**Comparative Analysis of the Transcriptome across Distant Species**

Mark B. Gerstein1,2,3,\*,#, Joel Rozowsky1,2,\*, Koon-Kiu Yan1,2,\*, Daifeng Wang1,2,\*, Chao Cheng4,5,\*, James B. Brown6,7,\*, Carrie A. Davis8,\*, LaDeana Hillier9,\*, Cristina Sisu1,2,\*, Jingyi Jessica Li7,\*, Baikang Pei1,2,\*, Arif O. Harmanci1,2,\*, Michael O. Duff10,\*, Sarah Djebali11,12,\*, Roger P. Alexander1,2, Burak H. Alver13, Raymond K. Auerbach1,2, Kimberly Bell8, Peter J. Bickel7, Max E. Boeck9, Nathan P. Boley6,7, Benjamin W. Booth6, Lucy Cherbas14,15, Peter Cherbas14,15, Chao Di16, Alex Dobin8, Jorg Drenkow8, Brent Ewing9, Gang Fang1,2, Megan Fastuca8, Elise A. Feingold17, Adam Frankish18, Guanjun Gao16, Peter J. Good17, Phil Green9, Roderic Guigó11,12, Ann Hammonds6, Jen Harrow18, Roger A. Hoskins6, Cédric Howald19,20, Long Hu16, Haiyan Huang7, Tim J. P. Hubbard18, Chau Huynh9, Sonali Jha8, Dionna Kasper21, Masaomi Kato22, Thomas C. Kaufman14, Robert R. Kitchen1,2, Erik Ladewig23, Julien Lagarde11,12, Eric Lai23, Jing Leng1,2, Zhi Lu16, Michael MacCoss9, Gemma May10,24, Rebecca McWhirter25, Gennifer Merrihew9, David M. Miller25, Ali Mortazavi26,27, Rabi Murad26,27, Brian Oliver28, Sara Olson10, Peter Park13, Michael J. Pazin17, Norbert Perrimon29,30, Dmitri Pervouchine11,12, Valerie Reinke21, Alexandre Reymond19, Garrett Robinson7, Anastasia Samsonova29,30, Gary I. Saunders18, Felix Schlesinger8, Anurag Sethi1,2, Frank J. Slack22, William C. Spencer25, Marcus H. Stoiber7, Pnina Strasbourger9, Andrea Tanzer31,32, Owen A. Thompson9, Kenneth H. Wan6, Guilin Wang21, Huaien Wang8, Kathie L. Watkins25, Jiayu Wen23, Kejia Wen16, Chenghai Xue8, Li Yang10,33, Kevin Yip34,35, Chris Zaleski8, Yan Zhang1,2, Henry Zheng1,2, Steven E. Brenner36,37,#, Brenton R. Graveley10,#, Susan E. Celniker6,#, Thomas R Gingeras8,#, Robert Waterston9,#

\* These authors contributed equally to this work.

# Co-senior authors.

1 Program in Computational Biology and Bioinformatics, Yale University, Bass 432, 266 Whitney Avenue, New Haven, Connecticut 06520, USA.

2 Department of Molecular Biophysics and Biochemistry, Yale University, Bass 432, 266 Whitney Avenue, New Haven, Connecticut 06520, USA.

3 Department of Computer Science, Yale University, 51 Prospect St, New Haven, Connecticut 06511, USA.

4 Department of Genetics, Geisel School of Medicine at Dartmouth, Hanover, New Hampshire 03755, USA

5 Institute for Quantitative Biomedical Sciences, Norris Cotton Cancer Center, Geisel School of Medicine at Dartmouth, Lebanon, New Hampshire 03766, USA

6 Department of Genome Dynamics, Lawrence Berkeley National Laboratory, Berkeley, California 94720, USA.

7 Department of Statistics, University of California, Berkeley, 367 Evans Hall Berkeley, CA 94720-3860

8 Cold Spring Harbor Laboratory, Cold Spring Harbor, New York 11724, USA.

9 Department of Genome Sciences and University of Washington School of Medicine, William H. Foege Bldg. S350D, 1705 N.E. Pacific Street, Box 355065 Seattle , Washington 98195-5065, USA.

10 Department of Genetics and Developmental Biology, Institute for Systems Genomics, University of Connecticut Health Center, 400 Farmington Avenue, Farmington, CT 06030 USA

11 Centre for Genomic Regulation, Barcelona, Catalonia, Spain

12 Departament de Ciències Experimentals i de la Salut, Universitat Pompeu Fabra, Barcelona, Catalonia, Spain

13 Center for Biomedical Informatics, Harvard Medical School, 10 Shattuck St. Boston, Massachusetts 02115, USA.

14 Department of Biology, Indiana University, 1001 E. 3rd Street, Bloomington, Indiana 47405-7005, USA.

15 Center for Genomics and Bioinformatics, Indiana University, 1001 E. 3rd Street, Bloomington, Indiana 47405-7005, USA.

16 MOE Key Lab of Bioinformatics, School of Life Sciences, Tsinghua University, Beijing, China 100084

17 National Human Genome Research Institute, National Institutes of Health, 5635 Fishers Lane, Bethesda, Maryland 20892-9307, USA.

18 Wellcome Trust Sanger Institute, Wellcome Trust Genome Campus, Hinxton, UK

19 Center for Integrative Genomics, University of Lausanne, 1015 Lausanne, Switzerland

20 Swiss Institute of Bioinformatics, 1015 Lausanne, Switzerland

21 Department of Genetics, Yale University School of Medicine, New Haven, Connecticut 06520-8005, USA.

22 Department of Molecular, Cellular and Developmental Biology, PO Box 208103, Yale University, New Haven, Connecticut 06520, USA.

23 Sloan-Kettering Institute, 1275 York Avenue, Box 252, New York, New York 10065, USA.

24 Department of Biological Sciences, Carnegie Mellon University, Pittsburgh, PA 15213 USA

25 Department of Cell and Developmental Biology, Vanderbilt University, 465 21st Avenue South, Nashville, Tennessee 37232-8240, USA.

26 Developmental and Cell Biology, University of California, Irvine, CA 92697

27 Center for Complex Biological Systems, University of California, Irvine, CA 92697

28 Section of Developmental Genomics, Laboratory of Cellular and Developmental Biology, National Institute of Diabetes and Digestive and Kidney Diseases, National Institutes of Health, Bethesda MD 20892, USA.

29 Department of Genetics and Drosophila RNAi Screening Center, Harvard Medical School, 77 Avenue Louis Pasteur, Boston, Massachusetts 02115, USA.

