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Mapping the Human miRNA Interactome by CLASH Reveals Frequent Noncanonical Binding

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SUMMARY

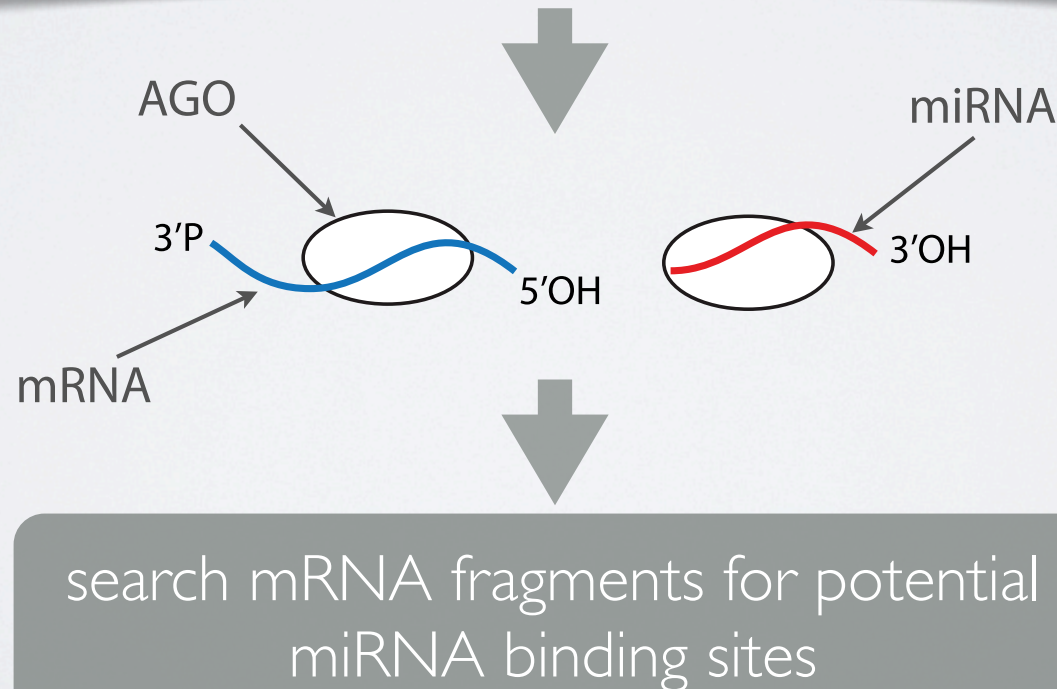
MicroRNAs (miRNAs) play key roles in gene regulation, but reliable bioinformatic or experimental identification of their targets remains difficult. To provide an unbiased view of human miRNA targets, we devel-

pairs with the target; (2) nucleotides paired outside the seed region stabilize interactions but are reported not to influence miRNA efficacy (Garcia et al., 2011; Grimson et al., 2007); and (3) functional miRNA targets are localized close to the extremes of the 3' UTRs of protein-coding genes in relatively unstructured regions (Grimson et al., 2007). Recently,

Argonaute HITS-CLIP decodes microRNA–mRNA interaction maps

Sung Wook Chi¹, Julie B. Zang¹, Aldo Mele¹ & Robert B. Darnell¹

MicroRNAs (miRNAs) have critical roles in the regulation of gene expression; however, as miRNA activity requires base pairing with only 6–8 nucleotides of messenger RNA, predicting target mRNAs is a major challenge. Recently, high-throughput sequencing of RNAs isolated by crosslinking immunoprecipitation (HITS-CLIP) has identified functional protein–RNA interaction sites. Here we use HITS-CLIP to covalently crosslink native argonaute (Ago, also called Eif2c) protein–RNA complexes in mouse brain. This produced two simultaneous data sets—Ago–miRNA and Ago–mRNA binding sites—that were combined with bioinformatic analysis to identify interaction sites between miRNA and target mRNA. We validated genome-wide interaction maps for miR-124, and generated additional maps for the 20 most abundant miRNAs present in P13 mouse brain. Ago HITS-CLIP provides a general platform for exploring the specificity and range of miRNA action *in vivo*, and identifies precise sequences for targeting clinically relevant miRNA–mRNA interactions.



HITS-CLIP

Mapping the Human miRNA Interactome by CLASH Reveals Frequent Noncanonical Binding

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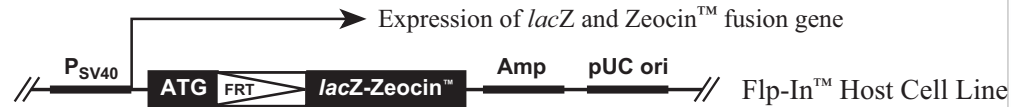
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SUMMARY

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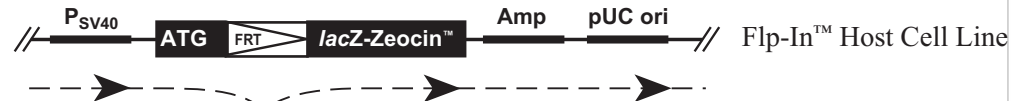
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1. pFRT/*lacZeo* is stably transfected into the mammalian cells of interest to generate the Zeocin[™]-resistant Flp-In[™] Host Cell Line(s)

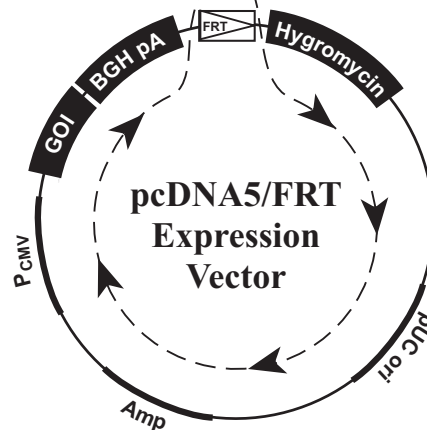


2. The pcDNA5/FRT expression vector containing your gene of interest (GOI) is cotransfected with pOG44 into the Flp-In[™] Host Cell Line.

