

# Analysis of Information Leakage in Phenotype and Genotype Datasets

---

Arif Harmanci<sup>1,2</sup>, Mark Gerstein<sup>1,2,3,\*</sup>

1 Program in Computational Biology and Bioinformatics, Yale University, 260 Whitney Avenue, New Haven, CT 06520, USA

2 Department of Molecular Biophysics and Biochemistry, Yale University, 260 Whitney Avenue, New Haven, CT 06520, USA

3 Department of Computer Science, Yale University, 260 Whitney Avenue, New Haven, CT 06520, USA

\*Corresponding authors: Mark Gerstein [pi@gersteinlab.org](mailto:pi@gersteinlab.org)

[[HIGHLIGHTED PARTS TO BE MOVED TO SUPPLEMENTARY]]

## ABSTRACT

Privacy is receiving much attention with the unprecedented increase in the breadth and depth of biomedical datasets, particularly personal genomics datasets. Moreover, considering the legislative plans for encouraging public data sharing in biomedical research fields, privacy will be the key consideration in designing data sharing mechanisms. Most studies on genomic privacy are focused on protection of variants in personal genomes. Molecular phenotype datasets, however, can also contain substantial amount of sensitive information. Although there is no explicit genotypic information in them, subtle genotype-phenotype correlations can be used to statistically link the phenotype and genotype datasets. The links can then be used to characterize individuals by identifying their sensitive phenotypes and breaching privacy. Here, we develop a formalism for the quantification and analysis of individual characterizing information leakage in a linking attack. We analyze the tradeoff between the predictability of the genotypes and the amount of leaked information that can be used in linking and individual characterization. Then we show how one could practically instantiate an attack focusing on the most commonly available data sets, those of RNA-seq and eQTL. We develop a three step procedure showing how an attacker would select eQTLs, statistically predict the genotypes, and perform linking based on the predicted genotypes. The linking can be very accurate considering the high dimensionality of phenotypes. The linking attack becomes particularly easy to perform when one deals with outlier gene expression levels. To study this, we developed a particular realization of this attack for the outlier cases and quantified the amount of information leaked.

## 1 BACKGROUND

Privacy is one of the most important topics of debate in data science that stands at the corner of many different fields, including ethics, sociology, law, political science, and forensic science. Recently, genomics has emerged as one of the major foci of studies on privacy. This can mainly be attributed to the advancement of technologies for high throughput biomedical data acquisition that bring about a surge of datasets<sup>1,2</sup>. Among these, high throughput molecular phenotype datasets, like functional genomic and metabolomic measurements, substantially grow the list of the *quasi-identifiers* (such as birth date, ZIP code, gender<sup>3</sup>) for participating individuals, which can be used by an adversary for re-identification of the identities. With the recent announcement of Precision Medicine Initiative<sup>4</sup>, a large body of datasets are to be generated and shared among researchers<sup>5</sup>. The National Institutes of Health also released the plans to encourage public access to biomedical datasets from scientific studies<sup>5-7</sup>. Considering the fact that one does not need many identifiers to uniquely pinpoint an individual<sup>3,8,9</sup>, these datasets have the potential to exacerbate the risk of privacy breach.

Many consortia, like GTex<sup>10</sup>, ENCODE<sup>11</sup>, 1000 Genomes<sup>12</sup>, and TCGA<sup>13</sup>, are generating large amount of personalized biomedical datasets. Coupled with the generated data, sophisticated analysis methods are being developed to discover correlations between genotypes and phenotypes, some of which can contain sensitive information like disease status. Although these correlations are useful for discovering how genotypes and phenotypes interact, they could also be utilized by an adversary in a linking attack for matching the entries in genotype and phenotype datasets. For example, when a phenotype dataset is available, the adversary can utilize the genotype-phenotype correlations to statistically predict the genotypes, compare the predicted genotypes with the entries in another dataset that contains genotypes. For the entries that are correctly matching, he/she can reveal sensitive phenotypes of the individuals and characterize them. Even when the strength of each genotype-phenotype correlation is not high, the availability of a large number of genotype-phenotype correlations increases the scale of linking. In fact, an adversary can perform correct linking with relatively small number of genotypes<sup>14,15</sup>.

Different aspects of privacy have been intensely studied. Recently, genomic privacy is receiving much attention as a result of the deluge of personalized genomics datasets that are being generated<sup>16,17</sup>. With the increase in the number of large scale genotyping and phenotyping studies, the protection of privacy of participating individuals emerged as an important issue. Homer et al<sup>18</sup> proposed a statistical testing procedure that enables testing whether a genotyped individual is in a pool of samples, for which only the allele frequencies are known. Im et al<sup>19</sup> showed that, given the genotypes of a large set of markers for an individual, an attacker can reliably predict whether the individual participated to a QTL study or not. These attacks, which we refer to as “detection of a genome in a mixture”, are one type of attacks on privacy (Fig S6). There is yet another important attack where the attacker links two or more datasets to pinpoint individuals in datasets and reveal sensitive information. One well-known and illustrative example of these “linking attacks”, although not in a genomic context, is the linking attack that matched the entries in Netflix Prize Database and the Internet Movie Database (IMDB)<sup>20</sup>. For research purposes,

Netflix released an anonymized dataset of movie ratings of thousands of viewers, which they thought was secure as the viewers' names were removed. However, Narayanan et al<sup>20</sup> used IMDB database, a seemingly unrelated and very large database of movie viewers, linked the two databases, and revealed identities and personal information (movie history and choices) of many viewers in the Netflix database. The fact that Netflix and IMDB host millions of individuals in their databases renders the question of detection of an individual in these database irrelevant since any random individual is very likely to be in one or both of these databases but the focus of attacks turns to matching individuals in the databases. Consequently, as the databases grow, the attacks for detection of an individual in a database become unimportant and the linking attacks become more admissible in order to characterize individuals' sensitive information. In the genomic privacy context, as the size and number of the genotype and phenotype datasets increase, possibility of potentially linkable datasets will increase, which may make scenarios similar to Netflix attacks a reality in genomic privacy (Fig S6, Section S2).

Several studies on genomic privacy address the linking of different datasets for re-identifying individuals and characterizing their sensitive information. Gymrek et al<sup>21</sup> revealed the identities of several male participants of 1000 Genomes Project<sup>12</sup> by using the short tandem repeats on Y-chromosome as an individual identifying biomarker and linking the genotypes to online genetic genealogy databases. A detailed review can be found elsewhere<sup>22</sup>. In addition, different formalisms for protecting sensitive information have been proposed and applied to genomic privacy. These censor or hide information, or aim at ensuring statistical indistinguishability of individuals in the released data. For example, differential privacy<sup>23</sup> involves building data release mechanisms that have guaranteed bounds on the leakage of sensitive information. The release mechanisms track how much information is leaked and stops release when the estimated leakage is above a predetermined threshold. Although this approach is theoretically very appealing, it can substantially decrease the utility of the biological data<sup>24</sup>. In addition, the release mechanism must keep track of all the queries, which can cause complications in data sharing<sup>25</sup>. Homomorphic encryption<sup>26</sup> enables performing analysis on encrypted data directly. Complete protection of sensitive information is guaranteed as the data processors never interact with the unencrypted sensitive information. The drawback, however, is high computational and storage requirements. Another well-established formalism is k-anonymization<sup>27,28</sup>. Before releasing the dataset, it is anonymized by data perturbation techniques for ensuring that no combination of features in the dataset are shared by fewer than k individuals. In this approach the anonymization process has, however, excessive computational complexity and is not practical for high dimensional biomedical datasets<sup>29</sup>. Several variants have been proposed for extending k-anonymity framework<sup>30,31</sup>. A majority of these studies aim at protecting the genomic variants and identities of individuals in databases. Different aspects of genomic privacy, pertaining linkability of high dimensional phenotype datasets to genotypes, are yet to be explored.

In this paper, we focus on characterizability of the individuals' sensitive information in the context of linking attacks, where the adversary exploits the genotype-phenotype correlations to link different datasets and potentially reveal sensitive information. In general, the high dimensional phenotype datasets generated in genomic studies harbor a number of phenotypes that contain sensitive information, like disease status, and other phenotypes, while not sensitive, may have subtle correlations

with genomic variant genotypes. Many quantitative phenotypes can be linked to genotypes using public quantitative trait loci (QTL) datasets. Some of the high dimensional genomic quantitative traits and corresponding QTLs are gene expression levels (eQTLs), protein levels (pQTLs<sup>32,33</sup>), DNase hypersensitivity site signals (dsQTLs<sup>34</sup>), ribosome occupancy (rQTLs<sup>35</sup>), DNA methylation levels (meQTLs<sup>36</sup>), histone modification levels (haQTLs<sup>37-39</sup>), RNA splicing (sQTLs<sup>40</sup>), and also higher order traits like network modularity (modQTLs<sup>41</sup>). Other QTLs associated with single dimensional non-genomic phenotypes include body mass index<sup>42</sup>, basal glucose levels<sup>43</sup>, and serum cholesterol levels<sup>32,44</sup>. Each QTL can potentially cause a small amount of genotypic information leakage. As these QTLs are often identified and reported at genomic scale, when an adversary utilizes a large number of QTLs in the attack, he/she can accurately link the sensitive phenotypes to the genotype dataset. Since genotypes can almost perfectly identify an individual, this linking attack can potentially cause a breach of privacy for the individuals who participated in the studies.

Among all the datasets, the most abundant and well-studied genotype-phenotype correlation dataset is expression quantitative trait loci (eQTL) datasets. These datasets are generated by genome-wide screening for correlations between the variant genotypes and gene expression levels usually through RNA sequencing or expression arrays<sup>40,45,46</sup>. The eQTL datasets are especially useful in the context of linking attacks since there is a large and growing compendium of public eQTL datasets<sup>47</sup>. For example, the GTex Project hosts a sizable set of eQTL dataset from multiple studies where the users can view in detail how the genotypes and expression levels are associated<sup>10,41</sup>. In order to demonstrate our results and build the formulations in a specific context, we will focus on eQTL datasets and linking of gene expression and genotype datasets. It is, however, worth noting that most of the results and analyses can be trivially generalized to other types of genotype-phenotype correlations.

One publication<sup>48</sup> relates to our study, where the authors demonstrate that an adversary can build a model for predicting genotypes for eQTLs using gene expression levels. The authors show that given the model, individuals can be identified with high accuracy. Our study follows<sup>48</sup> and generalizes the results in two ways: First we study quantification of characterizing information leakage versus risk of characterization in an information theoretic setting. Secondly, we show that the linking can be performed in a much simplified setting by just utilizing the outliers in the data. For this, we introduce a new metric, we termed extremity, and show that this metric can be utilized in genotype prediction and linking attacks with high accuracy.