30 Howard Hughes Medical Institute, Harvard Medical School, 77 Avenue Louis Pasteur, Boston, Massachusetts 02115, USA.

31 Bioinformatics and Genomics Programme, Center for Genomic Regulation, Universitat Pompeu Fabra (CRG-UPF), Barcelona, Catalonia, Spain

32 Institute for Theoretical Chemistry, Theoretical Biochemistry Group (TBI), University of Vienna

33 Key Laboratory of Computational Biology, CAS-MPG Partner Institute for Computational Biology, Shanghai Institutes for Biological Sciences, Chinese Academy of Sciences, Shanghai 200031, China

34 Hong Kong Bioinformatics Centre, The Chinese University of Hong Kong, Shatin, New Territories, Hong Kong

35 5 CUHK-BGI Innovation Institute of Trans-omics, The Chinese University of Hong Kong, Shatin, New Territories, Hong Kong

36 Department of Molecular and Cell Biology, University of California, Berkeley, California 94720, USA.

37 Department of Plant & Microbial Biology, University of California, Berkeley, California 94720, USA.

The transcriptome is the readout of the genome. Identifying common features in it across distant species can reveal fundamental principles. To this end, the ENCODE and modENCODE consortia have generated large amounts of matched RNA-sequencing data for human, worm and fly. Uniform processing and comprehensive annotation of these data allow comparison across metazoan phyla, extending beyond earlier within-phylum transcriptome comparisons and revealing ancient, conserved features.\cite{22012392,23258891,23258890,22560298,21150996,20969771} In particular, we discovered co-expression modules shared across animals, many of which are enriched in developmental genes. We used expression patterns to align the stages in worm and fly development, finding a novel pairing between worm embryo and fly pupae in addition to the expected embryo-to-embryo and larvae-to-larvae pairings. Furthermore, we found that the extent of non-canonical non-coding transcription is similar in each organism, per base-pair. Finally, we found the gene expression levels in all three organisms, both coding and non-coding, can be quantitatively predicted from their upstream histone marks using a “universal model," based on a single set of organism-independent parameters.

Our comparison used the ENCODE-modENCODE RNA resource (Fig. ED1). This resource comprises: (1) deeply sequenced RNA-Seq data from many distinct samples from all three organisms; (2) comprehensive annotation of transcribed elements and (3) uniformly processed, standardized analysis files, focusing on non-coding transcription and expression patterns. Where practical, these datasets match comparable samples across organisms and to other types of functional genomics data. The resource represents 575 different experiments containing >67B reads. It encompasses many different RNA types, including poly(A)+, poly(A)- and ribosomal-RNA-depleted RNA and short and long RNA. It represents a capstone for the decade-long annotation efforts in human, worm, and fly.

The new annotation sets have numbers, sizes and families of protein-coding genes similar to previous compilations; however, the number of pseudogenes and annotated ncRNAs differ (Figs. ED2, ED3). Also, the number of splicing events is greatly increased. We find the proportion of the different types of alternative splicing (e.g., exon skipping or intron retention) is approximately similar across the three organisms; however, skipped exons predominate in human while retained introns are most common in fly\cite{Talerico-M-&-Berget} (Figs. ED4, S1 and Table S1). Moreover, a considerable fraction of the transcription comes from genomic regions not associated with standard annotations, representing "non-canonical transcription” (Table S2)\cite{22955620}. Uniform processing of reads mapping outside of protein-coding transcripts, pseudogenes and annotated ncRNAs identified read-clusters (transcriptionally active regions, TARs) using a minimum-run/maximum-gap algorithm. We found across all three genomes roughly one third of the bases gives rise to "non-canonical" transcription (Fig. ED3). To determine the extent that this transcription represents an expansion of the current established classes of ncRNAs, we identified the TARs most similar to known annotated ncRNAs using a supervised classifier\cite{21177971} (Fig. S2, Table S2). We validated these predictions using RT-PCR, demonstrating high accuracy. Overall, the predictions encompass only a small fraction of all TARs, suggesting that most TARs have features distinct from annotated ncRNAs and that the majority of ncRNAs of established classes have already been identified. To shed further light on the possible roles of TARs we intersected them with enhancers and HOT regions \cite{mod3,21177976,21177974,mod2,22955620}, finding statistically significant overlaps (Fig. ED7, Table S2).

Given the uniformly processed nature of the data and annotations, we were able to make comparisons across organisms. First, we built co-expression modules, extending earlier analysis\cite{12934013}(Fig. 1a). To detect modules consistently across the three species, we combined across-species orthology and within-species co-expression relationships. We then searched for dense subgraphs (modules) in the resulting multilayer network, using simulated annealing\cite{17813860,15601068}. We found some modules dominated by one species, whereas others contain genes from two or three. As expected, the modules with genes from multiple species are enriched in orthologs. Moreover, a phylogenetic analysis shows that the genes in such modules are more preserved across 56 animal species (Fig. ED5, S3). To focus on the cross-species conserved functions, we restricted the clustering to orthologs, arriving at 16 conserved modules, which are enriched in a variety of functions, ranging from morphogenesis to chromatin remodeling (Fig. 1a, Table S3). Finally, we annotated many TARs based on correlating their expression profiles with these modules (Fig. ED7).

Next, we used the expression profiles of orthologous genes to align the developmental stages in worm and fly (Fig. 1b, ED6). Specifically, for every developmental stage, we identified stage-associated genes, i.e. genes highly expressed at a particular stage but not across all stages. We then counted the number of orthologous pairs among these stage-associated genes for each possible worm-and-fly stage correspondence, aligning stages by the significance of the overlap. Strikingly, worm stages map to two sets of fly stages: first, they match in the expected one-to-one fashion to the fly (i.e. embryos-to-embryos, larvae-to-larvae). However, worm late embryonic stages also match fly pupal stages, suggesting a shared expression program between embryogenesis and metamorphosis. The ~50 genes involved in this dual-stage mapping are enriched in functions such as ion transport and cation-channel activity (Table S3).

To gain further insight into the stage alignment, we examined our 16 conserved modules in terms of the "hourglass hypothesis", which posits that all animals go through a particular stage in embryonic development (the tight point of the hourglass or "phylotypic" stage) during which the expression divergence across species for orthologous genes is smallest\cite{21150996,22560298,21150997}. For genes in 12 of the modules, we observed canonical hourglass behavior, i.e. "inter-organism" expression divergence across closely related fly species during development is minimal\cite{21150996}(Fig. S3). Moreover, we find a subset of TARs also exhibit "hourglass" behavior (Fig. S2). Beyond looking at *inter*-species divergence, we also investigated the *intra*-species divergence within just *D. melanogaster* and *C. elegans.* Strikingly, we observed that divergence of gene expression between modules is minimized during the worm and fly phylotypic stages (Fig. 1c). This suggests, for an individual species, the expression patterns of different modules are most tightly coordinated (low divergence) during the phylotypic stage, but each module has its own signature before and after this. One can, in fact, directly see this coordination as a local maximum in the between-module correlation for the worm (Fig. ED5). Finally, using genes from just the 12 "hourglass modules," we found that the alignment between worm and fly stages becomes stronger (Fig. 1b, S3). The alignment shows a gap where no changes are observed, perfectly matching the phylotypic stage.