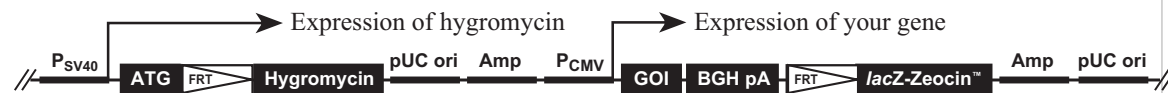
↓ + pOG44
+ pcDNA5/FRT



3. The Flp recombinase expressed from pOG44 catalyzes a homologous recombination event between the FRT sites in the host cells and the pcDNA5/FRT expression vector.



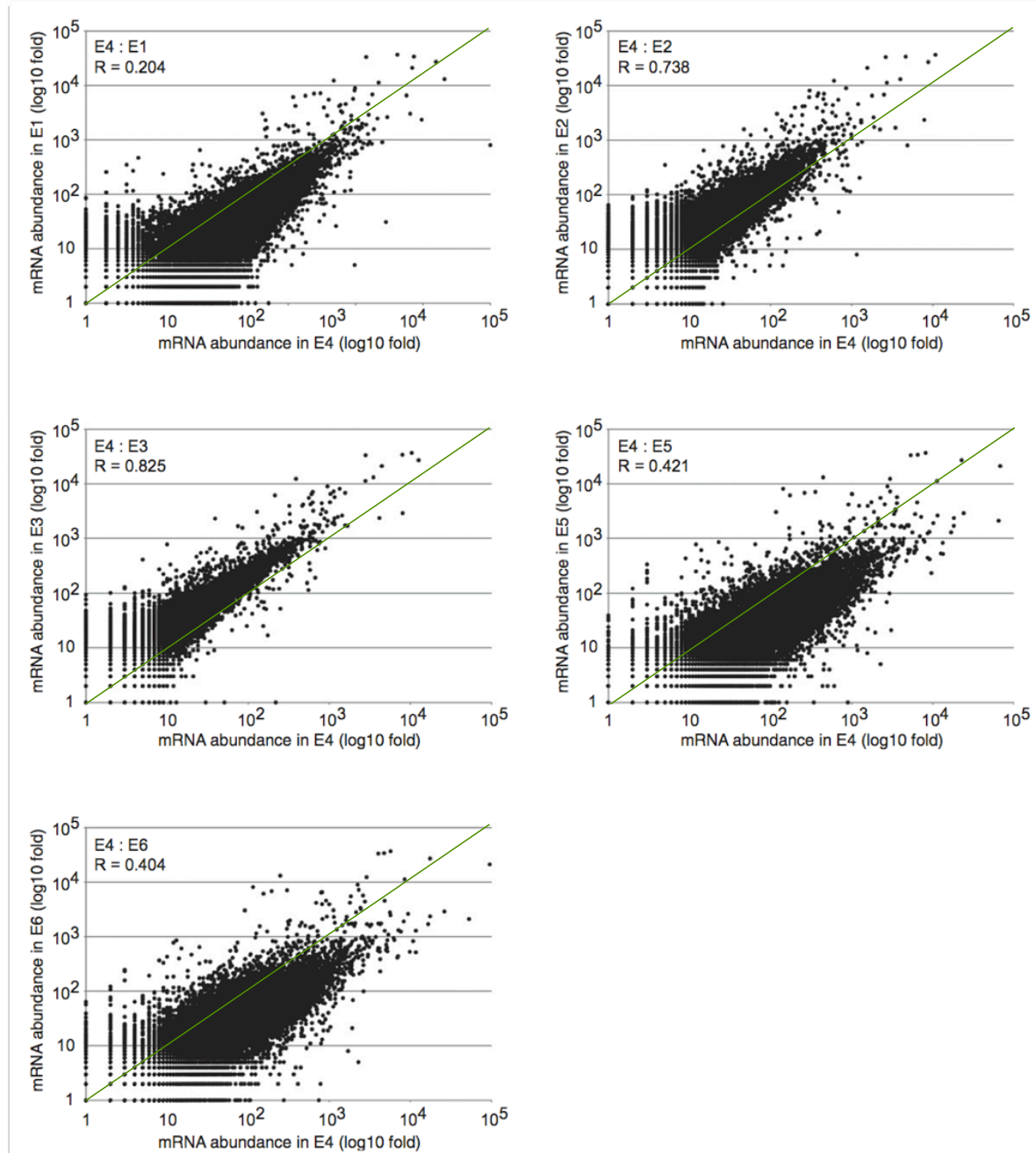
4. Integration of the expression construct allows transcription of the gene of interest (GOI) and confers hygromycin resistance and Zeocin[™] sensitivity to the cells.