The paper is organized as follows: We first analyze the genotype predictability and evaluate the tradeoff between the amount of information leakage and correct predictability of the genotypes. Next we present the 3 step individual characterization framework and study different aspects of vulnerability using the framework. In the last section, to illustrate the practicality of the attack scenario, we present extremity based genotype prediction method and evaluate the fraction of characterizable individuals on the representative dataset and we also present several different scenarios to test accuracy of the proposed linking attack. The analysis tools and code are available for download at <http://privaseq.gersteinlab.org>.

## 2 RESULTS

### 2.1 Overview of the Individual Characterization Scenario by Linking Attacks

Figure 1a illustrates the general privacy breaching scenario that is considered. There are three datasets in the context of the breach. First dataset contains the phenotype information for a set of individuals. The phenotypes can include sensitive information such as disease status in addition to several molecular phenotypes such as gene expression levels. The second dataset contains the genotypes and the identities for another set of individuals. The third dataset contains correlations between one or more of the phenotypes in the phenotype dataset and the genotypes. In this dataset, each entry contains a phenotype, a variant, and the degree to which these values are correlated. In order to formulate and demonstrate the results, we will focus on the gene expression datasets as the representative phenotype dataset. As explained earlier, the abundance of gene expression-genotype correlation (eQTL) datasets makes these datasets most suitable for linking attacks.

Figure 1b illustrates the eQTL, expression, and genotype datasets. The eQTL dataset is composed of a list of gene-variant pairs such that the gene expression levels and variant genotypes are significantly correlated. We will denote the number of eQTL entries with  $q$ . The eQTL (gene) expression levels and eQTL (variant) genotypes are stored in  $q \times n_e$  and  $q \times n_v$  matrices  $e$  and  $v$ , respectively, where  $n_e$  and  $n_v$  denotes the number of individuals in gene expression dataset and individuals in genotype dataset. The  $k^{th}$  row of  $e$ ,  $e_k$ , contains the gene expression values for  $k^{th}$  eQTL entry and  $e_{k,j}$  represents the expression of the  $k^{th}$  gene for  $j^{th}$  individual. Similarly,  $k^{th}$  row of  $v$ ,  $v_k$ , contains the genotypes for  $k^{th}$  eQTL variant and  $v_{k,j}$  represents the genotype ( $v_{k,j} \in \{0,1,2\}$ ) of  $k$  variant for  $j^{th}$  individual. The coding of the genotypes from homozygous or heterozygous genotype categories to the numeric values are done according to the correlation dataset (See Methods Section 4.1). We assume that the variant genotypes and gene expression levels for the  $k^{th}$  eQTL entry are distributed randomly over the samples in accordance with random variables (RVs) which we denote with  $V_k$  and  $E_k$ , respectively. We denote the correlation between the RVs with  $\rho(E_k, V_k)$ . In most of the eQTL studies, the value of the correlation is reported in terms of a gradient (or the regression coefficient) in addition to the significance of association (p-value) between genotypes and expression levels. The absolute value of  $\rho(E_k, V_k)$  indicates the strength of association between the eQTL genotype and the eQTL expression level. The sign of  $\rho(E_k, V_k)$  represents the direction of association, i.e., which homozygous genotype corresponds to higher expression levels. This forms the basis for correct predictability of the eQTL genotypes using eQTL expression levels: The homozygous genotypes associate with the extremes of the gene expression levels and the heterozygous genotypes associate with moderate levels of expression. The eQTL studies utilize linear models to identify the gene and variant pairs whose expressions and genotypes that are significantly correlated. Given this knowledge, the adversary aims at reversing this operation so as to predict genotypes for each individual, using the respective gene expression levels and the genotype-phenotype correlation. For general applicability of the analysis, we assume that he/she utilizes a prediction model that estimates correctly the *a posteriori* distribution of the eQTL genotypes given the eQTL expression levels, i.e.,  $p(V_k|E_k)$ , as illustrated in Fig S2b. This enables us to perform the analysis independent of the prediction methodology that the attacker utilizes without making any assumptions on the prediction model that is utilized by the attacker.

## 2.2 Quantification of Tradeoff between Correct Predictability of Genotypes and Leakage of Individual Characterizing Information

[[MOVE TO SUPP, REPHRASE FOLLOWING]]

We assume that the attacker will behave in a way that maximizes his/her chances of characterizing the most number of individuals. Thus, he/she will try and predict the genotypes, using the phenotype measurements, for the largest set of variants that he/she believes he/she can predict correctly. The most obvious way that the attacker does this is by first sorting the genotype-phenotype pairs with respect to decreasing strength of correlation as illustrated in Fig 2a. He/She will then predict the genotypes starting from the top genotype-phenotype pair. As he/she predicts more genotypes, he/she increases his/her chances of characterizing more individuals. As the attacker goes down the list, however, the correct predictability of the genotypes diminish, i.e., the strength of genotype-phenotype correlation decreases. Thus, each time he/she predicts a new genotype, he/she will encounter a tradeoff between the number of genotypes that can be predicted correctly versus the cumulative correctness of all the predicted genotypes. This tradeoff can also be viewed as the tradeoff between precision (fraction of the linkings that are correct) and recall (fraction of individuals that are correctly linked). In this section we will propose two measures to quantify this tradeoff.

In the context of the linking attack, the attacker aims to correctly characterize  $n_e$  individuals in the expression dataset among  $n_v$  individuals in the genotype dataset. In order to correctly characterize an individual, he/she should select a set of eQTLs that he/she believes he/she can predict correctly. Next, given the individual's expression levels, the attacker should predict the genotypes for the selected eQTLs correctly such that the predicted set of genotypes are not shared by more than 1 individual, i.e., the predicted genotypes can be matched to the correct individual. In other words, the joint frequency of the set of predicted genotypes for the selected eQTLs should be  $\frac{1}{n_v}$ . We can rephrase this condition as following in information theoretic terms: Given the genotypes of an individual, if the attacker can correctly predict a subset of genotypes that contain at least  $\log_2(n_v)$  bits of information, the individual is vulnerable to characterization of his/her phenotypes. Following this statement, we can quantify the leakage from a set of correctly predicted eQTL variant genotypes as the logarithm of their joint frequency. Assuming that the genotypes of different eQTLs (See Section 5) are independent from each other, we can decompose the quantity of individual characterizing information that is leaked for a set of  $n$  correctly predicted eQTL genotypes:

$$ICI(\{V_1 = g_1, V_2 = g_2, \dots, V_n = g_n\}) = \sum_{k=1}^n \underbrace{-\log(p(V_k = g_k))}_{\substack{\text{Convert the genotype} \\ \text{frequency to number of bits} \\ \text{that can be used to characterize} \\ \text{individual}}} \quad \text{Sum individual characterizing information from all variants} \quad (1)$$

where  $V_k$  is the random variable that corresponds to the genotypes for the  $k^{\text{th}}$  eQTL,  $g_k$  is a specific genotype (Refer to Methods Section 3.1 for more details), and  $p(V_k = g_k)$  denotes the genotype

frequency of  $g_k$  within the population, and  $ICI$  denotes the total individual characterizing information. Evaluating the above formula,  $ICI$  increases as the frequency of the variant's genotype  $g_k$  decreases. In other words, the more rare genotypes contribute higher to  $ICI$  compared to the more common ones. Thus, individual linking information can be interpreted as a quantification of how rare the predicted genotypes are. The attacker aims to predict as many eQTLs as possible such that  $ICI$  for the predicted genotypes is at least  $\log(n_v)$ .  $ICI$  can also be interpreted as the number of rare SNP genotypes that an individual harbors.

In order to maximize the amount of  $ICI$ , the attacker will aim at correctly predicting as many eQTL genotypes as possible. The (correct) predictability of the eQTL genotypes from expression levels, however, varies over the eQTL dataset as some of the eQTL genotypes are more highly correlated (i.e., more correctly predictable) with the expression levels compared to others, given in  $|\rho(E_k, V_k)|$ . Thus, the attacker will try to select the eQTLs whose genotypes are the most correctly predictable to maximize  $ICI$  leakage. Although  $\rho(E_k, V_k)$  is a measure of predictability, it is computed differently in different studies. In addition, there is no easy way to combine these correlation values when we would like to estimate the joint predictability of multiple eQTL genotypes. In order to uniformly quantify the joint (correct) predictability of the eQTL genotypes using the expression levels, we use the exponential of entropy of the conditional genotype distribution given gene expression levels. Given the expression levels for  $j^{th}$  individual, we compute the predictability of the  $k^{th}$  eQTL genotypes as

$$\pi(V_k | E_k = e_{k,j}) = \exp(-1 \times \underbrace{H(V_k | E_k = e_{k,j})}_{\text{Convert the entropy to average probability}})$$

Randomness left in  $V_k$   
given  $E_k = e_{k,j}$

(2)

where  $\pi$  denotes the predictability of  $V_k$  given the gene expression level  $e_{k,j}$ .  $\pi$  can be interpreted as the average probability (when sampling individuals from the population) that the attacker can correctly predict the eQTL genotype at the given expression level. In the above equation for  $\pi$ , the conditional entropy of the genotypes is a measure for the randomness that is left in genotype distribution when the expression level is known. In the case of high predictability, the conditional entropy is close to 0, and there is little randomness left in the genotype distribution. Taking the exponential of negative of the entropy converts the entropy to average probability of correct prediction of the genotype. In the most predictable case (conditional entropy close to 0),  $\pi$  is close to 1, indicating very high predictability (Refer to Methods Section 4.1 for more details).

We first considered each eQTL and evaluated the genotype predictability versus the characterizing information leakage. We use the GEUVADIS dataset as a representative dataset for this computation (Refer for Section 5). We computed, for each eQTL, average  $\pi$  and average  $ICI$  over all the individuals, which is plotted in Fig 2b. Most of the data points are spread along the diagonal, which indicate that there is a natural tradeoff between correct predictability and  $ICI$  leakage. The eQTL variants with high frequency major allele frequencies have high predictability and low  $ICI$  and vice versa for eQTL variants with lower major allele frequency (Fig 2b, left). This is expected because the genotypes of the high frequency variants can be predicted, on average, easily (most individuals will harbor one dominant

genotype) and consequently does not deliver much characterizing information. The genotypes for the eQTLs with smaller major frequency alleles, however, are harder to predict as they are mostly uniformly distributed among population. On the other hand, these eQTLs contain high *ICI* on average. The eQTLs with high correlation (Fig 2b, right) deviate from the diagonal with high *ICI* and high predictability. In principle, the adversary will aim at identifying and using these highly informative eQTLs. The shuffled gene-variant pairs, on the other hand, are distributed mainly along the diagonal (Fig S1a).