The uniformly processed and matched nature of the transcriptome data also facilitated integration with upstream factor-binding and chromatin-modification signals. We investigated the degree to which these upstream signals can quantitatively predict gene expression and how consistent this prediction is across organisms. As previously found\cite{20133639,ENCODE-main-paper ,modencode2010-paper}, we found consistent correlations in each of the three species between the histone-modification signal and the expression level of the downstream gene: around the TSS, H3K4me1, H3K4me2, H3K4me3 and H3K27ac are positively correlated, whereas H3K27me3 is negatively correlated (Figs. 2, ED8, S4). Then for each organism, we integrated these individual correlations into multivariate, statistical models, obtaining high accuracy in predicting expression for protein-coding genes and ncRNAs. The promoter-associated marks, H3K4me2 and H3K4me3, consistently have the highest contribution to the models.

A similar statistical analysis with TFs showed the correlation between gene expression and TF binding to be the greatest at the TSS, positively for activators and negatively for repressors (Fig. ED8). Integrated models in each organism also achieved high accuracy for protein-coding genes and ncRNAs, with only a few TFs necessary for accurate predictions, often as few as five (Fig. ED9). This perhaps reflects an intricate, correlated structure to regulation. The relative importance of the upstream regions is more peaked for the TF models than for the histone ones, likely reflecting the fact that histone modifications are spread over broad regions, including the gene body, whereas most TFs bind near the promoter.

Finally, we constructed a "universal model," containing a single set of organism-independent parameters. This achieved accuracy comparable to the organism-specific models. In the universal model, the consistently important promoter-associated marks such as H3K4me2 and H3K4me3 are weighted most highly. In contrast, the enhancer mark H3K4me1 is down-weighted, perhaps reflecting that signals for most human enhancers are not near the TSS. The same universal model also can predict ncRNA expression, i.e. using the same set of organism-independent parameters derived from training on protein-coding genes.

Our comparison of the transcriptomes of these three highly dissimilar metazoans, a comparison not previously reported, highlights fundamental features of transcription conserved across animal phyla. First, there are ancient co-expression modules across organisms, many of which are enriched for developmentally important “hourglass” genes. These conserved modules have highly coordinated intra-organism expression during the phylotypic stage, but display diversified expression before and after. The expression clustering also aligns developmental stages between worm and fly, revealing shared expression programs between embryogenesis and metamorphosis. Finally, we were able to build a single model that could predict transcription in all three organisms from upstream histone marks using a single set of parameters for both protein-coding genes and ncRNAs. Overall, our results underscore the importance of comparing divergent model organisms to human to highlight conserved biological principles (and dis-entangle them from lineage-specific adaptations).

## Methods

Detailed methods are in the supplement. (See first section of this for guide.) Data files are available from cmptxn.gersteinlab.org and [www.encodedcc.org/modencode/comparative\_RNA.html](http://www.encodedcc.org/modencode/comparative_RNA.html) .

**Fig 1: Expression Clustering.** (A) Left: Human, worm, and fly gene-gene co-association matrix; darker blocks reflect increased likelihood pairs of genes are assigned to the same module. Blocks along the diagonal represent groups of human, worm, and fly genes. Associated off diagonal blocks create cross-species modules, connecting matching genes in two species. Diagonal blocks without any off these form species-specific modules. Right: The functional enrichment of genes within each module is shown. (B) Alignment of worm-and-fly developmental stages based on all worm-fly orthologs. Inset shows worm-fly stage alignment using only hourglass orthologs is more significant and exhibits a gap (brown) matching the phylotypic stage. (C) Expression of 16 conserved modules shows the smallest intra-organism divergence during the phylotypic stage (brown).

**Fig 2: Histone Models for Gene Expression.** Top: Normalized correlations of two representative histone marks with expression. Left: Relative importance of the histone marks in organism-specific models and the universal model. Right: Prediction accuracy of the organism-specific and universal models.

## Acknowledgements

The authors thank the NHGRI and the ENCODE and modENCODE projects for support.

# Extended Data (ED) Figure Captions

**Fig ED1 - Overview of the data.** (A) Schematic of the RNA-seq data generated for human (red), worm (green), and fly (blue), showing how it samples developmental stages and various tissues and cell lines. (B) The number and size of data sets generated. The amount of new data beyond that in the previous ENCODE publications\cite{put-in-prev-pubs} is indicated by the degree the white bar extends over the colored ones. (See Supplement section XXX for a detailed description of these data.)

**Fig ED2 - Summary statistics for the protein coding gene annotations.**  (A) Distributions of key summary statistics - gene span, longest ORF per gene, CDS exon length, and CDS exons per gene; note that the x axes are in log scale. Both fly and worm genes span similar genomic lengths while human genes span larger regions (mostly due to the size of human introns). (B) Left: Venn diagram of protein domains (from the Pfam database version 26.0) present in annotated protein-coding genes in each species. Right: Shared domain combinations. (For more information on domain combinations, see Fig S1h and Supplement section XXX.)

**Fig ED3 - Summary of annotated ncRNAs, TARs, and ncRNA predictions in each species.** This shows the number of elements, the base pairs covered and the fraction of the genome for each class (see also Supplement). There are comparable numbers of tRNAs in humans and worms but about half as many in fly. While the number of lncRNAs in human is more than an order of magnitude greater than in either worms or flies, the fractional genomic coverage in all three species is, in fact, similar. Finally, humans have at least 5-fold more miRNAs, snoRNAs and snRNAs as compared to worm or fly.The fraction of the genome covered by TARs (highlighted squares) for each species is similar. A large amount of non canonical transcription occurs in the introns of annotated genes, presumably representing a mixture of unprocessed mRNAs and internally initiated transcripts. The remaining non-canonical transcription (249Mb, 16Mb, and 14Mb in human, worm, and fly) is intergenic and occurs at low levels, comparable to that observed for introns (Table S2). Overall, the fraction of the genome transcribed -- including intronic, exonic, and non-canonical transcription -- is consistent with that previously reported for human despite the methodological differences in the analysis (Fig. S2, Supplement section C).

**Fig ED4 - Analysis of Alternative Splicing.** (A) Representative orthologous genes do not share the same exon/intron structure or alternative splicing. (B) Distribution of the number of isoforms per gene. (C) Comparison of the fraction of various alternative splicing event classes in human, worm, and fly -- skipped exons “SE”, retained introns “RI”, alternative 3' splice sites “A3SS”, alternative 5' splice sites “A5SS”, alternative first exons “AFE”, alternative last exons “ALE”, tandem 3' UTRs “TandemUTR”, coordinately skipped exons “CSE”, and mutually exclusive exons “MXE”. (See Supplement section XXX for a further discussion of splicing.)