enzymatic reactions at 37°C, not 20°C

different ligase for 3' linker

linker ligation performed in solution for 16 hours



3' linker ligation performed on beads, not gel & different ligase for 3' linker

linker ligation performed in solution for 1 hour

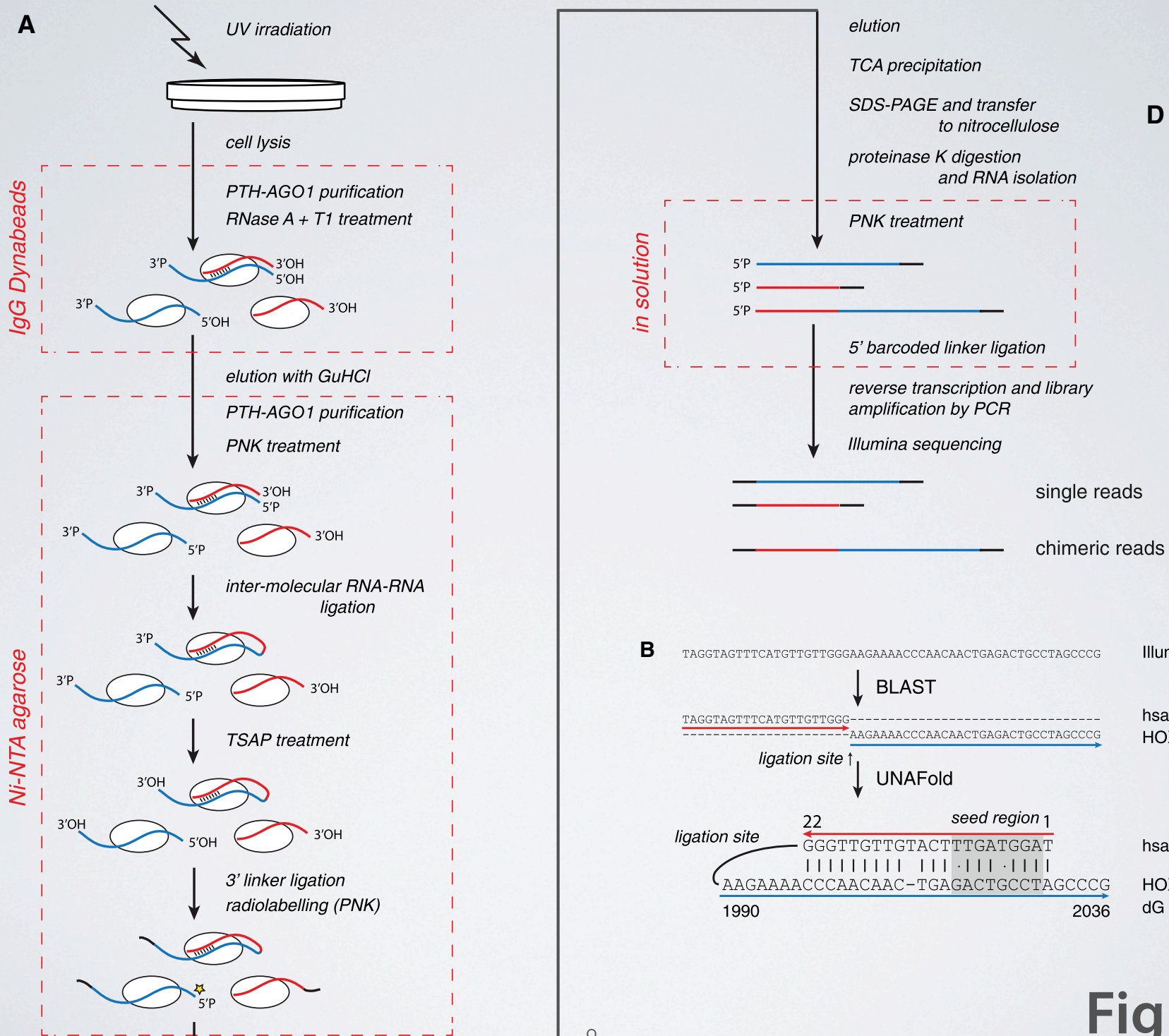
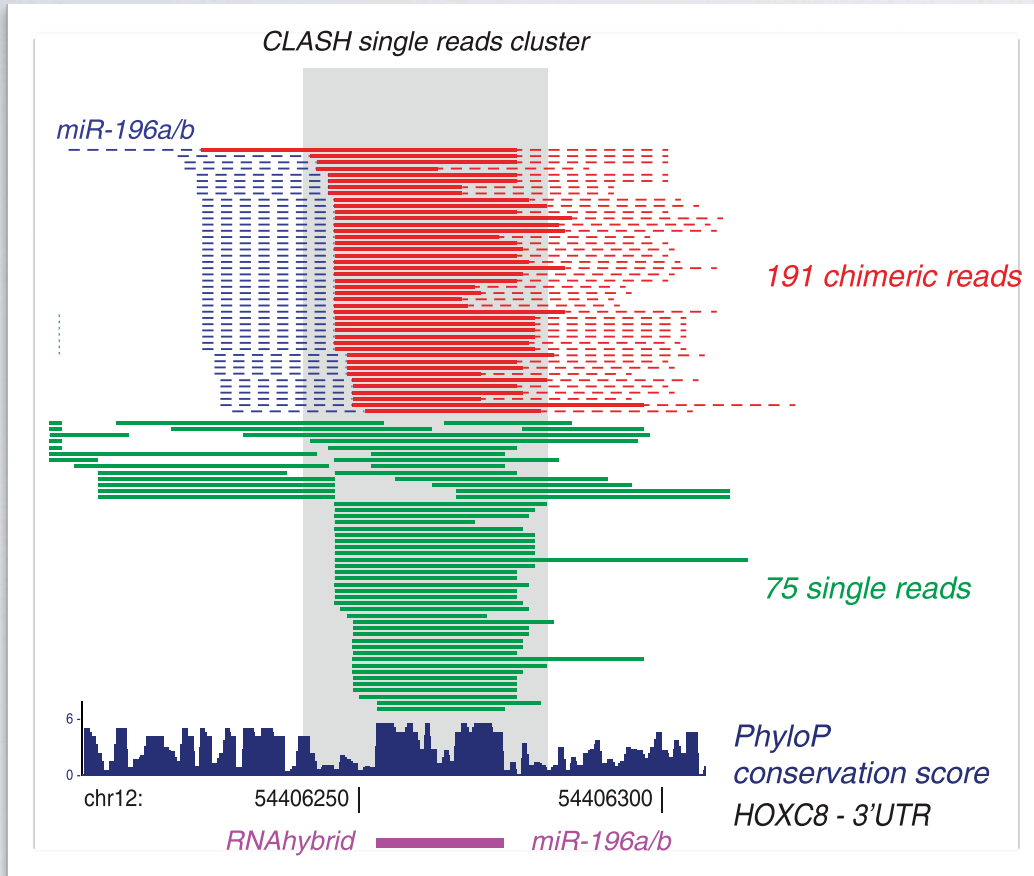
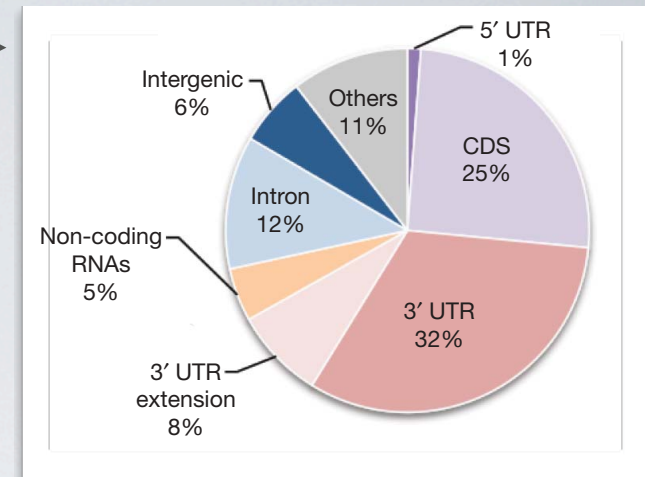