## [[FOLLOWING CAN BE MOVED AND REPHRASED]]

The risk of characterizability increases substantially when the adversary utilizes multiple genotype predictions at once. We will now use *ICI* and  $\pi$  to evaluate how predictability changes with increasing leakage when multiple genotypes are utilized. As discussed earlier, the attacker will aim at predicting the largest number of eQTL genotypes given the expression levels to maximize characterization power. For this, we assume the attacker will sort the eQTLs with respect to the absolute value of correlation then predict the eQTL genotypes starting from the first eQTL. In order to evaluate the tradeoff between the characterizing information of the top predictable eQTLs and their predictabilities, we plotted average *ICI* versus average  $\pi$  for top genotype predictions. For this, we first sorted the eQTLs with respect to the reported correlation,  $|\rho(E_k, V_k)|$ . Then for top  $n=1,2,3,\dots,20$  eQTLs, we estimated mean  $\pi$  and mean *ICI* over all the samples as illustrated in Fig S2a. We then plotted mean  $\pi$  versus mean *ICI* for each  $n$  which is shown in Fig 2c. From the plot, we can first estimate the number of vulnerable individuals at different predictability levels. For example, at 20% predictability, there is approximately 8 bits of *ICI* leakage. At this level of leakage, the adversary can correctly link all individuals, on average with 20% chance, in a sample of  $2^8 = 256$  individuals. At 5% predictability, the leakage is 11 bits and the characterizable sample size is  $2^{11} = 2048$  individuals, which can be interpreted as a higher risk of characterizability. These estimates are useful when releasing QTL datasets such that the leakage risks can be assessed besides the released list of genotype-phenotype correlations. Another view is to evaluate the risk at which a given sample of individuals can be characterized. For a dataset of  $n_v$  individuals, as explained earlier, it is necessary to predict  $\log(n_v)$  bits of genotypic information correctly. The risk of characterization can be determined from the graph as the predictability level at which  $\log(n_v)$  bits of *ICI* leakage is observed. The auxiliary information knowledge can also be incorporated into this analysis easily. For example, assuming that the sample set contains 10,000 individuals, it is necessary to correctly predict  $\log(n_v = 10,000) = 13.3$  bits of information. At around 5% predictability, the adversary can gain 11 bits of information. Even though this cannot uniquely characterize all individuals, if the attacker can gain  $13.3 - 11 = 2.3$  bits of auxiliary information, e.g. gender and ethnicity, he/she can characterize all individuals correctly. Since many phenotypic measurements have significant predictive power for gender, the attacker can predict it correctly, which gains the attacker 1 bit of auxiliary information. The presented quantification procedure can be utilized for evaluating the risk of information leakage while releasing QTL datasets. For example, the QTLs to be released can be assessed in terms of the characterizing information leakage versus the predictability so as to estimate the size and risk of a linking attack (Fig S8) that would be mediated by these QTLs.



## 2.3 A General Framework for Analysis of Individual Characterization

[[REPHRASE FOLLOWING]]

In this section, we present a 3 step framework for individual characterization in the context of linking attacks. Figure 3 summarizes the steps in the individual characterization for each individual. The input is the phenotype measurements for  $j^{th}$  individual. The aim of the attacker is to correctly link the disease state of the individual to the correct identity in the genotype dataset. In the first step, the attacker selects the QTLs, which will be used in linking  $j^{th}$  individual. The selection of QTLs can be based on different criteria. As described in the previous section, the most accessible criterion is selection based on the absolute gradient or the absolute strength of association between the phenotypes and genotypes. In the case of eQTLs, this is the reported correlation coefficient,  $|\rho(E_k, V_k)|$ . In our analysis, we evaluate the effect of changing correlation coefficient. It is worth noting that the adversary can use other measures of correct predictability to select the set of QTLs that he/she will utilize in the linking process. The second step is genotype prediction for the selected QTLs using a prediction model. For general applicability of our analysis we are assuming that the attacker's prediction model can reliably construct the posterior probability distribution of the genotypes given the phenotypes. The attacker then uses the posterior probabilities of the genotypes to identify the maximum *a posteriori* (MAP) genotype. In this prediction, the attacker assigns the genotype that has the highest *a posteriori* probability given the expression level (Refer to Methods Section 4.3). The third and final step of individual characterization is comparison of the predicted genotypes to the genotypes of the  $n_v$  individuals in genotype dataset to identify the individual that matches best to the predicted genotypes. In this step, the attacker links the predicted genotypes to the individual in the genotype dataset with the smallest number of mismatches compared to the predicted genotypes (Refer to Methods Section 4.4).

### 2.3.1 Fraction of Vulnerable Individuals with MAP Genotype Prediction

[[MOVE TO SUPPLEMENTARY, MENTION IN THE NEXT SECTION]]

To illustrate the results of linking attack, we evaluate the fraction of individuals that are vulnerable to characterization using gene expression and genotype data in GEUVADIS Project. We assume that the attacker uses the absolute value of the reported correlation between the variant genotypes and gene expression levels to select the eQTLs for characterization. The genotypes for the selected eQTLs are predicted using MAP prediction (Refer to Methods Section 4.3). Figure 4a shows, for each correlation threshold, the number of selected eQTLs and the fraction correctly predicted genotypes.

Using the list of predicted eQTL genotypes selected at each absolute correlation cutoff, the attacker performs the 3<sup>rd</sup> step in the attack and links the predicted genotypes to the genotype dataset to identify individuals (Refer to Methods Section 4.4). Each individual in expression dataset, who is linked to the right individual are flagged as vulnerable. Figure 4b shows the fraction of vulnerable individuals. The fraction of vulnerable individuals increase as the absolute correlation threshold increases and fraction is maximized at around 0.35 (Fig S3). At this value, 95% of the individuals are vulnerable. This behavior can

be explained by the increase in characterizing information leakage as the accuracy of the predicted genotypes increase while there is a balancing decrease in the characterizing information leakage with decreasing number of eQTL genotypes predicted.

We also evaluate the scenario when the attacker gains access to auxiliary information. As the sources of auxiliary information, we use the gender and population information that is available for all the participants of 1000 Genomes Project on the project web site. It has been previously shown that gene expression levels show widespread differences with respect to gender<sup>49</sup>. In addition, it has been shown that the ethnicity and population differences can be observed in the gene expression levels<sup>50,51</sup>. These indicate that gender and ethnicity can be inferred from gene expression levels. We assume that the attacker either gains access to or predicts the gender and/or the population of the individuals and uses the information in the 3<sup>rd</sup> step of the attack (Refer to Methods Section 4.4). Figure 4b shows the fraction of vulnerable individuals when the auxiliary information is available. When the auxiliary information is available, more than 95% of the individuals are vulnerable to characterization for all the eQTL selections up to when the absolute correlation threshold is 0.6. These results show that a significant fraction of individuals are vulnerable for most of the correlation thresholds that the attacker can choose.

## 2.4 Individual Characterization using Extremity based Genotype Prediction

[[REPHRASE FOLLOWING]]

In the previous section, we presented a general framework for analysis of vulnerability. For the applicability of the framework in different genotype prediction scenarios, we assumed that the attacker can correctly reconstruct the *a posteriori* distribution of genotypes given the gene expression levels, which is then used to estimate the MAP genotype. In general, correct reconstruction of the *a posteriori* distribution of the genotypes given expression levels may not be possible because the knowledge of only the genotype-phenotype correlation coefficient via eQTLs is not enough to regenerate the *a posteriori* distribution of genotypes given the expression levels.

The attacker can, however, utilize a priori knowledge about the relation between gene expression levels and genotypes and build the joint genotype-expression distributions using models with varying complexities and parameters (See Methods Section 4.8). Even though the genotype prediction with these models may not be very accurate, the attacker can utilize a large number of eQTLs to maximize the accuracy of linking (Detailed in the Background Section). We focus on a highly simplified model here. We will assume the attacker exploits the knowledge that the eQTL genotypes and expression levels are correlated such that the allelic effects on expression are additive and extremes of the gene expression levels (highest and smallest expression levels) are observed with extremes of the genotypes (homozygous genotypes). Therefore, given the gradient of association, the attacker can estimate coarsely the joint distribution of the genotypes and expression levels. This idea is illustrated in Fig 5a. Using an estimate of the joint distribution, the attacker can compute the *a posteriori* distribution of genotypes given gene expression levels. To quantify the extremeness of expression levels, we use a statistic we termed *extremity*. For the gene expression levels for  $k^{th}$  eQTL,  $e_k$ , *extremity* of the  $j^{th}$  individual's expression level,  $e_{k,j}$ , is defined as

$$extremity(e_{k,j}) = \frac{\text{rank of } e_{k,j} \text{ in } \{e_{k,1}, e_{k,2}, \dots, e_{k,n_e}\} - 0.5}{n_e} \quad (3)$$

Extremity can be interpreted as a normalized rank, which is bounded between -0.5 and 0.5. Figure S4a shows the median absolute extremity distribution of all the gene expression levels among the individuals. The average median extremity is uniformly distributed among individuals. Figure S4b shows the median number of genes with minimum extremity. Almost half of the genes in each individual have higher than 0.3 extremity in the population. Also, around 1000 genes have higher than 0.45 absolute extremity. In other words, each individual harbors substantial number of genes whose expressions are at the extremes within the population. These can potentially serve as quasi-identifiers. It is worth noting, however, that not all of these extreme genes are associated with eQTLs (See Sections S1 and S6, Figure S7).

Following from the above discussion, the adversary builds the posterior distribution for  $k^{th}$  eQTL genotypes as

$$P(V_k = 0 \mid E_k = e_{k,j}) = \begin{cases} 0 & \text{if } extremity(e_{k,j}) \times \rho(E_k, V_k) > 0 \\ 1 & \text{otherwise} \end{cases} \quad (4)$$

$$P(V_k = 2 \mid E_k = e_{k,j}) = \begin{cases} 1 & \text{if } extremity(e_{k,j}) \times \rho(E_k, V_k) > 0 \\ 0 & \text{otherwise} \end{cases} \quad (5)$$

$$P(V_k = 1 \mid E_k = e_{k,j}) = 0. \quad (6)$$

From the *a posteriori* probabilities, when the sign of the extremity and the reported correlation are the same, the attacker assigns the genotype value 2, and otherwise, genotype value 0. Finally, the genotype value 1 is never assigned in this prediction method, i.e., the *a posteriori* probability is zero. This is expected since we are focusing on the extremes and heterozygous genotype is observed at medium levels of expression. The posterior distribution of genotypes in equations (4-6) can be derived from a simplified model of the genotype-expression distribution that utilizes just one parameter (See Methods Section 4.8, Fig S9). As yet another way of interpretation, the genotype prediction can be interpreted as a rank correlation between the genotypes and expression levels and choosing the homozygous genotypes that maximize the absolute values of the rank correlation. Thus, this process can be generalized as a rank correlation based prediction. We used the posterior genotype probabilities in extremity based prediction and assessed the genotype prediction accuracy. Figure 5b shows the accuracy of genotype predictions with changing correlation threshold. As expected, the accuracy of genotype predictions increases with increasing correlation threshold. The slight decrease of genotype accuracy at correlation thresholds higher than 0.7 is caused by the fact that the accuracy (fraction of correct genotype predictions within all genotypes) is not robust at very small number of SNPs. Although we expect very high accuracy, even one wrong prediction among small number of total genotypes decreases the accuracy significantly.