**Fig ED5 - Further Detail on Expression Clustering.** (A) Pie charts showing gene conservation across 56 Ensembl species for the blocks in the Fig. 1 heatmap enclosed with the same symbol (i.e. pentagon here matches pentagon in Fig.1a). Overall, species-specific modules tend to have fewer orthologs across 56 Ensembl species. (B) The expression levels of a conserved module (Module No. 5) in *D. melanogaster* and its orthologous counterparts in other 5 *Drosophila* species are plotted against time. The x-axis represents the middle time points of two-hour periods at fly embryo stages. The boxes represent the log10 modular expression levels from microarray data of 6 *Drosophila* species centered by their medians. The modular expression divergence (inter-quartile region) becomes minimal during the fly phylotypic stage (brown, 8-10 hours). Part (C) shows the modular expression correlations over a sliding 2-hour window (Pearson correlation per 5 stages, middle time of two-hour period in x-axis) among 16 modules in worm. We found that the modular correlations (median shown as bar height in y-axis) are highest during the worm phylotypic stages (brown), 6-8 hours. One can, in fact, directly see this coordination as a local maximum in the between-module correlation for the worm, which has a more densely sampled developmental time course. (This figure provides more detail on Fig. 1a and 1c. More details on all parts of this figure are in Supplement section D and Figure S3.)

**Fig ED6 - Further Detail on Stage Alignment.** This figure provides further detail beyond Fig. 1b. (A) An alignment of worm and fly developmental stages based on all worm-fly orthologs (11,403 pairs, including one-to-one, one-to-many, many-to-many pairs). (B) Alignment of worm and fly developmental stages based on just worm-fly hourglass orthologs. Note the prominent gap in the aligned stages coincides with the worm and fly phylotypic stages (brown band). This make sense: since the expression values of genes in all hourglass modules converge at the phylotypic stage, no hourglass genes can be phylotypic-stage specific, and hence, the gap. (C) Key aligned stages from part (A). Worm “early embryo” and “late embryo” stages are matched with fly “early embryo” and “late embryo” respectively in the “lower diagonal” set of matches, and they are also matched with fly “L1” and “prepupa-pupa” stages respectively in the “upper diagonal” set of matches. (More details on all parts of this figure are in Supplement section XXX and Table S3.)

**Fig ED7 - Characterizing Non-canonical Transcription.** (A) The left column highlights ncRNA/TARs that are highly correlated with corresponding HOX orthologues in human (HOXB4), worm (lin-39), and fly (Dfd). The expression of mir-10 correlates strongly with Dfd in fly (r=0.66, p<6e-4 in fly), as does mir-10a in human, which correlates strongly with HOXB4 (r=0.88, p<2e-9). A TAR (chrIII:8871234-2613) strongly correlates with lin-39 (r=0.91, p<4e-13) in worm. The right column shows TARs in human (chr19:7698570-7701990), worm (chrII:11469045-440), and fly (chr2L:2969620-772) that are negatively correlated with the expression of three orthologous genes: SGCB (r=-0.91, p<3e-16), sgcb-1 (r=-0.86, p<2e-7), and Scgb (r=-0.82, p<4e-8), respectively. (B) The overlap of enhancers and distal HOT regions with supervised ncRNA predictions and TARs in human, worm, and fly. The overlap of enhancers and distal HOT regions with respect to both supervised ncRNA predictions as well as TARs are significantly enriched compared to a randomized expectation (More details on all parts of this figure are in Supplement section C and Table XXX.)

**Fig ED8 - Further Detail on Statistical Models for Predicting Gene Expression.** This figure provides further information beyond that in Fig. 2. Binding/expression correlations of (A) various histone marks and (C) TFs. For instance, H3K36me3 shows positive correlation in worm and fly, but weak negative correlation in human at the promoter, with positive correlation over the gene body. (B) The positional accuracy from the TF and histone-mark models for predicting mRNA and ncRNA expression about the TSS. (More details on all parts of this figure are in Supplement section XXX and Fig. XXX.)

**Fig ED9** - **Average predictive accuracy of models with different number of randomly selected TFs.** We randomly selected n TFs as predictors and examined the predictive accuracy by cross-validation, with n was taken from 2 to 28. The curve shows the average predictive accuracy (Fig. S4 indicate the standard deviation of all models with the same number of predictors). Surprisingly, models with as few as 5 TFs have predictive accuracy. This may reflect an intricate, correlated structure to regulation. However, it could also be that open chromatin is characteristic of gene expression and TFs bind somewhat indiscriminately. (More details on all parts of this figure are in Supplement section XXX.)

