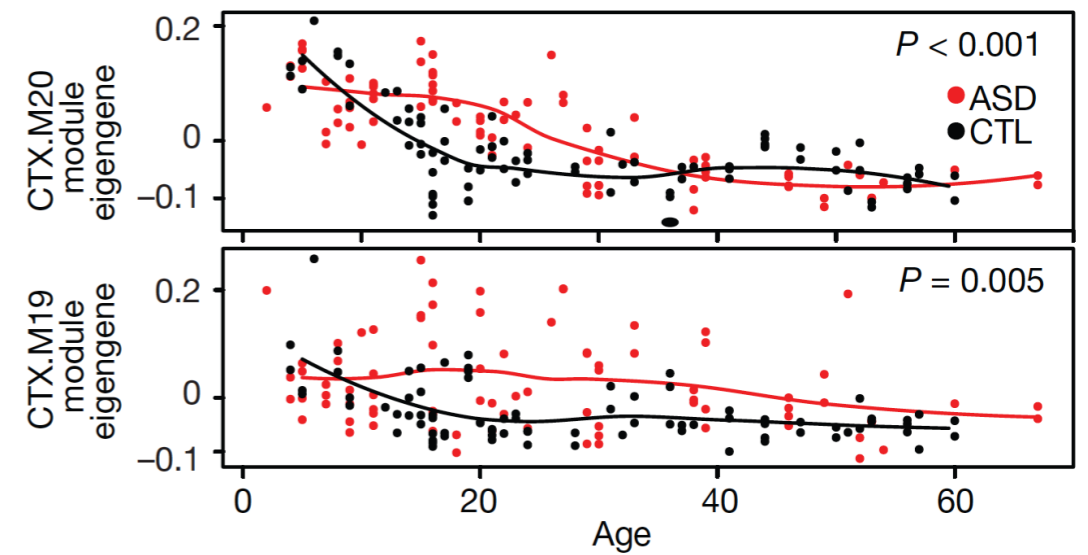
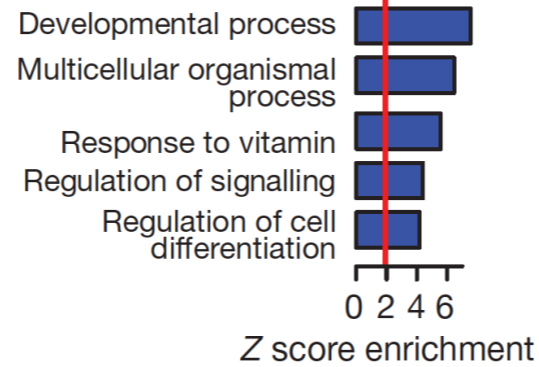
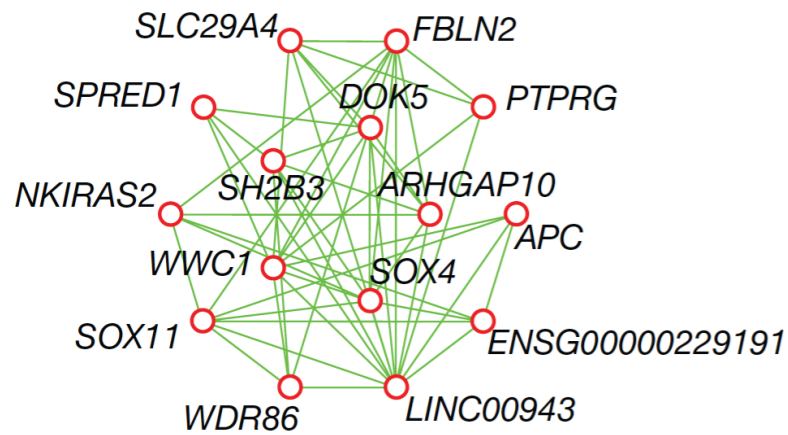
CTX.M16



CTX.M19



CTX.M20



## Related publications from the same lab

- Voineagu et al., *Nature* 2011  
Transcriptomic analysis of autistic brain reveals convergent molecular pathology
- Parikshak et al., *Cell* 2013  
Integrative Functional Genomic Analyses Implicate Specific Molecular Pathways and Circuits in Autism
- Parikshak et al., *Nature Rev Genet*, 2015  
Systems biology and gene networks in neurodevelopmental and neurodegenerative disorders
- de la Torea-Ubieta et al., *Nature Med*, 2016  
Advancing the understanding of autism disease mechanisms through genetics
- Gandal et al., *Nature Neurosci*, 2016  
The road to precision psychiatry: translating genetics into disease mechanisms
- Leppa et al., *Am J Hum Genet*, 2016  
Rare Inherited and De Novo CNVs Reveal Complex Contributions to ASD Risk in Multiplex Families
- Werling et al., *Nature Comm*, 2016  
Gene expression in human brain implicates sexually dimorphic pathways in autism spectrum disorders
- Sun et al., *Cell*, 2016  
Histone Acetylome-wide Association Study of Autism Spectrum Disorder
- Seyfried et al., *Cell Systems*, 2016  
A Multi-network Approach Identifies Protein-Specific Co-expression in Asymptomatic and Symptomatic Alzheimer's Disease
- Won et al., *Nature*, 2016  
Chromosome conformation elucidates regulatory relationships in developing human brain