We next utilized extremity based genotype prediction in the 2<sup>nd</sup> step of the individual characterization framework (Fig 3) and evaluated the fraction of characterizable individuals in the GEUVADIS dataset. We utilized the correlation based eQTL selection in step 1, then extremity based genotype prediction in step 2. In order to demonstrate the utility of the 3-step analysis framework; we evaluated two different distance measures for linking the predicted genotypes to the individuals in genotype dataset in the 3<sup>rd</sup> step of the attack. First is based on comparison of the predicted genotypes to all the genotypes in genotype dataset. Second is based on comparison of the predicted genotypes to only the homozygous genotypes in the genotype dataset (See Methods Section 4.5 for details). The motivation for using this distance measure is that the extremity based genotype prediction never assigns heterozygous genotypes. Thus the heterozygous genotypes are excluded from distance computation.

## [[FOLLOWING RESULTS CAN BE MOVED TO SUPPLEMENTARY AND CAN BE REPHRASED HERE BRIEFLY.]]

For each measure, the attacker links the predicted genotypes to the individual whose genotypes minimize the selected distance measure. Figure 5c and 5d show the fraction of vulnerable individuals for both distance measures. More than 95% of the individuals are vulnerable for most of the parameter selections for both distance measures. The homozygous genotype matching distance measure has slightly higher linking accuracy. When the gender and/or population information is present as auxiliary information (red and green plots), the fraction of vulnerable individuals increases to 100% for most of the eQTL selections. These results show that linking attack with extremity based genotype prediction, although technically simple, can be extremely effective in characterizing individuals. We will focus on homozygous genotype matching based distance computation in the rest of the paper for simplicity of presentation.

The previous results show that extremity based linking attacks are highly effective when the eQTLs are identified and linking attack is performed using the same expression and genotype datasets. In order to assess the accuracy when the eQTLs are computed and tested on different datasets, we divided the dataset into a training and a testing dataset. The training dataset, of 210 individuals, is used to discover the eQTLs, using Matrix eQTL<sup>52</sup> method (See Methods Section for details). The testing dataset, of 211 individuals, is utilized for assessing the accuracy of linking. Figure 6a shows the linking accuracy for individuals in testing dataset. The accuracy is very high, around 95%, which suggests that extremity based linking attacks are potentially effective when the datasets where eQTLs are identified do not match the data being tested. This is an important aspect of genotype prediction based linking attacks, as they exploit the generalizability of the correlations between phenotypes and genotypes. We also evaluated the accuracy of linking attack in comparison to the linking attack proposed by Schadt et al (See Section S3, Table S2). We observed that the two methods have comparable and very high accuracy, while extremity based linking attack uses much less input information compared to Schadt et al linking attack.

We evaluated whether the attacker can estimate the reliability of the linkings. This may potentially increase the effectiveness of the linking and increase the risk associated with linking attacks because the

attacker can estimate reliability of the linkings and choose the ones that are more likely to be correct. This increases the risk associated with the linking attacks because although he/she may not have a high overall accuracy of linkings, the high ranking linkings may be much higher in accuracy. We observed that the measure we termed, *first distance gap*, denoted by  $d_{1,2}$  (See Methods Section 4.6), serves as a good reliability estimate for each linking. For a given linking,  $d_{1,2}$  is the difference between the genotype distances of the 1<sup>st</sup> closest and 2<sup>nd</sup> closest individuals to the predicted genotypes. When the linking is incorrect, we observed that  $d_{1,2}$  is very likely to be smaller than the distance difference when the linking is correct.

To evaluate this measure further, we computed the positive predictive value (PPV) versus sensitivity of the linkings of individuals in the testing set with changing  $d_{1,2}$  threshold. For this, we first computed  $d_{1,2}$  for each linking, then filtered the linkings that did not satisfy the threshold. Then we computed PPV and sensitivity of the linkings (See Methods Section 4.9), which is plotted in Fig 6b. It can be seen that the PPV of linkings can get very high at the same time with high sensitivity. For example, the attacker can link around 79% of the individuals at a PPV higher than 95%. The random sorting of the linkings, on the other hand, have significantly lower PPV (cyan in the plots) at the same sensitivity levels. These results suggest that the attacker can increase the potential risk (accuracy of linkings) of the attack by focusing on a slightly smaller set of linkings with high reliability.

An important practical question is how well the linking accuracy changes with increasing genotype data size. In order to evaluate this, we simulated the genotypes of the eQTLs (discovered in the training set) for 100,000 individuals. The 100,000 simulated individuals are then merged with the testing dataset of 211 individuals to build the large testing dataset. We then performed the extremity attack using the expression levels of the testing dataset and linked them to the merged testing genotype dataset of 100,211 individuals. The linking accuracy is plotted in Fig 7a with changing eQTL selection criteria. The linking accuracy is very high (Around 96%). This result suggests that the extremity attack can be extended to a large testing sample set. Figure 7b shows the sensitivity versus PPV (with changing first distance gap) for the eQTLs for which the overall linking accuracy is 70% (Yellow dashed lines on Fig. 7b). It can be seen that the attacker can link around 55% of the individuals with PPV higher than 95%. Only the remaining 15% are predicted with accuracy lower than 95%.

We also studied how the linking accuracy changes when the training and testing datasets are measured in different populations. For this, we used the 1000 Genomes Project sample information and divided the GEUVADIS samples into 5 populations. Then we used each population's samples to discover the population specific eQTLs, then used the other populations to test the linking accuracy. Table S1a shows the accuracies in each case. It can be seen that when the eQTLs are discovered in European populations (CEU, GBR, TSI, FIN), the linking accuracies are very high (higher than 95%). When the eQTLs are discovered in YRI (African) population, the linking accuracies are smaller in European populations. Similarly, when eQTLs are discovered on European populations, the linking accuracy in YRI sample is relatively smaller. These results illustrate that extremity attack can still be effective when eQTLs are identified in populations that are genetically close to the population(s) of testing sample and decrease when the discovery and testing populations are diversified. We next studied scenario where the eQTLs are identified in tissues that are different from the tissues on which the expression data is generated.

For this, we used the eQTLs that are identified by GTex Project<sup>41</sup>. We downloaded the eQTLs for 6 tissues and performed the linking attack on the whole GEUVADIS samples as test samples. The results are shown in Table S1b. The accuracy is general high (>80%) and is highest for Whole Blood eQTLs, which is 88%. This is expected since the expression levels in GEUVADIS project are measured in blood cell lines. The accuracy is smallest for Muscle Skeletal eQTLs, which is 76%. It is worth noting that the decrease in the accuracies stem also from the differences in data handling and processing between GEUVADIS and GTex projects.

We also studied whether having close relatives in the genotype dataset affects the accuracy. To test this, we used the expression and genotype data from 30 CEU trios (mother-father-child) available from HAPMAP project<sup>53,54</sup>. We first identified the eQTLs from the 90 individuals and performed linking over the same individuals. We then computed the average rank of the close relatives in each linking. For example, when the tested individual is a father or mother, we computed the rank of his/her child and if the tested individual is a child, we computed the rank of his/her mother and father. We also selected, for each tested individual, a random individual and computed his/her rank in the linking. The distribution of the ranks are shown in Fig 8. It can be seen that the ranks of the related individuals are significantly shifted to smaller values compared to random individuals. This result shows that the close relatives can get linked to each other. This result indicates that the individuals that are close relatives may potentially be confused with each other. While the correct person may not get characterized, the attacker can still reveal sensitive information about the individual's family, which might extend the reach of privacy breach and cause privacy concerns for the family.

### 3 CONCLUSION AND DISCUSSION

Increasing pace of data generation and the policies to encourage genomic data sharing will make genomic privacy a topic of hot debate. In the analysis of genomic privacy, it is necessary to consider the basic premise of sharing any type of personal information: There is always an amount of leakage in the sensitive information<sup>55</sup>. In addition, as shown by previous studies, we often cannot propose black-and-white solutions to problems in privacy which mainly roots from the multifaceted nature of privacy. We believe these make it necessary for the genomic data sharing and publishing mechanisms to incorporate statistical quantification methods before the datasets are released. This is recently recently recognised<sup>56</sup>. Legislative decision making processes should incorporate the quantified risk estimates of leakage as an objective factor. The quantification methodology and the analysis frameworks presented in this study can be applied for analysis of the information leakage in the datasets where the correlative relations between datasets can be exploited for performing linking attacks (Section S5, Figure S8). In accordance to a utility policy, the leakage risk can be evaluated against the utility requirements so as to assess the suitability of different data release mechanisms.

Our study focuses on the individual privacy breaches in the context of linking attacks, where an individual's existence in two seemingly independent databases (e.g., phenotype and the genotype) can cause a privacy concern when an attacker links statistically the databases using the a priori information about correlation of different entries in the databases. The fact that the available molecular phenotypes are (i.e., gene expression levels) generally very high in dimension makes this attack much more

probable. The obvious risk management strategy against these attacks is restricting access to the phenotype datasets. The statistical techniques like k-anonymization and differential privacy can also be utilized. These, however, have associated drawbacks about loss of biological utility, and high computational complexity. Moreover, some studies also demonstrated that there are still risks associated with linkability of the anonymized data<sup>24,30,31,57</sup>. We believe new studies should address protection and risk management strategies for serving utility maximized and privacy aware high dimensional phenotype datasets.

The analysis of tradeoff between predictability and leakage of *ICI* can be generalized in two ways in future studies: First, the information theoretic measures that we proposed for measuring predictability versus the *ICI* leakage can be utilized for analyzing the tradeoff in other biomedical datasets where correlations can be exploited in linking attacks. Second, the analysis that we performed can be used to extrapolate the number of vulnerable individuals at different predictability levels. Depending on the risk of leakage that can be tolerated, the predictability versus *ICI* leakage can be utilized to assess whether the dataset can be released to public access or not. The 3-step framework aims at representing the framework for studying specific instantiations of the linking attacks.

The 3-step decomposition of the attack makes the analysis of different attacks easier as each step can be separately evaluated. For example, the genotype prediction and linking steps can be replaced with different approaches so as to evaluate how the linking accuracy varies. These can reveal insight into how the datasets should be protected. We also presented a simple yet accurate linking attack that utilizes genotype prediction method based on the extremity statistic. This approach capitalizes on the fact that an individual who is an outlier for a phenotype will most likely harbor a homozygous genotype. When employed in the individual identification framework, this simple approach renders a very significant number of individuals vulnerable. In addition, we also showed that the attacker can estimate the reliability of the linkings using the first distance gap statistic so as to increase the risk of correct characterization. This illustrates the viability of individual characterization utilizing technically simple approaches. Even though we observed that the attacker can characterize a large fraction of individuals with high PPV, the smaller fraction of individuals that are linked at the top with high gap distance statistic are under higher risk of being characterized.