# 

# 

# References:

1. ENCODE Project Consortium, Dunham, I., Kundaje, A., Aldred, S. F., Collins, P. J., Davis, C. A., Doyle, F., Epstein, C. B., Frietze, S., Harrow, J., Kaul, R., Khatun, J., Lajoie, B. R., Landt, S. G., Lee, B.-K., Pauli, F., Rosenbloom, K. R., Sabo, P., Safi, A., Sanyal, A., Shoresh, N., Simon, J. M., Song, L., Trinklein, N. D., Altshuler, R. C., Birney, E., Brown, J. B., Cheng, C., Djebali, S., Dong, X., Dunham, I., Ernst, J., Furey, T. S., Gerstein, M., Giardine, B., Greven, M., Hardison, R. C., Harris, R. S., Herrero, J., Hoffman, M. M., Iyer, S., Kelllis, M., Khatun, J., Kheradpour, P., Kundaje, A., Lassman, T., Li, Q., Lin, X., Marinov, G. K., Merkel, A., Mortazavi, A., Parker, S. C. J., Reddy, T. E., Rozowsky, J., Schlesinger, F., Thurman, R. E., Wang, J., Ward, L. D., Whitfield, T. W., Wilder, S. P., Wu, W., Xi, H. S., Yip, K. Y., Zhuang, J., Bernstein, B. E., Birney, E., Dunham, I., Green, E. D., Gunter, C., Snyder, M., Pazin, M. J., Lowdon, R. F., Dillon, L. A. L., Adams, L. B., Kelly, C. J., Zhang, J., Wexler, J. R., Green, E. D., Good, P. J., Feingold, E. A., Bernstein, B. E., Birney, E., Crawford, G. E., Dekker, J., Elinitski, L., Farnham, P. J., Gerstein, M., Giddings, M. C., Gingeras, T. R., Green, E. D., Guigó, R., Hardison, R. C., Hubbard, T. J., Kellis, M., Kent, W. J., Lieb, J. D., Margulies, E. H., Myers, R. M., Snyder, M., Starnatoyannopoulos, J. A., Tennebaum, S. A., Weng, Z., White, K. P., Wold, B., Khatun, J., Yu, Y., Wrobel, J., Risk, B. A., Gunawardena, H. P., Kuiper, H. C., Maier, C. W., Xie, L., Chen, X., Giddings, M. C., Bernstein, B. E., Epstein, C. B., Shoresh, N., Ernst, J., Kheradpour, P., Mikkelsen, T. S., Gillespie, S., Goren, A., Ram, O., Zhang, X., Wang, L., Issner, R., Coyne, M. J., Durham, T., Ku, M., Truong, T., Ward, L. D., Altshuler, R. C., Eaton, M. L., Kellis, M., Djebali, S., Davis, C. A., Merkel, A., Dobin, A., Lassmann, T., Mortazavi, A., Tanzer, A., Lagarde, J., Lin, W., Schlesinger, F., Xue, C., Marinov, G. K., Khatun, J., Williams, B. A., Zaleski, C., Rozowsky, J., Röder, M., Kokocinski, F., Abdelhamid, R. F., Alioto, T., Antoshechkin, I., Baer, M. T., Batut, P., Bell, I., Bell, K., Chakrabortty, S., Chen, X., Chrast, J., Curado, J., Derrien, T., Drenkow, J., Dumais, E., Dumais, J., Duttagupta, R., Fastuca, M., Fejes-Toth, K., Ferreira, P., Foissac, S., Fullwood, M. J., Gao, H., Gonzalez, D., Gordon, A., Gunawardena, H. P., Howald, C., Jha, S., Johnson, R., Kapranov, P., King, B., Kingswood, C., Li, G., Luo, O. J., Park, E., Preall, J. B., Presaud, K., Ribeca, P., Risk, B. A., Robyr, D., Ruan, X., Sammeth, M., Sandu, K. S., Schaeffer, L., See, L.-H., Shahab, A., Skancke, J., Suzuki, A. M., Takahashi, H., Tilgner, H., Trout, D., Walters, N., Wang, H., Wrobel, J., Yu, Y., Hayashizaki, Y., Harrow, J., Gerstein, M., Hubbard, T. J., Reymond, A., Antonarakis, S. E., Hannon, G. J., Giddings, M. C., Ruan, Y., Wold, B., Carninci, P., Guigó, R., Gingeras, T. R., Rosenbloom, K. R., Sloan, C. A., Learned, K., Malladi, V. S., Wong, M. C., Barber, G. P., Cline, M. S., Dreszer, T. R., Heitner, S. G., Karolchik, D., Kent, W. J., Kirkup, V. M., Meyer, L. R., Long, J. C., Maddren, M., Raney, B. J., Furey, T. S., Song, L., Grasfeder, L. L., Giresi, P. G., Lee, B.-K., Battenhouse, A., Sheffield, N. C., Simon, J. M., Showers, K. A., Safi, A., London, D., Bhinge, A. A., Shestak, C., Schaner, M. R., Kim, S. K., Zhang, Z. Z., Mieczkowski, P. A., Mieczkowska, J. O., Liu, Z., McDaniell, R. M., Ni, Y., Rashid, N. U., Kim, M. J., Adar, S., Zhang, Z., Wang, T., Winter, D., Keefe, D., Birney, E., Iyer, V. R., Lieb, J. D., Crawford, G. E., Li, G., Sandhu, K. S., Zheng, M., Wang, P., Luo, O. J., Shahab, A., Fullwood, M. J., Ruan, X., Ruan, Y., Myers, R. M., Pauli, F., Williams, B. A., Gertz, J., Marinov, G. K., Reddy, T. E., Vielmetter, J., Partridge, E. C., Trout, D., Varley, K. E., Gasper, C., Bansal, A., Pepke, S., Jain, P., Amrhein, H., Bowling, K. M., Anaya, M., Cross, M. K., King, B., Muratet, M. A., Antoshechkin, I., Newberry, K. M., McCue, K., Nesmith, A. S., Fisher-Aylor, K. I., Pusey, B., DeSalvo, G., Parker, S. L., Balasubramanian, S., Davis, N. S., Meadows, S. K., Eggleston, T., Gunter, C., Newberry, J. S., Levy, S. E., Absher, D. M., Mortazavi, A., Wong, W. H., Wold, B., Blow, M. J., Visel, A., Pennachio, L. A., Elnitski, L., Margulies, E. H., Parker, S. C. J., Petrykowska, H. M., Abyzov, A., Aken, B., Barrell, D., Barson, G., Berry, A., Bignell, A., Boychenko, V., Bussotti, G., Chrast, J., Davidson, C., Derrien, T., Despacio-Reyes, G., Diekhans, M., Ezkurdia, I., Frankish, A., Gilbert, J., Gonzalez, J. M., Griffiths, E., Harte, R., Hendrix, D. A., Howald, C., Hunt, T., Jungreis, I., Kay, M., Khurana, E., Kokocinski, F., Leng, J., Lin, M. F., Loveland, J., Lu, Z., Manthravadi, D., Mariotti, M., Mudge, J., Mukherjee, G., Notredame, C., Pei, B., Rodriguez, J. M., Saunders, G., Sboner, A., Searle, S., Sisu, C., Snow, C., Steward, C., Tanzer, A., Tapanan, E., Tress, M. L., van Baren, M. J., Walters, N., Washieti, S., Wilming, L., Zadissa, A., Zhengdong, Z., Brent, M., Haussler, D., Kellis, M., Valencia, A., Gerstein, M., Raymond, A., Guigó, R., Harrow, J., Hubbard, T. J., Landt, S. G., Frietze, S., Abyzov, A., Addleman, N., Alexander, R. P., Auerbach, R. K., Balasubramanian, S., Bettinger, K., Bhardwaj, N., Boyle, A. P., Cao, A. R., Cayting, P., Charos, A., Cheng, Y., Cheng, C., Eastman, C., Euskirchen, G., Fleming, J. D., Grubert, F., Habegger, L., Hariharan, M., Harmanci, A., Iyenger, S., Jin, V. X., Karczewski, K. J., Kasowski, M., Lacroute, P., Lam, H., Larnarre-Vincent, N., Leng, J., Lian, J., Lindahl-Allen, M., Min, R., Miotto, B., Monahan, H., Moqtaderi, Z., Mu, X. J., O'Geen, H., Ouyang, Z., Patacsil, D., Pei, B., Raha, D., Ramirez, L., Reed, B., Rozowsky, J., Sboner, A., Shi, M., Sisu, C., Slifer, T., Witt, H., Wu, L., Xu, X., Yan, K.-K., Yang, X., Yip, K. Y., Zhang, Z., Struhl, K., Weissman, S. M., Gerstein, M., Farnham, P. J., Snyder, M., Tenebaum, S. A., Penalva, L. O., Doyle, F., Karmakar, S., Landt, S. G., Bhanvadia, R. R., Choudhury, A., Domanus, M., Ma, L., Moran, J., Patacsil, D., Slifer, T., Victorsen, A., Yang, X., Snyder, M., White, K. P., Auer, T., Centarin, L., Eichenlaub, M., Gruhl, F., Heerman, S., Hoeckendorf, B., Inoue, D., Kellner, T., Kirchmaier, S., Mueller, C., Reinhardt, R., Schertel, L., Schneider, S., Sinn, R., Wittbrodt, B., Wittbrodt, J., Weng, Z., Whitfield, T. W., Wang, J., Collins, P. J., Aldred, S. F., Trinklein, N. D., Partridge, E. C., Myers, R. M., Dekker, J., Jain, G., Lajoie, B. R., Sanyal, A., Balasundaram, G., Bates, D. L., Byron, R., Canfield, T. K., Diegel, M. J., Dunn, D., Ebersol, A. K., Ebersol, A. K., Frum, T., Garg, K., Gist, E., Hansen, R. S., Boatman, L., Haugen, E., Humbert, R., Jain, G., Johnson, A. K., Johnson, E. M., Kutyavin, T. M., Lajoie, B. R., Lee, K., Lotakis, D., Maurano, M. T., Neph, S. J., Neri, F. V., Nguyen, E. D., Qu, H., Reynolds, A. P., Roach, V., Rynes, E., Sabo, P., Sanchez, M. E., Sandstrom, R. S., Sanyal, A., Shafer, A. O., Stergachis, A. B., Thomas, S., Thurman, R. E., Vernot, B., Vierstra, J., Vong, S., Wang, H., Weaver, M. A., Yan, Y., Zhang, M., Akey, J. A., Bender, M., Dorschner, M. O., Groudine, M., MacCoss, M. J., Navas, P., Stamatoyannopoulos, G., Kaul, R., Dekker, J., Stamatoyannopoulos, J. A., Dunham, I., Beal, K., Brazma, A., Flicek, P., Herrero, J., Johnson, N., Keefe, D., Lukk, M., Luscombe, N. M., Sobral, D., Vaquerizas, J. M., Wilder, S. P., Batzoglou, S., Sidow, A., Hussami, N., Kyriazopoulou-Panagiotopoulou, S., Libbrecht, M. W., Schaub, M. A., Kundaje, A., Hardison, R. C., Miller, W., Giardine, B., Harris, R. S., Wu, W., Bickel, P. J., Banfai, B., Boley, N. P., Brown, J. B., Huang, H., Li, Q., Li, J. J., Noble, W. S., Bilmes, J. A., Buske, O. J., Hoffman, M. M., Sahu, A. O., Kharchenko, P. V., Park, P. J., Baker, D., Taylor, J., Weng, Z., Iyer, S., Dong, X., Greven, M., Lin, X., Wang, J., Xi, H. S., Zhuang, J., Gerstein, M., Alexander, R. P., Balasubramanian, S., Cheng, C., Harmanci, A., Lochovsky, L., Min, R., Mu, X. J., Rozowsky, J., Yan, K.-K., Yip, K. Y., & Birney, E. An integrated encyclopedia of DNA elements in the human genome. *Nature* 489, 57-74, (2012)

