Integrating SV calls across pipelines

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Challenge

2000+ whole cancer genomes

- (At least) 3 different pipelines
 - 3 different sets of SV calls (outputting genomic breakpoints)
 - 3 different sets of copy number segmentations
- How do we generate a 'consensus' set of SV & CN calls for each genome?

1,000 Genomes experience

- Multiple different platforms
 - Read depth; aberrant read-pairs; split read mapping; de novo assembly etc

- Relied heavily on:
 - Orthogonal data sets (PacBio, trios, validation by PCR/capillary sequencing, copy number arrays)
 - Genotype concordance across individuals

Scenarios for integrating SV calls

Concordance

 All pipelines call same variant at same location and with same microhomology / NTS

Semi-concordance

 Same basic variant but details of exact location or microhom/ NTS differ

Differential calls

Variant called by some pipelines but not others

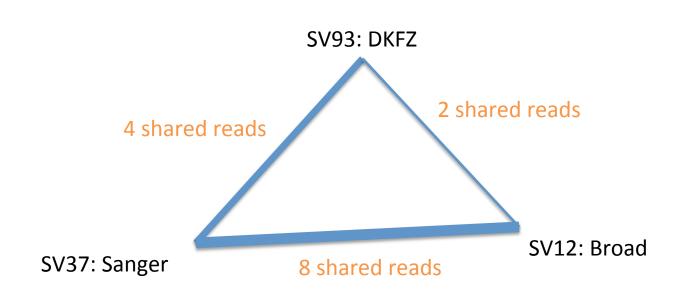
Split calls

 Some pipelines call 2 different variants, where others lump together as a single variant

Proposal: Cliques based on supporting reads

- Each SV call in each pipeline is made on the basis of supporting reads
- Where the pipelines use the same supporting reads in SV calls, probably reporting the same variant
- One can quantify concordance between pipelines in clique membership

Cliques of SV calls in PCAWG – Concordance & semi-concordance

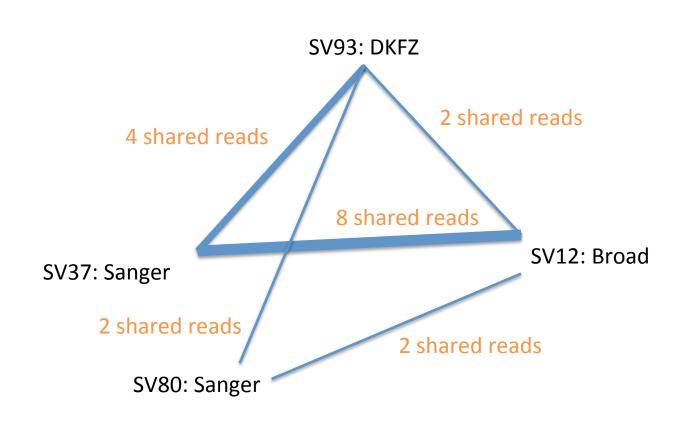


Cliques of SV calls in PCAWG – Differential calls

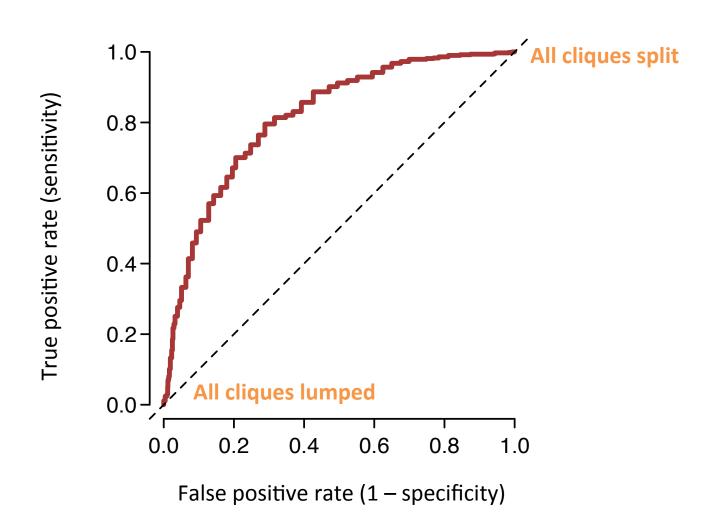
SV93: DKFZ

SV37: Sanger 8 shared reads

Cliques of SV calls in PCAWG – Split calls



Lumping and splitting cliques



Potential 'gold standard' data sets to train cut-offs for lumping vs splitting

- A set of visually inspected copy number changes from core 50 genomes
- 3 cell lines with 10% BAC libraries sequenced to finishing standard
 - Bignell et al, Genome Research 2007
- Samples with several lesions sequenced
 - Eg primary & met; multiple lesions of primary
- PCR validation of novel calls
- Ultra-high depth medulloblastoma genome