Compared to other formalisms, our study aims to develop and build on other studies for quantifying the information leakage and help setup a framework for analysis of the leakage of individual characterizing information. Differential privacy, for example, aims at proposing release mechanisms for statistical databases where the mechanism guarantees that queries return results such that the probability of identifying a specific individual's contribution to the result is vanishingly small. In order to maximize the utility of the biological data, however, it is necessary to analyze the sources of sensitive information leakage so that one can design the utility maximizing release mechanisms<sup>58</sup>. The metrics that we presented can be used to analyze the correlative structures as the leakage sources and quantify the risk and amount of leakage associated with these sources.

## 4 METHODS

### 4.1 Quantification of Individual Characterizing Information and Predictability

The genotype RV  $V_k$  takes 3 different values,  $\{0,1,2\}$ , where the genotype coding is done per counting the number of alternate alleles in the genotype. Given that the genotype is  $g_{k,j}$ , we quantify the individual characterizing information in terms of *self-information*<sup>59</sup> of the event that RV takes the value  $g_{k,j}$ :

$$ICI(V_k = g_{k,j}) = I(V_k = g_{k,j}) = -\log(p(V_k = g_{k,j})) \quad (7)$$

where  $V_k$  is the RV that represents the  $k^{\text{th}}$  eQTL genotype,  $p(V_k = g_{k,j})$  is the probability (frequency) of that  $V_k$  takes the value  $g_{k,j}$ , and  $ICI$  denotes the individual characterizing information. Given multiple eQTL genotypes, assuming that they are independent, the total individual characterizing information is simply summation of those:

$$\begin{aligned} ICI(\{V_1 = v_{1,j}, V_2 = v_{2,j}, \dots, V_N = v_{N,j}\}) \\ = -\sum_{k=1}^N \log(p(V_k = v_{k,j})). \end{aligned} \quad (8)$$

The genotype probabilities are estimated by the frequency of genotypes in the genotype dataset. As presented in the Results Section 2.2, we measure the predictability of eQTL genotypes using an entropy based measure. Given the genotype RV,  $V_k$ , and the correlated gene expression RV,  $E_k$ ,

$$\pi(V_k|E_k = e) = \exp(-H(V_k|E_k = e)) \quad (9)$$

where  $\pi$  denotes the predictability of  $V_k$  given the gene expression level  $e$ , and  $H$  denotes the entropy of  $V_k$  given gene expression level  $e$  for  $E_k$ . The extension to multiple eQTLs is straightforward. For the  $j^{\text{th}}$  individual, given the expression levels  $e_{k,j}$  for all the eQTLs, the total predictability is computed as

$$\begin{aligned} \pi(\{V_k\}, \{E_k = e_{k,j}\}) &= \exp(-H(\{V_k\} | \{E_k = e_{k,j}\})) \\ &= \exp\left(-\sum_k H(V_k|E_k = e_{k,j})\right) \end{aligned} \quad (10)$$

In addition, this measure is guaranteed to be between 0 and 1 such that 0 represents no predictability and 1 representing perfect predictability. The measure can be thought as mapping the prediction process to a uniform random guessing where the average correct prediction probability is measured by  $\pi$ .



## 4.2 Estimation of Genotype Entropy

We estimate the genotype entropy using the Shannon's entropy<sup>59</sup>:

$$H(V_k) = - \sum_{v \in \{0,1,2\}} p(V_k = v) \times \log(p(V_k = v)) \quad (11)$$

where  $V_k$  represents the RV for  $k^{th}$  eQTL variant genotypes and  $p(V_k = v)$  represents the probability that  $V_k$  takes the value  $v$ . This probability can be also interpreted as the population frequency of the genotype  $v$  at the  $k^{th}$  eQTL's variant locus. These probabilities are estimated from the distribution of genotypes over all the samples. As the genotypes are discrete valued, the above formula can be computed in a straightforward way by the summation after the probabilities are estimated.

In the formulation for conditional predictability of genotypes given expression levels, we also use the conditional specific entropies<sup>59</sup> of the genotypes given the gene expression levels. For this, we use the following formulation:

$$H(V_k | E_k = e_{k,j}) = - \sum_{v \in \{0,1,2\}} p(V_k = v | E_k = e_{k,j}) \times \log(p(V_k = v | E_k = e_{k,j})) \quad (12)$$

where  $p(V_k = v | E_k = e_{k,j})$  represents the conditional probability that  $V_k$  takes the value  $v$  under the condition that the RV representing gene expression level for  $k^{th}$  eQTLs ( $E_k$ ) is  $e_{k,j}$ . Since the gene expression levels are continuous, to estimate the conditional probabilities of genotypes given expression levels; we start with the joint distribution of  $E_k$  and  $V_k$ , then bin the gene expression levels. For this, we use Sturges' rule<sup>60</sup> to choose the number of bins. This rule states that the number of bins should be selected as:

$$n_b = \lceil \log_2(n_e) \rceil + 1 = \lceil \log_2(421) \rceil + 1 = 10 \quad (13)$$

The binning is done for each gene by first sorting the expression levels for all the individuals, then the range of gene expression levels are divided into  $n_b = 10$  bins of equal size and each expression level is mapped to a value between in  $[0, n_b - 1]$ . The expression level of  $k^{th}$  gene in  $j^{th}$  individual,  $e_{k,j}$ , is mapped to

$$\tilde{e}_{k,j} = \left\lfloor \frac{(e_{k,j} - \min(\mathbf{e}_k)) \times n_b}{\max(\mathbf{e}_k) - \min(\mathbf{e}_k)} \right\rfloor \quad (14)$$

where  $\min(\mathbf{e}_k)$  and  $\max(\mathbf{e}_k)$  represents the minimum and maximum values, respectively, for the  $k^{th}$  expression level over all the samples and  $\tilde{e}_{k,j}$  represents the binned expression level. After the gene expression levels are binned, we use the binned expression levels and compute the conditional distribution of the variant genotypes at each binned gene expression level using the histograms:

$$p(V_k = v | \tilde{E}_k = \tilde{e}_{k,j}) = \frac{\sum_i I(\tilde{e}_{k,i} = \tilde{e}_{k,j}, V_{k,i} = v)}{\sum_i I(\tilde{e}_{k,i} = \tilde{e}_{k,j})} \quad (15)$$

where  $I(\cdot)$  is an indicator function for counting the number of matching mapped expression and genotype values:

$$I(\tilde{e}_{k,i} = \tilde{e}_{k,j}, V_{k,i} = v) = \begin{cases} 1; & \text{if } \tilde{e}_{k,i} = \tilde{e}_{k,j}, V_{k,i} = v \\ 0; & \text{otherwise} \end{cases} \quad (16)$$

Finally, we utilize compute the Shannon entropy of the estimated conditional distribution as the condition specific entropies.

### 4.3 Maximum *a posteriori* (MAP) Genotype Prediction

While assigning the genotypes using maximum *a posteriori* prediction, the attacker assigns to  $V_k$  the genotype that maximizes the estimated conditional probability:

$$\text{MAP}(V_k | \tilde{E}_k = \tilde{e}_{k,j}) = \tilde{v}_{k,j} = \underset{v}{\text{argmax}}(p(V_k = v | \tilde{E}_k = \tilde{e}_{k,j})) \quad (17)$$

where the conditional probabilities are estimated as in Methods Section 4.2 and  $\tilde{v}_{k,j}$  denotes the predicted genotype for  $V_k$ , given  $\tilde{E}_k = \tilde{e}_{k,j}$ .

### 4.4 Linking of the Predicted Genotypes to Genotype Dataset

The linking is the 3<sup>rd</sup> and last step of the linking attack. The aim is to compare the predicted genotypes from the phenotype dataset to the genotypes in the genotype dataset so as to match the samples in the phenotype dataset to those in genotype dataset. We will use the linking approach that evaluates the minimal distance between the compared genotypes but different methods can be used for genotype comparison. Given a set of predicted eQTL genotypes for individual  $j$ ,  $\tilde{\mathbf{v}}_{\cdot,j} = \{\tilde{v}_{1,j}, \tilde{v}_{2,j}, \dots, \tilde{v}_{n_q,j}\}$ , the attacker links the predicted genotypes to the individual whose genotypes have the smallest distance to the predicted genotypes:

$$\text{pred}_j = \underset{a}{\text{argmin}}\{d(\tilde{\mathbf{v}}_{\cdot,j}, \mathbf{v}_{\cdot,a})\}. \quad (18)$$

$\text{pred}_j$  denotes the index for the linked individual and  $d(\tilde{\mathbf{v}}_{\cdot,j}, \mathbf{v}_{\cdot,a})$  represents the distance between the predicted eQTL genotypes and the genotypes of the  $a^{\text{th}}$  individual:

$$d(\tilde{\mathbf{v}}_{\cdot,j}, \mathbf{v}_{\cdot,a}) = \sum_{k=1}^{n_q} (1 - I(\tilde{v}_{k,j}, v_{k,a})) \quad (19)$$

where  $I(\tilde{v}_{k,j}, v_{k,a})$  is the match indicator:

$$I(\tilde{v}_{k,j}, v_{k,a}) = \begin{cases} 1 & \text{if } \tilde{v}_{k,j} = v_{k,a} \\ 0 & \text{otherwise} \end{cases} \quad (20)$$

Finally,  $j^{\text{th}}$  individual is vulnerable if  $\text{pred}_j = j$ . When auxiliary information is available, the attacker constrains the set of individuals while computing  $d(\tilde{\mathbf{v}}_{\cdot,j}, \mathbf{v}_{\cdot,a})$  to the individuals with matching auxiliary information. For example, if the gender of the individual is known, the attacker excludes the individuals

whose gender does not match while computing  $d(\tilde{\mathbf{v}}_{\cdot,j}, \mathbf{v}_{\cdot,a})$ . This way the auxiliary information decreases the search space of the attacker.

#### 4.5 Homozygous Genotype Matching based Linking of the Predicted Genotypes to Genotype Dataset

The extremity based genotype prediction predicts only homozygous genotypes. Therefore heterozygous genotypes in the genotype dataset will always increase the distance in linking step. To correct for this, the attacker can focus only on the homozygous genotypes while he/she is linking the predicted genotypes to the genotype dataset. For this, a simple modification of the distance function is sufficient:

$$d^H(\tilde{\mathbf{v}}_{\cdot,j}, \mathbf{v}_{\cdot,a}) = \frac{\sum_{k=1}^{n_q} (1 - I^H(\tilde{v}_{k,j}, v_{k,a}))}{n_a^H} \quad (21)$$

where  $n_a^H$  represents the number of homozygous genotypes in  $\mathbf{a}$ th individual and  $I^H(\tilde{v}_{k,j}, v_{k,j})$  represents the homozygous match indicator:

$$I^H(\tilde{v}_{k,j}, v_{k,j}) = \begin{cases} 1 & \text{if } v_{k,a} = 0, \tilde{v}_{k,j} = v_{k,a} \\ 1 & \text{if } v_{k,a} = 2, \tilde{v}_{k,j} = v_{k,a} \\ 1 & \text{if } v_{k,a} = 1 \\ 0 & \text{otherwise} \end{cases} \quad (22)$$

This indicator function does comparison only when the genotype being matched ( $v_{k,a}$ ) is homozygous. When  $v_{k,a}$  is heterozygous, it acts as if the genotypes are the same, thus the distance function is updated only when the genotype being matched is a homozygous genotype. The normalization is necessary to convert the distance into a fraction so that the distances can be compared among different genotype samples.