Figure 1

HITS-CLIP



interactions
(chimeric reads)

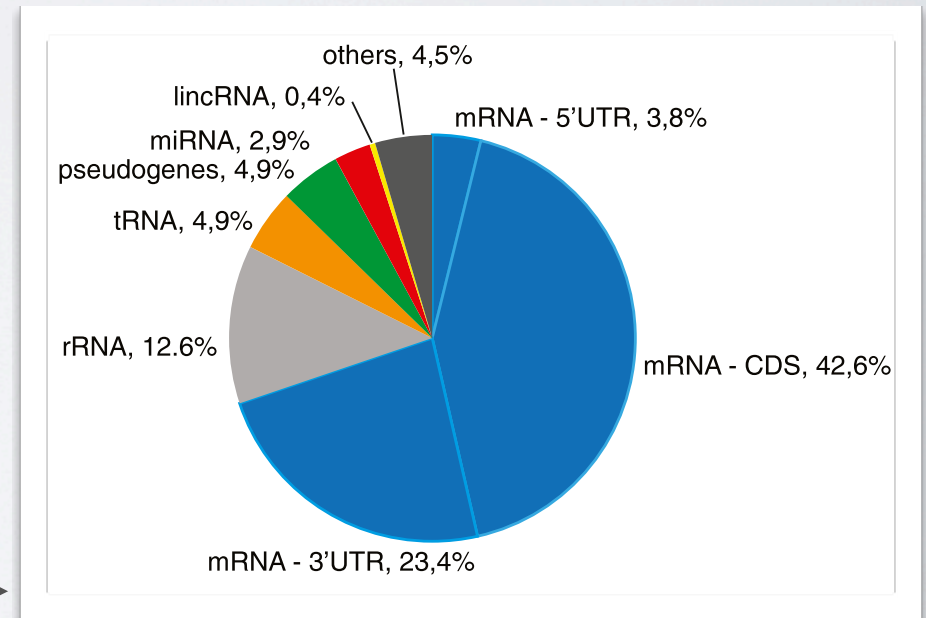


Figure 1

Table S2A

	ALL DATASETS		DATASET E4	
	Single reads	%	Single reads	%
microRNA*	21,832,771	20.6	1,722,779	7.6
mRNA	20,134,598	19.0	3,767,064	16.5
tRNA	40,910,266	38.5	11,596,776	50.9
rRNA	13,491,752	12.7	3,709,881	16.3
snRNA	1,546,938	1.5	300,695	1.3
pseudogenes	957,987	0.9	183,904	0.8
lincRNA	519,538	0.5	138,261	0.6
snoRNA	489,682	0.5	88,446	0.4
other	6,288,134	5.9	1,272,717	5.6
Total:	106,171,666		22,780,523	