2. Gerstein, M. B., Lu, Z. J., Van Nostrand, E. L., Cheng, C., Arshinoff, B. I., Liu, T., Yip, K. Y., Robilotto, R., Rechtsteiner, A., Ikegami, K., Alves, P., Chateigner, A., Perry, M., Morris, M., Auerbach, R. K., Feng, X., Leng, J., Vielle, A., Niu, W., Rhrissorrakrai, K., Agarwal, A., Alexander, R. P., Barber, G., Brdlik, C. M., Brennan, J., Brouillet, J. J., Carr, A., Cheung, M.-S., Clawson, H., Contrino, S., Dannenberg, L. O., Dernburg, A. F., Desai, A., Dick, L., Dosé, A. C., Du, J., Egelhofer, T., Ercan, S., Euskirchen, G., Ewing, B., Feingold, E. A., Gassmann, R., Good, P. J., Green, P., Gullier, F., Gutwein, M., Guyer, M. S., Habegger, L., Han, T., Henikoff, J. G., Henz, S. R., Hinrichs, A., Holster, H., Hyman, T., Iniguez, A. L., Janette, J., Jensen, M., Kato, M., Kent, W. J., Kephart, E., Khivansara, V., Khurana, E., Kim, J. K., Kolasinska-Zwierz, P., Lai, E. C., Latorre, I., Leahey, A., Lewis, S., Lloyd, P., Lochovsky, L., Lowdon, R. F., Lubling, Y., Lyne, R., MacCoss, M., Mackowiak, S. D., Mangone, M., McKay, S., Mecenas, D., Merrihew, G., Miller, 3rd, D. M., Muroyama, A., Murray, J. I., Ooi, S.-L., Pham, H., Phippen, T., Preston, E. A., Rajewsky, N., Rätsch, G., Rosenbaum, H., Rozowsky, J., Rutherford, K., Ruzanov, P., Sarov, M., Sasidharan, R., Sboner, A., Scheid, P., Segal, E., Shin, H., Shou, C., Slack, F. J., Slightam, C., Smith, R., Spencer, W. C., Stinson, E. O., Taing, S., Takasaki, T., Vafeados, D., Voronina, K., Wang, G., Washington, N. L., Whittle, C. M., Wu, B., Yan, K.-K., Zeller, G., Zha, Z., Zhong, M., Zhou, X., modENCODE Consortium, Ahringer, J., Strome, S., Gunsalus, K. C., Micklem, G., Liu, X. S., Reinke, V., Kim, S. K., Hillier, L. W., Henikoff, S., Piano, F., Snyder, M., Stein, L., Lieb, J. D., & Waterston, R. H. Integrative analysis of the Caenorhabditis elegans genome by the modENCODE project. *Science* 330, 1775-87, (2010)

3. Graveley, B. R., Brooks, A. N., Carlson, J. W., Duff, M. O., Landolin, J. M., Yang, L., Artieri, C. G., van Baren, M. J., Boley, N., Booth, B. W., Brown, J. B., Cherbas, L., Davis, C. A., Dobin, A., Li, R., Lin, W., Malone, J. H., Mattiuzzo, N. R., Miller, D., Sturgill, D., Tuch, B. B., Zaleski, C., Zhang, D., Blanchette, M., Dudoit, S., Eads, B., Green, R. E., Hammonds, A., Jiang, L., Kapranov, P., Langton, L., Perrimon, N., Sandler, J. E., Wan, K. H., Willingham, A., Zhang, Y., Zou, Y., Andrews, J., Bickel, P. J., Brenner, S. E., Brent, M. R., Cherbas, P., Gingeras, T. R., Hoskins, R. A., Kaufman, T. C., Oliver, B., & Celniker, S. E. The developmental transcriptome of Drosophila melanogaster. *Nature* 471, 473-9, (2011)

4. Brawand, D., Soumillon, M., Necsulea, A., Julien, P., Csárdi, G., Harrigan, P., Weier, M., Liechti, A., Aximu-Petri, A., Kircher, M., Albert, F. W., Zeller, U., Khaitovich, P., Grützner, F., Bergmann, S., Nielsen, R., Pääbo, S., & Kaessmann, H. The evolution of gene expression levels in mammalian organs. *Nature* 478, 343-8, (2011)

5. Merkin, J., Russell, C., Chen, P., & Burge, C. B. Evolutionary dynamics of gene and isoform regulation in Mammalian tissues. *Science* 338, 1593-9, (2012) [[JJL: delete if necessary]]