#### 4.6 First Distance Gap Statistic For Reliability Estimation of Each Linking

Following the previous section, the attacker computes, for each individual, the distance to all the genotypes in genotype dataset, then identifies the individual with smallest distance. Let  $d_{j,(1)}$  and  $d_{j,(2)}$  denote the minimum and second minimum genotype distances (among  $d^H(\tilde{\mathbf{v}}_{\cdot,j}, \mathbf{v}_{\cdot,a})$  for all  $\mathbf{a}$ ) for  $j^{\text{th}}$  individual. We propose using the difference between these distances, termed *first distance gap statistic*, as a measure of reliability of linking. For this, the attacker computes following difference:

$$d_{1,2}(j) = d_{j,(2)} - d_{j,(1)} \quad (23)$$

First distance gap can be computed without the knowledge of the true genotypes, and is immediately accessible by the attacker with no need for auxiliary information. The basic motivation for this statistic comes from the observation that the first distance gap for correctly linked individuals are much higher compared to the incorrectly linked individuals (See Figure S5).

#### 4.7 eQTL Identification on Training Sets with Matrix eQTL<sup>52</sup>

For identification of eQTLs, we used Matrix eQTL<sup>52</sup> method. We first generated the testing and training sample lists by randomly picking 210 and 211 individuals, respectively, for testing and training sets. We

then separated the genotype and expression matrices into training and testing sets. Matrix eQTL is run to identify the eQTLs using the training dataset. In order to decrease the run time, Matrix eQTL is run in cis-eQTL identification mode. After the eQTLs are generated, we filtered out the eQTLs whose FDR (as reported by Matrix eQTL) was larger than 5%. We finally removed the redundancy by ensuring that each gene and each SNP is used only once in the eQTL final list. To accomplish this, we selected the eQTL that is correlated with highest association with each gene. The association statistic reported by Matrix eQTL was used as the measure of strength of association between expression levels and genotypes. Similar procedure is applied when eQTLs for 30 trios are identified.

#### 4.8 On Modeling of Genotype-Phenotype Distribution for Genotype Prediction in Linking Attacks

In the second step of the linking attack, the genotype predictions are performed. The genotype predictions are used, as an intermediate information, as input to the third step (Fig 3), where linking is performed. The main aim of attacker is to maximize the linking accuracy (not the genotype prediction accuracy), which depends jointly on the genotype prediction accuracy and the accuracy of the genotype matching in the 3<sup>rd</sup> step. Other than the accuracy of linking, another important consideration, for risk management purposes, is the amount of auxiliary input data (like training data for prediction model) that the genotype prediction takes. The prediction methods that require high amount of auxiliary data would decrease the applicability of the linking attack as the attacker would need to gather extra information before performing the attack. On the other hand, the prediction methods that require little or no auxiliary data makes the linking attack much more realistic and prevalent. It is therefore useful, in the risk management strategies, to study complexities of genotype prediction methods and evaluate how these translate into assessing the accuracy and applicability of the linking attack. We study different simplifications of genotype prediction, and illustrate different levels of complexity for genotype prediction.

As we presented in Section 2.3, we assume that the attacker estimates the posterior distribution of genotypes and utilizes the maximum *a posteriori* estimate of the genotype as the general prediction method. For this, attacker must first model the joint genotype-phenotype distribution and then build the posterior genotype distribution. Figure S9a shows the joint genotype-expression distribution for an eQTL. Figure S9b shows the modeling of the joint distribution using 3 conditional distributions of expression levels at each genotype. First, the means and variances of the distributions are assumed independent. Assuming that mean and variance are sufficient statistics for the conditional distributions (e.g., normally distributed), the joint distributions can be modeled when the 6 parameters (3 means and 3 variances) are trained. The training can be performed using unsupervised methods like expectation maximization or can be performed using training data. This would, however, increase the required auxiliary data and decrease the applicability of the linking attack. Figure S9c shows a simplification of the model by assuming the variances of the conditional expression distributions are same for each genotype. This decreases the number of parameters to be trained to 4 (3 means and 1 variance). Figure S9d shows an equally complex model with 4 parameters where the conditional distributions are uniform at non-overlapping ranges of expression for each genotype. This model requires 4 parameters to be trained corresponding to the expression range limits. Figure S9e shows the final simplification of the

genotype prediction, which requires only one parameter to be trained. In this model, the prediction only assigns uniform probability for homozygous genotypes when expression levels higher or lower than  $e_{mid}$  and assigns 0 conditional probability to the heterozygous genotypes, which brings up an important point: This simplified model is exactly the distribution that is utilized in the extremity based genotype prediction. In the extremity based prediction, we estimate  $e_{mid}$  simply as the mid-point of the range of gene expression levels within the expression dataset (Equations 3 and 4-6).

#### 4.9 Accuracy, Positive Predictive Value, and Sensitivity of Linkings

The accuracy of the linkings are evaluated with respect to 3 measures. First is the overall accuracy of linkings, which measures the fraction of correctly linked individuals. This is simply among the all the linkings that we performed, the fraction of correct linkings:

$$\text{Overall Accuracy} = \frac{\sum_{j \in (1, n_e)} I(\text{pred}_j = j)}{n_e} \quad (24)$$

where  $\text{pred}_j$  is defined in Equation (18) and  $I(\text{pred}_j = j)$  returns 1 if  $\text{pred}_j$  is equal to  $j$  (i.e., correct linking) and 0 otherwise. When we are studying the effectiveness of first distance gap,  $d_{1,2}$ , for estimating reliability of linkings, we select the subset of linkings for which  $d_{1,2}$  is above the first distance gap threshold and computed the sensitivity and positive predictive value (PPV) for the selected set of linkings, which are defined below:

$$\text{Sensitivity for } d_{1,2} \text{ threshold } \tau = \frac{\sum_{j \in (1, n_e)} I(\text{pred}_j = j, d_{1,2}(j) > \tau)}{n_e} \quad (25)$$

$$\text{PPV for } d_{1,2} \text{ threshold } \tau = \frac{\sum_{j \in (1, n_e)} I(\text{pred}_j = j, d_{1,2}(j) > \tau)}{n_\tau} \quad (26)$$

where  $n_\tau$  represents the number of individuals for which the first distance gap for the linking is greater than the threshold  $\tau$ ,

$$n_\tau = \sum_{j \in (1, n_e)} I(d_{1,2}(j) > \tau) \quad (27)$$

In summary, sensitivity measures the fraction of correctly linked individuals among all the  $n_e$  individuals and PPV measures the fraction of correctly linked individuals among the selected  $n_\tau$  individuals, for which the first distance gap for linking is greater than  $\tau$ . When  $\tau$  is increased,  $n_\tau$  will decrease (Less number of linkings will be higher than  $\tau$ ), the reliabilities of the linking will increase and PPV will increase. Sensitivity, however, decreases as we select smaller number of individuals. When we are evaluating the random sortings of the data, we first randomly sorted the linkings but did not shuffle the first distance gap values. Then we computed the accuracies and plotted the sensitivity vs PPV curves.

## 5 DATASETS

The normalized gene expression levels for 462 individuals and the eQTL dataset are obtained from gEUVADIS mRNA sequencing project<sup>61</sup>. The eQTL dataset contains all the significant (Identified at most 5% false discovery rate) gene-variant pairs with high genotype-expression correlation. To ensure that there are no dependencies between the variant genotypes and expression levels, we used the eQTL entries where gene and variants are unique. In other words, each variant and gene are found exactly once in the final eQTL dataset (See Section S4 for a detailed discussion). The genotype, gender, and population information datasets for 1092 individuals are obtained from 1000 Genomes Project<sup>12</sup>. For 421 individuals, both the genotype data and gene expression levels are available. For tissue analysis, the publicly available significant eQTLs for 6 tissues that are computed by the GTex project are downloaded from the GTex Portal.

## 6 ACKNOWLEDGEMENTS

Authors would like to thank Akdes Serin Harmanci for constructive comments on study design and running of external tools. Authors also would like to thank the anonymous reviewers for constructive criticism that made the manuscript complete and comprehensive.

## 7 AUTHOR CONTRIBUTIONS

AH designed the study, gathered datasets, performed experiments, and drafted the manuscript. MG conceived the study, oversaw the experiments and wrote the manuscript. Both authors

Authors declare no conflict of interest.

## 8 FIGURE CAPTIONS

**Figure 1:** Illustration of the linking attack. (a) Phenotype dataset contains  $q$  different phenotype measurements and the HIV Status for a list of individuals. Genotype dataset contains the variants genotypes for  $m$  individuals. Phenotype-Genotype correlation datasets contains  $q$  phenotypes, variants, and their correlations. The attacker does genotype prediction for all the variants. The attacker then links the phenotype dataset to the genotype dataset by matching the genotypes. The linking potentially reveals the HIV status for the subjects in the genotypes dataset. The IDs and HIV Status are colored to illustrate how the linking combines the entries in the two datasets. The non-shaded columns are used for linking. (b) Illustration of the expression and genotype datasets. Variant genotype dataset contains the genotypes for  $q$  eQTL variants for  $n_v$  individuals.  $j^{th}$  entry for  $k^{th}$  eQTL is denoted by  $v_{k,j}$ . Similarly, the expression dataset contains the expression levels for  $q$  genes. The  $k^{th}$  expression level for  $j^{th}$  individual is denoted by  $e_{k,j}$ . The variant genotypes for  $k^{th}$  variant is distributed over samples with distribution specified by the random variable  $V_k$ . Likewise, the expression levels for  $k^{th}$  gene is distributed per random variable  $E_k$ . These random variables are correlated with each other with correlation coefficient, denoted by  $\rho(E_k, V_k)$  (right).

**Figure 2:** Quantification of ICI and correct genotype predictability (a) Adversary’s genotype prediction strategy. The phenotype-genotype correlations  $\rho_1, \rho_2, \dots$  are sorted with respect to decreasing absolute value, as shown on each line. For a selected set of  $n$  variants, the genotypes are predicted using the phenotypes. The green and red individuals on the right represent the vulnerable and non-vulnerable individuals, respectively. (b) Plots show, for each eQTL, the information leakage (x-axis) versus correct genotype predictability (y-axis). For each eQTL, the estimated ICI leakage and genotype predictability are plotted. The dots are colored with respect to the major allele frequency (left) and with respect to absolute correlation of the eQTL (right). (c) Average predictability versus average individual characterizing information leakage. For the top 20 eQTLs, the plot shows the distribution of average predictability and average ICI leakage for the top eQTLs. The number of eQTLs that are used for computing the values at each point are shown next to the point. Only 10 of them are numbered in the figure. The error bars show one standard deviation around the mean among the sample set. The cyan plot shows the mean leakage plot for shuffled gene-variant pairs. The error bars are left out for simplification.