Table S2B

	All	containing miRNAs	miRNA-mRNA (% of all)
ALL DATASETS			
CHIMERIC READS:	1,700,221	563,984	66,304 (3.9%)
collapsed reads	622,721	106,765	28,255 (4.5%)
number of supported interactions	240,135	26,543	18,514 (7.7%)
number of identified miRNAs			399
number of identified mRNAs [ENSG/ENST]			6,959 / 7,390
PUBLISHED DETAILED PROTOCOL (E4)			
CHIMERIC READS:	794,729	303,154	29,943 (3.8%)
collapsed reads	290,221	58,337	17,237 (5.9%)
number of supported interactions	110,148	16,386	12,198 (11.1%)
number of identified miRNAs			340
number of identified mRNAs [ENSG/ENST]			5,427 / 5,705

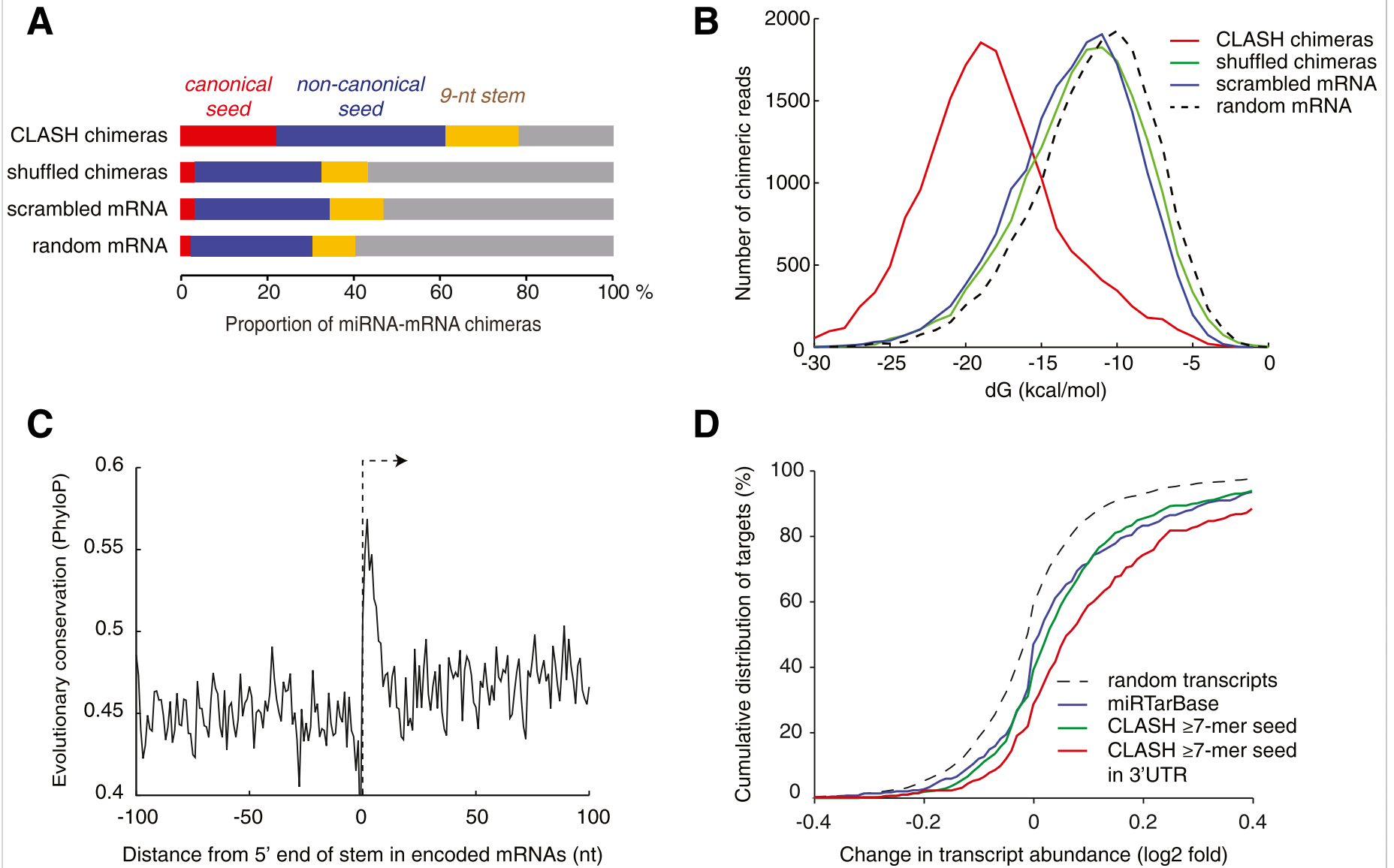


Figure 2

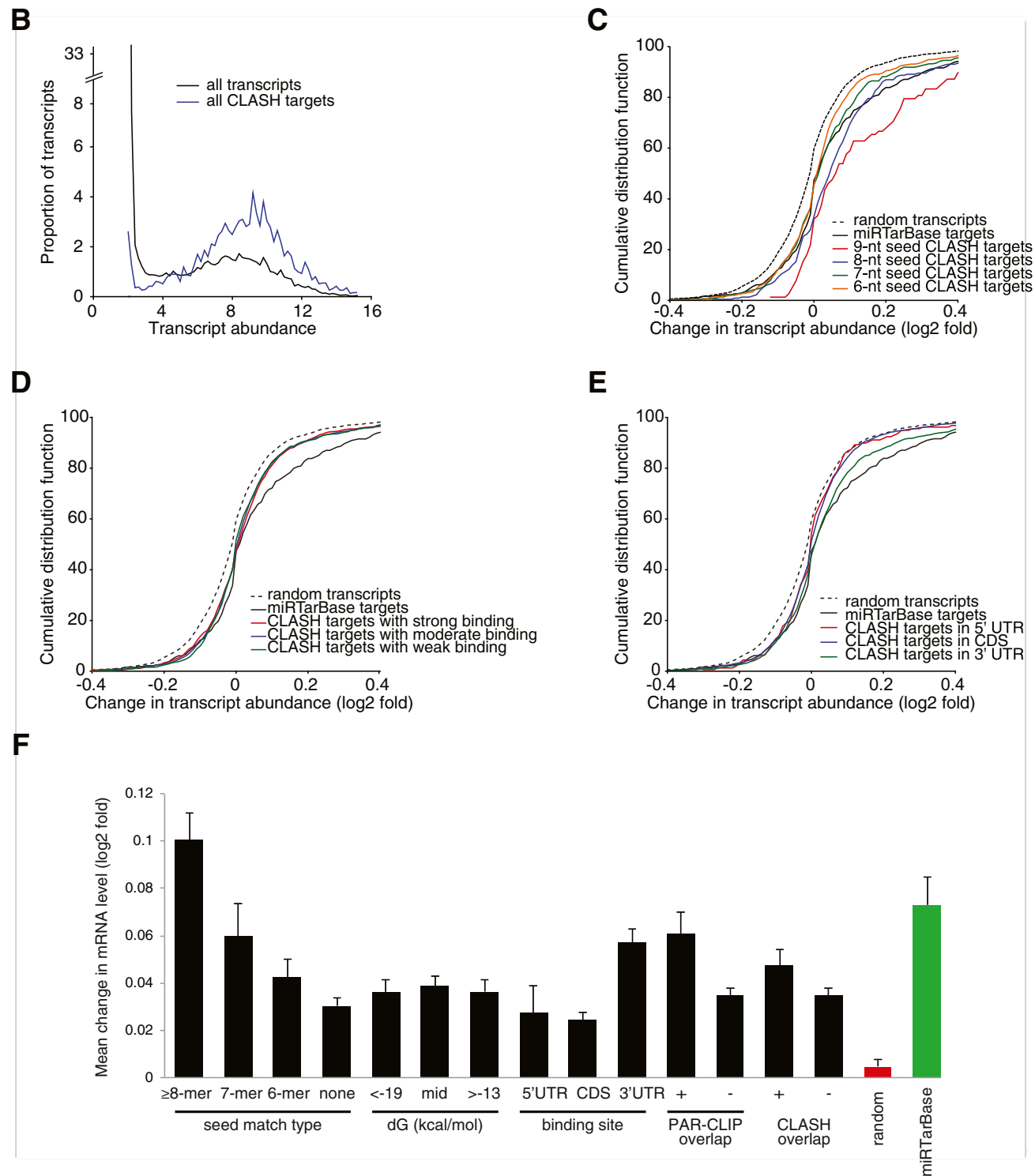


Table S4A

		PAR-CLIP clusters	CLASH single read clusters
	Number of interactions	15,823	28,515
CLASH chimeras	18,514	1,596 (9%)	3,066 (17%)
random localization within the same gene	18,514	1,125	2,036
random localization within a random gene with similar expression level	18,514	540	968
random localization within a random gene	18,514	268	439
CLASH single read clusters	28,515	2,528 (9%)	
random localization within the same gene	28,515	1,417	
random localization within a random gene with similar expression level	28,515	702	
random localization within a random gene	28,515	343	

Table S4B

		matches with CLASH	matches with control	enrichment
	Number of interactions	6,248	6,248	
miRanda	687,208	411	29	14.2 ×
PicTar	205,263	224	9	24.9 ×
PITA	192,255	195	2	97.5 ×
RNAhybrid	992,584	310	25	12.4 ×
TargetScan	54,199	170	5	34.0 ×
all predictions	2,131,509	802	59	13.6 ×