6. Barbosa-Morais, N. L., Irimia, M., Pan, Q., Xiong, H. Y., Gueroussov, S., Lee, L. J., Slobodeniuc, V., Kutter, C., Watt, S., Colak, R., Kim, T., Misquitta-Ali, C. M., Wilson, M. D., Kim, P. M., Odom, D. T., Frey, B. J., & Blencowe, B. J. The evolutionary landscape of alternative splicing in vertebrate species. *Science* 338, 1587-93, (2012) [[JJL: delete]]

7. Levin, M., Hashimshony, T., Wagner, F., & Yanai, I. Developmental milestones punctuate gene expression in the Caenorhabditis embryo. *Dev Cell* 22, 1101-8, (2012)

8. Kalinka, A. T., Varga, K. M., Gerrard, D. T., Preibisch, S., Corcoran, D. L., Jarrells, J., Ohler, U., Bergman, C. M., & Tomancak, P. Gene expression divergence recapitulates the developmental hourglass model. *Nature* 468, 811-4, (2010)

9. Simola, D. F., Francis, C., Sniegowski, P. D., & Kim, J. Heterochronic evolution reveals modular timing changes in budding yeast transcriptomes. *Genome Biol* 11, R105, (2010)

10. Busby, M. A., Gray, J. M., Costa, A. M., Stewart, C., Stromberg, M. P., Barnett, D., Chuang, J. H., Springer, M., & Marth, G. T. Expression divergence measured by transcriptome sequencing of four yeast species. *BMC Genomics* 12, 635, (2011)[[JJL: delete]]

11. Boyle, A. P., Araya, C. L., Brdlik, C., Cayting, P., Cheng, C., Cheng, Y., Gardner, K., Hillier, L., Janette, J., Jiang, L., Kasper, D., Kawli, T., Kheradpour, P., Kundaje, A., Li, J. J., Ma, L., Niu, W., Rehm, E. J., Rozowsky, J., Slattery, M., Spokony, R., Terrell, R., Vafeados, D., Wang, D., Weisdepp, P., Wu, Y.-C., Xie, D., Yan, K.-K., Feingold, E. A., Good, P. J., Pazin, M. J., Huang, H., Bickel, P. J., Brenner, S. E., Reinke, V., Waterston, R. H., Gerstein, M., White, K. P., Kellis, M., Snyder, M., the modENCODE & ENCODE Consortia Comparative analysis of regulatory information and circuits across distant species. *Nature* submitted, (2013)

12. Ho, J. W. K., Liu, T., Jung, Y. L., Alver, B. H., Lee, S., Ikegami, K., Sohn, K.-A., Minoda, A., Tolstorukov, M. Y., Appert, A., Parker, S. C. J., Gu, T., Kundaje, A., Riddle, N. C., Bishop, E., Egelhofer, T. A., Hu, S. S., Alekseyenko, A. A., Rechtsteiner, A., Asker, D., Belsky, J. A., Bowman, S. K., Chen, Q. B., Chen, R. A.-J., Day, D. S., Dong, Y., Dosé, A. C., Duan, X., Epstein, C. B., Ercan, S., Feingold, E. A., Garrigues, J. M., Gehlenborg, N., Good, P. J., Haseley, P., He, D., Herrmann, M., Hoffman, M. M., Jeffers, T. E., Kharchenko, P. V., Kolasinska-Zwierz, P., Kotwaliwale, C. V., Kumar, N., Langley, S. A., Larschan, E. N., Latorre, I., Libbrecht, M. W., Lin, X., Park, R., Pazin, M. J., Pham, H. N., Plachetka, A., Qin, B., Schwartz, Y. B., Shoresh, N., Stempor, P., Vielle, A., Wang, C., Whittle, C. M., Xue, H., Kingston, R. E., Kim, J. H., Bernstein, B. E., Dernburg, A. F., Pirrotta, V., Kuroda, M. I., Noble, W. S., Tullius, T. D., Kellis, M., MacAlpine, D. M., Strome, S., Elgin, S. C. R., Ahringer, J., Liu, X. S., and Gary H. Karpen, and Jason D. Lieb, & Park, P. J. modENCODE and ENCODE resources for analysis of metazoan chromatin organization. *Nature* submitted, (2013)

13. Berget, S. M. Exon recognition in vertebrate splicing. *J Biol Chem* 270, 2411-4, (1995)[[JJL: delete]]

14. Talerico, M. & Berget, S. M. Intron definition in splicing of small Drosophila introns. *Mol Cell Biol* 14, 3434-45, (1994)[[JJL: delete]]

15. Stuart, J. M., Segal, E., Koller, D., & Kim, S. K. A gene-coexpression network for global discovery of conserved genetic modules. *Science* 302, 249-55, (2003)

16. Kirkpatrick, S., Gelatt, Jr, C. D., & Vecchi, M. P. Optimization by simulated annealing. *Science* 220, 671-80, (1983)

17. Reichardt, J. & Bornholdt, S. Detecting fuzzy community structures in complex networks with a Potts model. *Phys Rev Lett* 93, 218701, (2004)

18. Domazet-Lošo, T. & Tautz, D. A phylogenetically based transcriptome age index mirrors ontogenetic divergence patterns. *Nature* 468, 815-8, (2010) [[JJL: delete?]]

19. Djebali, S., Davis, C. A., Merkel, A., Dobin, A., Lassmann, T., Mortazavi, A., Tanzer, A., Lagarde, J., Lin, W., Schlesinger, F., Xue, C., Marinov, G. K., Khatun, J., Williams, B. A., Zaleski, C., Rozowsky, J., Röder, M., Kokocinski, F., Abdelhamid, R. F., Alioto, T., Antoshechkin, I., Baer, M. T., Bar, N. S., Batut, P., Bell, K., Bell, I., Chakrabortty, S., Chen, X., Chrast, J., Curado, J., Derrien, T., Drenkow, J., Dumais, E., Dumais, J., Duttagupta, R., Falconnet, E., Fastuca, M., Fejes-Toth, K., Ferreira, P., Foissac, S., Fullwood, M. J., Gao, H., Gonzalez, D., Gordon, A., Gunawardena, H., Howald, C., Jha, S., Johnson, R., Kapranov, P., King, B., Kingswood, C., Luo, O. J., Park, E., Persaud, K., Preall, J. B., Ribeca, P., Risk, B., Robyr, D., Sammeth, M., Schaffer, L., See, L.-H., Shahab, A., Skancke, J., Suzuki, A. M., Takahashi, H., Tilgner, H., Trout, D., Walters, N., Wang, H., Wrobel, J., Yu, Y., Ruan, X., Hayashizaki, Y., Harrow, J., Gerstein, M., Hubbard, T., Reymond, A., Antonarakis, S. E., Hannon, G., Giddings, M. C., Ruan, Y., Wold, B., Carninci, P., Guigó, R., & Gingeras, T. R. Landscape of transcription in human cells. *Nature* 489, 101-8, (2012)

20. Rozowsky, J. S., Newburger, D., Sayward, F., Wu, J., Jordan, G., Korbel, J. O., Nagalakshmi, U., Yang, J., Zheng, D., Guigó, R., Gingeras, T. R., Weissman, S., Miller, P., Snyder, M., & Gerstein, M. B. The DART classification of unannotated transcription within the ENCODE regions: associating transcription with known and novel loci. *Genome Res* 17, 732-45, (2007)