**Figure 3:** The figure illustrates three steps of the linking attack. The first step consists of selecting the phenotypes and genotype to be used in linking. The absolute value of correlation can be used as one of the selection criteria. The second step comprises the genotype prediction using the selected set of phenotypes. Maximum *a posteriori* genotype prediction can be used for prediction. Third step in characterization is the linking step, where the predicted genotypes are matched to the genotype dataset. The matching can be performed by comparing the distance between the predicted genotypes and individual genotypes in the dataset.

**Figure 4:** MAP genotype prediction accuracy and vulnerable fraction. (a) The number of selected eQTLs (blue) and the number of eQTLs with correctly predicted eQTL (red) are shown. At each absolute correlation threshold, the number of eQTLs passing the threshold are shown and the number of correctly predicted genotypes using MAP prediction are shown. The error bars show the distribution of accuracy over all the samples. (b) The fraction of vulnerable individuals with MAP genotype prediction. X-axis shows the absolute correlation threshold used to select eQTLs. Y-axis shows the fraction of vulnerable individuals. At correlation threshold of 0.35, the fraction is maximized, as indicated by the yellow arrow. The red, green, and cyan lines show the fraction of vulnerable individuals when gender, population, and gender and population information, respectively, are available as auxiliary information.

**Figure 5:** Extremity based genotype prediction and extremity based linking attack (a) Figure illustrates the extremity based genotype prediction. The joint distribution of expression levels and genotypes is shown on left. Given the relation between expression and genotypes, the lower expression levels (Labelled with “Negative Extremity” shown in red ellipse on left) are assigned the genotype “TT” and higher expression levels (Labelled with “Positive Extremity” shown in green ellipse on left) are assigned the genotype “CC”. The extremity value 0 is indicated with the dashed grey line which is the midpoint between the highest and lowest expression levels. The heterozygous genotype (TC) is shaded with grey to indicate that it is not used in the genotype prediction. (b) The extremity based genotype prediction accuracy versus the absolute correlation threshold used to select the eQTLs. (c) The fraction of vulnerable individuals versus the correlation threshold in blue. The red, green, and cyan plots show the

vulnerable fraction when gender, population, and gender + population are available, respectively, as auxiliary information. (d) The vulnerable individual fraction with homozygous only matching.

**Figure 6:** The accuracy of linking on the testing dataset. (a) The fraction of vulnerable individuals on the testing dataset. (b) The sensitivity versus positive predictive value (PPV) of linking with changing  $d_{1,2}$  threshold. The plot is generated with eQTLs with higher than 11.3 absolute association strength (indicated with yellow dashed lines) strength. Blue line shows the ranking with  $d_{1,2}$  and cyan lines shows PPV and sensitivity with 10 random rankings for comparison. The grey lines indicate 95% PPV, at which approximately 80% sensitivity is achieved.

**Figure 7:** The vulnerable individual fraction statistics for simulated genotypes dataset. (7a) The fraction of vulnerable individuals. (7b) The sensitivity versus positive predictive value. The plot is for the eQTL selection at association strength 9.9, as indicated by the yellow dashed line in (a), where the vulnerable fraction is around 70%. The dashed grey line show the 95% PPV level, where the sensitivity is around 55%. The cyan plots show the sensitivity-PPV plot for 10 randomized rankings of the data for comparison.

**Figure 8:** The distribution of linking ranks (in genotype comparison) of close relatives in 30 trios from the HAPMAP project. The blue plot shows the distribution of ranks assigned to close relatives (child-mother or child-father relations) excluding the self ranks of the individual being linked. Red plot shows, as a control, the same distribution for randomly selected individuals for each individual being linked. The ranks for close relatives are much smaller, indicating that they are scored much higher than random individuals.

**Table S1:** Linking accuracy of extremity based linking attack using the eQTLs are identified in different populations and different tissues. (a) The table shows the linking accuracies (for populations shown in the rows) when the eQTLs that are identified using data (indicated in each column) from different populations. (b) The linking accuracy of individuals in GEUVADIS project when eQTLs identified from different tissues are used in linking.

**Table S2:** Linking attack accuracy comparison. The table shows linking accuracy for Schadt et al and extremity based linking attack methods. Each row corresponds (for Schadt et al Method) to a different number of data points in the training datasets that is input to Schadt et al method.

**Figure S1:** Different properties of the eQTLs. (a) The average ICI leakage versus the genotype predictability is shown for real (red) and shuffled (blue) eQTL dataset is shown. (b) The absolute correlation versus predictability is shown.

**Figure S2:** Figure shows the attacker's presumed strategy for linking attack. (a) The phenotype and variant pairs are sorted with respect to decreasing absolute correlations values. For the top  $n$  pairs, joint predictability and ICI are computed. (b) Illustration of prior, joint, and posterior distributions of genotypes and expression levels. Leftmost figure shows the distribution of genotypes over the sample set, which is labelled as the prior distribution. Middle figure shows the joint distribution of genotypes and expression levels. Notice that there is a significant negative correlation between genotype values



and the expression levels. Rightmost figure shows the posterior distribution of genotypes given that the gene expression level is 10. The posterior distribution has a maximum (MAP prediction) at genotype 2, which is indicated by a star.

**Figure S3:** The distribution of ranks of the individuals in the linking step. At each gradient threshold, the box plots show, for each individual, their ranks in the genotype comparison in the 3<sup>rd</sup> step of linking attack with MAP genotype prediction. Notice that at around 0.35 correlation threshold, the assigned ranks are minimized, i.e., most of the individual are linked correctly.

**Figure S4:** The median absolute gene expression extremity statistics over 462 individuals in GEUVADIS dataset. (a) For each individual, the extremity is computed over all the genes (23,662 genes) reported in the expression dataset. The median of the absolute value of the extremity is plotted. X-axis shows the sample index and y-axis shows the extremity. The absolute median extremity fluctuates around 0.25, which is exactly the midpoint between minimum and maximum values of absolute extremity. (b) For each individual, we count the number of genes above the extremity threshold. The plot shows the extremity threshold versus the median number of genes (over 462 individuals) above the extremity threshold. Around half of the genes (indicated by dashed yellow lines) have higher than almost 0.3 extremity on average over all the individuals. Also, around median number of 1000 genes over the samples have higher than 0.45 extremity (indicated by dashed red lines).

**Figure S5:** Illustration of linking for  $j^{th}$  individual. The attacker first predicts the genotypes ( $\tilde{v}_{\cdot,j}$ ) which are then used to compute the distance to all the individuals in the genotype dataset. The computed distances are then sorted in decreasing. The top matching individual (in the example, individual  $a$ ) is assigned as the linked individual. The first distance gap,  $d_{1,2}$ , is computed as the difference between the second ( $d_{j,(2)}$ ) and the first ( $d_{j,(1)}$ ) distances in the sorted list.

**Figure S6:** Schematic comparison of linking attacks (Left) and detection of a genome in a mixture attacks (Right). Each box in the figure represents a dataset in the form of a matrix. Multiple boxes next to each other correspond to concatenation of matrices. Linking attacks aim at linking genotype and phenotype datasets. The phenotype datasets contain both “predicting” phenotypes and other phenotypes, some of which can be sensitive. The attacker first predict genotypes for each of the predicting phenotype. The predicted genotypes are then compared with the genotypes in the genotype dataset. After the linking, all the datasets are concatenated where the identifiers can be matched to the sensitive phenotypes. Different colors indicate how the linking merges different information. The detection of a genome in a mixture attacks start with a genotype dataset. The attacker gets access to the statistics of a GWAS or genotyping dataset (for example, regression coefficients or allele frequencies). Then the attacker generates a statistic and tests it against that of a reference population. The testing result can be converted into the study membership indicator (attended/not attended) which shows whether the tested individual was in the study cohort or not.

**Figure S7:** A representative example of extremity based linking. The phenotype dataset (Consisting of gene expression levels for 6 genes) is shown above. Each phenotype measurement is represented by green (negative extreme), red (positive extreme), or grey (non-extreme) dots. Based on the extremity of

phenotypes, the attacker performs prediction of genotypes, which are shown below in (2). He/she uses the eQTL dataset (with genes and SNPs) for prediction. Blue and brown triangles correspond to the correct genotype predictions. The grey crosses correspond to the incorrect or unavailable genotype predictions. The attacker compares the predicted genotypes to the genotype dataset in (3), where triangles show the genotypes, and performs linking. The attacker links the predicted genotypes to the genotype dataset. 3 individuals (Bob, Alice, and John) are highlighted. The attacker can link Bob and John by matching them to their genotypes. The correct prediction of rs7274244 (in yellow dashed rectangle) enables the attacker to distinguish between correct entries and reveal both of their disease status as positive. For Alice, the predicted genotypes are equally matching at two entries both of which match at 2 genotypes; PID-b and PID-k (with negative and positive disease status) thus the attacker cannot exactly reveal Alice's disease status.

**Figure S8:** Illustration of risk assessment procedure for joint genotyping/phenotyping data generation. There are two paths of risk assessment to be performed. The first path evaluates the risks associated with release of the QTL datasets. The genotype and phenotype data (on the left) is first used for quantitative trait loci identification (QTL identification box). This generates the significant QTLs. These are then utilized, in addition to the list of external QTL databases, in quantification of leakage versus predictability, as presented in Section 2.2. These results are then relayed to the risk assessment procedures. The second risk assessment procedure evaluates the release of genotype and phenotype datasets. For this, the datasets are input to application of a list of linking attacks (Presented in Sections 2.3, and 2.4, and other linking attacks in the literature) for evaluation of characterization risks. The results are then relayed to risk assessment procedures.

**Figure S9:** Models of joint genotype-expression distribution with varying numbers of parameters for a positively correlated eQTL. (a) shows the true distribution where grey boxes represent the expression distributions given different genotypes. Red line show the gradient of correlation between genotype and expression. First simplification of the model is shown in (b). The expression distribution can be modeled with Gaussians with different means and variances with total of 6 parameters. The variances can be assumed same for different genotypes (c), where 4 parameters are required. (d) illustrates a representation of the uniform expression distribution given genotypes, where 4 parameters are required. The conditional distribution of expression is uniform (cross shaded rectangles) over the ranges  $(e_1, e_2)$ ,  $(e_2, e_3)$ , and  $(e_3, e_4)$  given genotypes 0, 1, and 2, respectively. The transparent grey rectangles shows the original distributions. (e) is a simplification of (d) where no conditional probability of expression is assigned given genotype is 1. In this model, only 1 parameter ( $e_{mid}$ ) is necessary. The conditional probability of expression given genotypes 0 and 2 are uniform for expression levels below  $e_{mid}$  and above  $e_{mid}$ , respectively (shown with cross shaded rectangles). The original distribution is included with grey rectangles for comparison. Extremity based prediction is an instantiation of the model in (e).