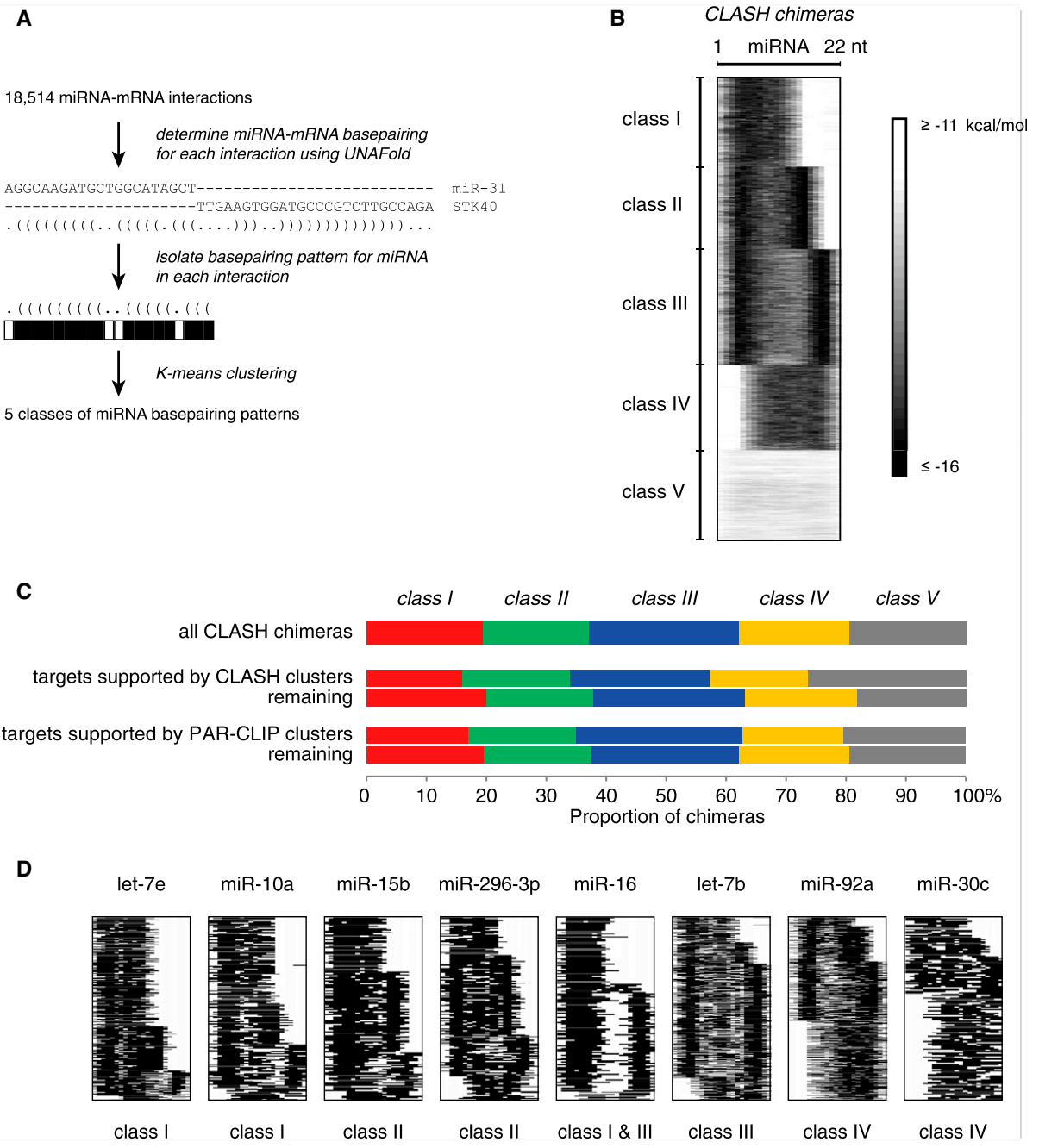


Figure 3

Table 1. Analysis of the Five miRNA-mRNA Base-Pairing Classes

Class	I	II	III	IV	V
Number of interactions	3,594	3,293	4,630	3,389	3,608
Number of base-paired nucleotides	13.0 ± 0.04	15.3 ± 0.03	16.8 ± 0.03	14.6 ± 0.04	11.9 ± 0.05
Number of base-paired nucleotides in seed	5.2 ± 0.02	5.1 ± 0.02	5.0 ± 0.01	2.6 ± 0.02	3.3 ± 0.03
Interaction energy (dG)	-18.3 ± 0.04	-20.2 ± 0.06	-20.5 ± 0.05	-19.0 ± 0.05	-11.1 ± 0.05
PhyloP conservation score	0.092 ± 0.017	0.127 ± 0.018	0.097 ± 0.017	0.011 ± 0.017	0.086 ± 0.018
Efficiency of inhibition by miRNA	0.042 ± 0.007	0.052 ± 0.009	0.047 ± 0.005	0.024 ± 0.005	0.039 ± 0.004
Targets in 5' UTR	4.8%	4.2%	4.1%	5.8%	4.7%
Targets in CDS	60.7%	61.1%	61.4%	63.9%	53.4%
Targets in 3' UTR	32.7%	32.2%	32.1%	28.1%	39.5%