21. Bertone, P., Stolc, V., Royce, T. E., Rozowsky, J. S., Urban, A. E., Zhu, X., Rinn, J. L., Tongprasit, W., Samanta, M., Weissman, S., Gerstein, M., & Snyder, M. Global identification of human transcribed sequences with genome tiling arrays. *Science* 306, 2242-6, (2004)

22. Kapranov, P., Drenkow, J., Cheng, J., Long, J., Helt, G., Dike, S., & Gingeras, T. R. Examples of the complex architecture of the human transcriptome revealed by RACE and high-density tiling arrays. *Genome Res* 15, 987-97, (2005)

23. Lu, Z. J., Yip, K. Y., Wang, G., Shou, C., Hillier, L. W., Khurana, E., Agarwal, A., Auerbach, R., Rozowsky, J., Cheng, C., Kato, M., Miller, D. M., Slack, F., Snyder, M., Waterston, R. H., Reinke, V., & Gerstein, M. B. Prediction and characterization of noncoding RNAs in C. elegans by integrating conservation, secondary structure, and high-throughput sequencing and array data. *Genome Res* 21, 276-85, (2011)

24. Manak, J. R., Dike, S., Sementchenko, V., Kapranov, P., Biemar, F., Long, J., Cheng, J., Bell, I., Ghosh, S., Piccolboni, A., & Gingeras, T. R. Biological function of unannotated transcription during the early development of Drosophila melanogaster. *Nat Genet* 38, 1151-8, (2006)

25. Dobin, A., Davis, C. A., Schlesinger, F., Drenkow, J., Zaleski, C., Jha, S., Batut, P., Chaisson, M., & Gingeras, T. R. STAR: ultrafast universal RNA-seq aligner. *Bioinformatics* 29, 15-21, (2013)

26. modENCODE Consortium, Roy, S., Ernst, J., Kharchenko, P. V., Kheradpour, P., Negre, N., Eaton, M. L., Landolin, J. M., Bristow, C. A., Ma, L., Lin, M. F., Washietl, S., Arshinoff, B. I., Ay, F., Meyer, P. E., Robine, N., Washington, N. L., Di Stefano, L., Berezikov, E., Brown, C. D., Candeias, R., Carlson, J. W., Carr, A., Jungreis, I., Marbach, D., Sealfon, R., Tolstorukov, M. Y., Will, S., Alekseyenko, A. A., Artieri, C., Booth, B. W., Brooks, A. N., Dai, Q., Davis, C. A., Duff, M. O., Feng, X., Gorchakov, A. A., Gu, T., Henikoff, J. G., Kapranov, P., Li, R., MacAlpine, H. K., Malone, J., Minoda, A., Nordman, J., Okamura, K., Perry, M., Powell, S. K., Riddle, N. C., Sakai, A., Samsonova, A., Sandler, J. E., Schwartz, Y. B., Sher, N., Spokony, R., Sturgill, D., van Baren, M., Wan, K. H., Yang, L., Yu, C., Feingold, E., Good, P., Guyer, M., Lowdon, R., Ahmad, K., Andrews, J., Berger, B., Brenner, S. E., Brent, M. R., Cherbas, L., Elgin, S. C. R., Gingeras, T. R., Grossman, R., Hoskins, R. A., Kaufman, T. C., Kent, W., Kuroda, M. I., Orr-Weaver, T., Perrimon, N., Pirrotta, V., Posakony, J. W., Ren, B., Russell, S., Cherbas, P., Graveley, B. R., Lewis, S., Micklem, G., Oliver, B., Park, P. J., Celniker, S. E., Henikoff, S., Karpen, G. H., Lai, E. C., MacAlpine, D. M., Stein, L. D., White, K. P., & Kellis, M. Identification of functional elements and regulatory circuits by Drosophila modENCODE. *Science* 330, 1787-97, (2010)

27. Yip, K. Y., Cheng, C., Bhardwaj, N., Brown, J. B., Leng, J., Kundaje, A., Rozowsky, J., Birney, E., Bickel, P., Snyder, M., & Gerstein, M. Classification of human genomic regions based on experimentally determined binding sites of more than 100 transcription-related factors. *Genome Biol* 13, R48, (2012)

28. Kim, T.-K., Hemberg, M., Gray, J. M., Costa, A. M., Bear, D. M., Wu, J., Harmin, D. A., Laptewicz, M., Barbara-Haley, K., Kuersten, S., Markenscoff-Papadimitriou, E., Kuhl, D., Bito, H., Worley, P. F., Kreiman, G., & Greenberg, M. E. Widespread transcription at neuronal activity-regulated enhancers. *Nature* 465, 182-7, (2010)

29. Ren, B. Transcription: Enhancers make non-coding RNA. *Nature* 465, 173-4, (2010)

30. van Bakel, H., Nislow, C., Blencowe, B. J., & Hughes, T. R. Most "dark matter" transcripts are associated with known genes. *PLoS Biol* 8, e1000371, (2010)

31. Clark, M. B., Amaral, P. P., Schlesinger, F. J., Dinger, M. E., Taft, R. J., Rinn, J. L., Ponting, C. P., Stadler, P. F., Morris, K. V., Morillon, A., Rozowsky, J. S., Gerstein, M. B., Wahlestedt, C., Hayashizaki, Y., Carninci, P., Gingeras, T. R., & Mattick, J. S. The reality of pervasive transcription. *PLoS Biol* 9, e1000625; discussion e1001102, (2011)

32. Punta, M., Coggill, P. C., Eberhardt, R. Y., Mistry, J., Tate, J., Boursnell, C., Pang, N., Forslund, K., Ceric, G., Clements, J., Heger, A., Holm, L., Sonnhammer, E. L. L., Eddy, S. R., Bateman, A., & Finn, R. D. The Pfam protein families database. *Nucleic Acids Res* 40, D290-301, (2012)

33. Campo-Paysaa, F., Sémon, M., Cameron, R. A., Peterson, K. J., & Schubert, M. microRNA complements in deuterostomes: origin and evolution of microRNAs. *Evol Dev* 13, 15-27, (2011)

PMID=20133639 Karlic R, Chung HR, Lasserre J, Vlahovicek K, Vingron M: **Histone modification levels are predictive for gene expression.***Proc Natl Acad Sci USA* 2010, **107:**2926-2931. [PubMed Abstract](http://genomebiology.com/pubmed/20133639) | [Publisher Full Text](http://dx.doi.org/10.1073/pnas.0909344107) |

PMID=21177976 [**Integrative analysis of the Caenorhabditis elegans genome by the modENCODE project.**](http://papers.gersteinlab.org/papers/wormawg/index.html)

PMID=22955616‎, [**An integrated encyclopedia of DNA elements in the human genome.**](http://papers.gersteinlab.org/papers/encodemain/index.html)