## 9 REFERENCES

1. Sboner, A., Mu, X., Greenbaum, D., Auerbach, R. K. & Gerstein, M. B. The real cost of sequencing: higher than you think! *Genome Biol.* **12**, 125 (2011).
2. Rodriguez, L. L., Brooks, L. D., Greenberg, J. H. & Green, E. D. The Complexities of Genomic Identifiability. *Science (80-. )*. **339**, 275–276 (2013).
3. Sweeney, L., Abu, A. & Winn, J. Identifying Participants in the Personal Genome Project by Name. *SSRN Electron. J.* 1–4 (2013). doi:10.2139/ssrn.2257732
4. infographic-printable.pdf. at <<http://www.nih.gov/precisionmedicine/infographic-printable.pdf>>
5. Collins, F. S. A New Initiative on Precision Medicine. *N. Engl. J. Med.* **372**, 793–795 (2015).
6. Plan for Increasing Access to Scientific Publications - NIH-Public-Access-Plan.pdf. at <<https://grants.nih.gov/grants/NIH-Public-Access-Plan.pdf>>
7. GENOMIC DATA SHARING (GDS) Home. at <<http://gds.nih.gov/index.html>>
8. Sweeney, L. *Uniqueness of Simple Demographics in the U.S. Population, LIDAP-WP4. Forthcom. B. entitled, Identifiability Data.* (2000).
9. Golle, P. Revisiting the uniqueness of simple demographics in the US population. in *Proc. 5th ACM Work. Priv. Electron. Soc.* 77–80 (2006). doi:<http://doi.acm.org/10.1145/1179601.1179615>
10. Consortium, T. G. The Genotype-Tissue Expression (GTEx) project. *Nat. Genet.* **45**, 580–5 (2013).
11. Bernstein, B. E. *et al.* An integrated encyclopedia of DNA elements in the human genome. *Nature* **489**, 57–74 (2012).
12. The 1000 Genomes Project Consortium. An integrated map of genetic variation. *Nature* **135**, 0–9 (2012).
13. Collins, F. S. The Cancer Genome Atlas (TCGA). *Online* 1–17 (2007).
14. Pakstis, A. J. *et al.* SNPs for a universal individual identification panel. *Hum. Genet.* **127**, 315–324 (2010).
15. Wei, Y. L., Li, C. X., Jia, J., Hu, L. & Liu, Y. Forensic Identification Using a Multiplex Assay of 47 SNPs. *J. Forensic Sci.* **57**, 1448–1456 (2012).
16. Church, G. *et al.* Public access to genome-wide data: Five views on balancing research with privacy and protection. *PLoS Genet.* **5**, (2009).
17. Lunshof, J. E., Chadwick, R., Vorhaus, D. B. & Church, G. M. From genetic privacy to open consent. *Nat. Rev. Genet.* **9**, 406–411 (2008).

18. Homer, N. *et al.* Resolving individuals contributing trace amounts of DNA to highly complex mixtures using high-density SNP genotyping microarrays. *PLoS Genet.* **4**, (2008).
19. Im, H. K., Gamazon, E. R., Nicolae, D. L. & Cox, N. J. On sharing quantitative trait GWAS results in an era of multiple-omics data and the limits of genomic privacy. *Am. J. Hum. Genet.* **90**, 591–598 (2012).
20. Narayanan, A. & Shmatikov, V. Robust de-anonymization of large sparse datasets. in *Proc. - IEEE Symp. Secur. Priv.* 111–125 (2008). doi:10.1109/SP.2008.33
21. Gymrek, M., McGuire, A. L., Golan, D., Halperin, E. & Erlich, Y. Identifying personal genomes by surname inference. *Science* **339**, 321–4 (2013).
22. Erlich, Y. & Narayanan, A. Routes for breaching and protecting genetic privacy. *Nat. Rev. Genet.* **15**, 409–21 (2014).
23. Dwork, C. Differential privacy. *Int. Colloq. Autom. Lang. Program.* **4052**, 1–12 (2006).
24. Fredrikson, M., Lantz, E., Jha, S. & Lin, S. Privacy in Pharmacogenetics: An End-to-End Case Study of Personalized Warfarin Dosing. in *23rd USENIX Secur. Symp.* (2014). at <<http://www.biostat.wisc.edu/~page/WarfarinUsenix2014.pdf>>
25. Adam, N. R. & Worthmann, J. C. Security-control methods for statistical databases: a comparative study. *ACM Comput. Surv.* **21**, 515–556 (1989).
26. Gentry, C. A FULLY HOMOMORPHIC ENCRYPTION SCHEME. *PhD Thesis* 1–209 (2009). doi:10.1145/1536414.1536440
27. SWEENEY, L. k-ANONYMITY: A MODEL FOR PROTECTING PRIVACY. *Int. J. Uncertainty, Fuzziness Knowledge-Based Syst.* **10**, 557–570 (2002).
28. Loukides, G., Gkoulalas-Divanis, A. & Malin, B. Anonymization of electronic medical records for validating genome-wide association studies. *Proc. Natl. Acad. Sci. U. S. A.* **107**, 7898–7903 (2010).
29. Meyerson, A. & Williams, R. On the complexity of optimal K-anonymity. in *Proc. twentythird ACM SIGMOD-SIGACT-SIGART Symp. Princ. database Syst. Pod. 04* 223–228 (2004). doi:10.1145/1055558.1055591
30. Machanavajjhala, A., Kifer, D., Gehrke, J. & Venkatasubramanian, M. L-diversity. *ACM Trans. Knowl. Discov. Data* **1**, 3–es (2007).
31. Ninghui, L., Tiancheng, L. & Venkatasubramanian, S. t-Closeness: Privacy beyond k-anonymity and  $\ell$ -diversity. in *Proc. - Int. Conf. Data Eng.* 106–115 (2007). doi:10.1109/ICDE.2007.367856
32. Holdt, L. M. *et al.* Quantitative trait loci mapping of the mouse plasma proteome (pQTL). *Genetics* **193**, 601–608 (2013).

33. Stark, A. L. *et al.* Protein Quantitative Trait Loci Identify Novel Candidates Modulating Cellular Response to Chemotherapy. *PLoS Genet.* **10**, (2014).
34. Degner, J. F. *et al.* DNase I sensitivity QTLs are a major determinant of human expression variation. *Nature* **482**, 390–394 (2012).
35. Battle, A. *et al.* Impact of regulatory variation from RNA to protein. *Science (80-. )*. **347**, 664–667 (2014).
36. Bell, J. T. *et al.* DNA methylation patterns associate with genetic and gene expression variation in HapMap cell lines. *Genome Biol.* **12**, R10 (2011).
37. McVicker, G. *et al.* Identification of genetic variants that affect histone modifications in human cells. *Sci. (New York, NY)* **342**, 747–749 (2013).
38. Kilpinen, H. *et al.* Coordinated effects of sequence variation on DNA binding, chromatin structure, and transcription. *Science* **342**, 744–7 (2013).
39. Kasowski, M. *et al.* Extensive variation in chromatin states across humans. *Sci. (New York, NY)* **342**, 750–752 (2013).
40. Pickrell, J. K. *et al.* Understanding mechanisms underlying human gene expression variation with RNA sequencing. *Nature* **464**, 768–772 (2010).
41. Ardlie, K. G. *et al.* The Genotype-Tissue Expression (GTEx) pilot analysis: Multitissue gene regulation in humans. *Science (80-. )*. **348**, 648–660 (2015).
42. Speliotes, E. K. *et al.* Association analyses of 249,796 individuals reveal 18 new loci associated with body mass index. *Nat. Genet.* **42**, 937–948 (2010).
43. Cheverud, J. M. *et al.* Quantitative trait loci for obesity- and diabetes-related traits and their dietary responses to high-fat feeding in LGXSM recombinant inbred mouse strains. *Diabetes* **53**, 3328–3336 (2004).
44. Beekman, M. *et al.* Evidence for a QTL on chromosome 19 influencing LDL cholesterol levels in the general population. *Eur. J. Hum. Genet.* **11**, 845–850 (2003).
45. Stranger, B. E. *et al.* Patterns of Cis regulatory variation in diverse human populations. *PLoS Genet.* **8**, (2012).
46. Montgomery, S. B. *et al.* Transcriptome genetics using second generation sequencing in a Caucasian population. *Nature* **464**, 773–777 (2010).
47. Xia, K. *et al.* SeeQTL: A searchable database for human eQTLs. *Bioinformatics* **28**, 451–452 (2012).
48. Schadt, E. E., Woo, S. & Hao, K. Bayesian method to predict individual SNP genotypes from gene expression data. *Nat. Genet.* **44**, 603–608 (2012).

49. Trabzuni, D. *et al.* Widespread sex differences in gene expression and splicing in the adult human brain. *Nat. Commun.* **4**, 2771 (2013).
50. Spielman, R. S. *et al.* Common genetic variants account for differences in gene expression among ethnic groups. *Nat. Genet.* **39**, 226–231 (2007).
51. Storey, J. D. *et al.* Gene-expression variation within and among human populations. *Am. J. Hum. Genet.* **80**, 502–509 (2007).
52. Shabalin, A. A. Matrix eQTL: Ultra fast eQTL analysis via large matrix operations. *Bioinformatics* **28**, 1353–1358 (2012).
53. Stranger, B. E. *et al.* Relative impact of nucleotide and copy number variation on gene expression phenotypes. *Science* **315**, 848–853 (2007).
54. The International HapMap 3 Consortium. Integrating common and rare genetic variation in diverse human populations. *Nature* **467**, 52–8 (2010).
55. Narayanan, A. *et al.* *Redefining Genomic Privacy: Trust and Empowerment*. *bioRxiv* (2014). doi:10.1101/006601
56. US Department of Commerce, N. NIST Requests Comments on a Draft Privacy Risk Management Framework. at <[http://www.nist.gov/itl/201506\\_privacy\\_framework.cfm](http://www.nist.gov/itl/201506_privacy_framework.cfm)>
57. Wong, R. C.-W. W., Fu, A. W.-C. C., Wang, K. & Pei, J. Minimality attack in privacy preserving data publishing. in *Proc. 33rd Int. Conf. Very large data bases* 543–554 (2007). at <<http://dl.acm.org/citation.cfm?id=1325851.1325914>>
58. Alvim, M. S., Andrés, M. E., Chatzikokolakis, K., Degano, P. & Palamidessi, C. Differential privacy: On the trade-off between utility and information leakage. in *Lect. Notes Comput. Sci. (including Subser. Lect. Notes Artif. Intell. Lect. Notes Bioinformatics)* **7140 LNCS**, 39–54 (2012).
59. Cover, T. M. & Thomas, J. A. *Elements of Information Theory*. *Elem. Inf. Theory* (2005). doi:10.1002/047174882X
60. Herbert A. Sturges. The Choice of a Class Interval. *J. Am. Stat. Assoc.* **21**, 65–66 (1926).
61. Lappalainen, T. *et al.* Transcriptome and genome sequencing uncovers functional variation in humans. *Nature* **501**, 506–11 (2013).