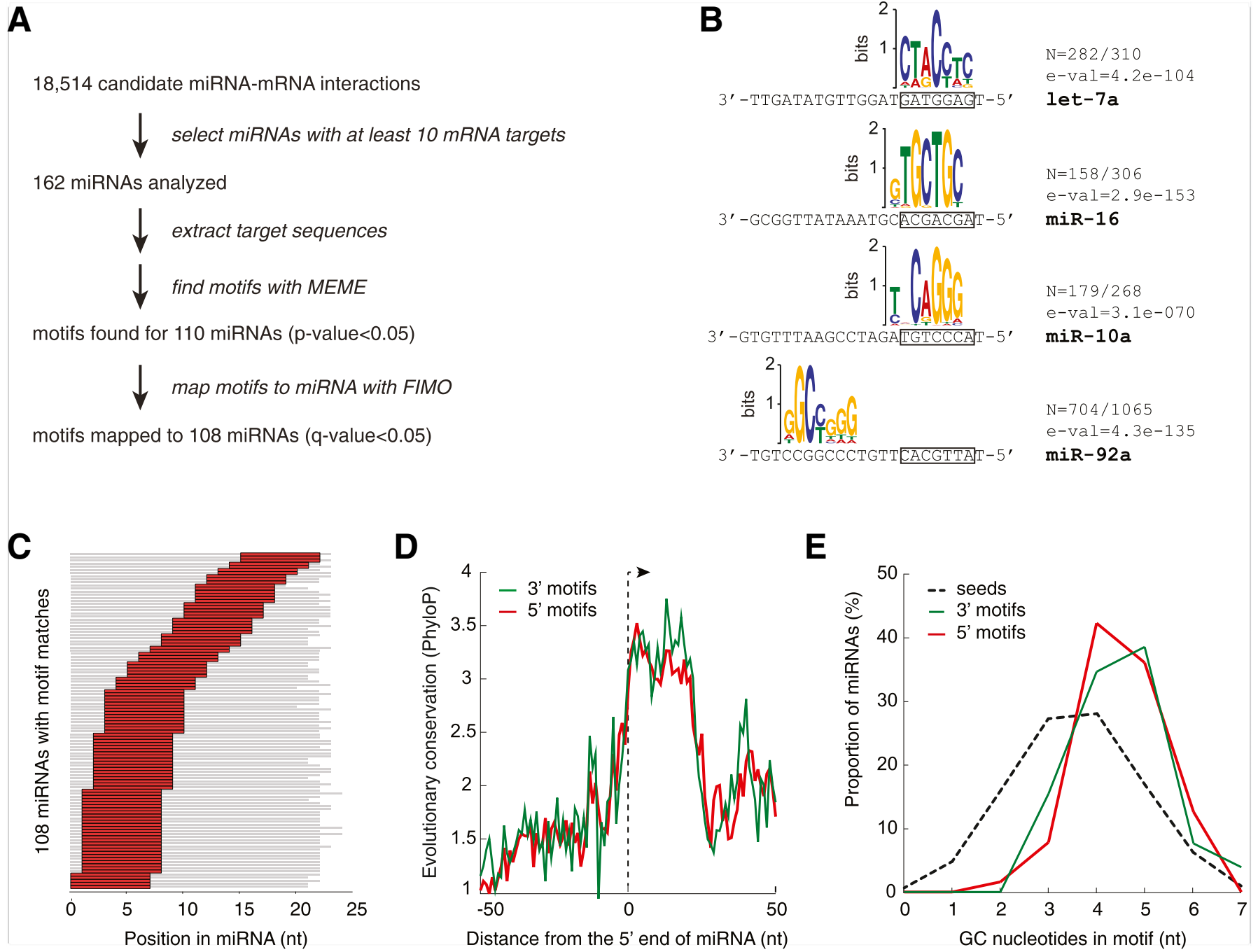
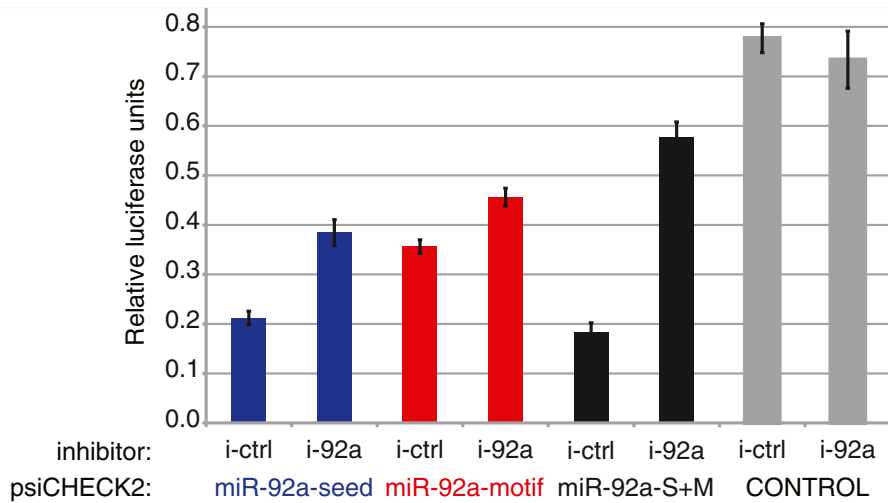


Figure 4

A



B

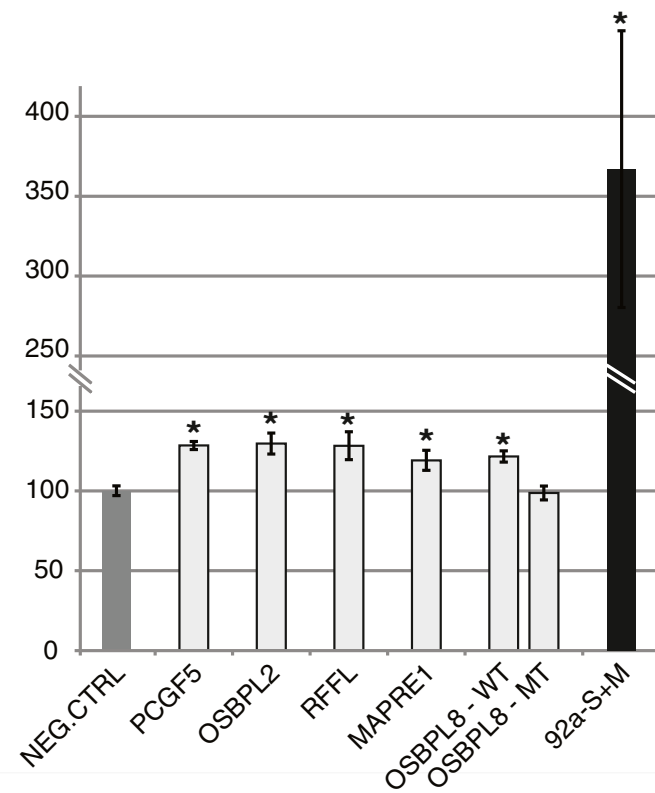


Figure 5

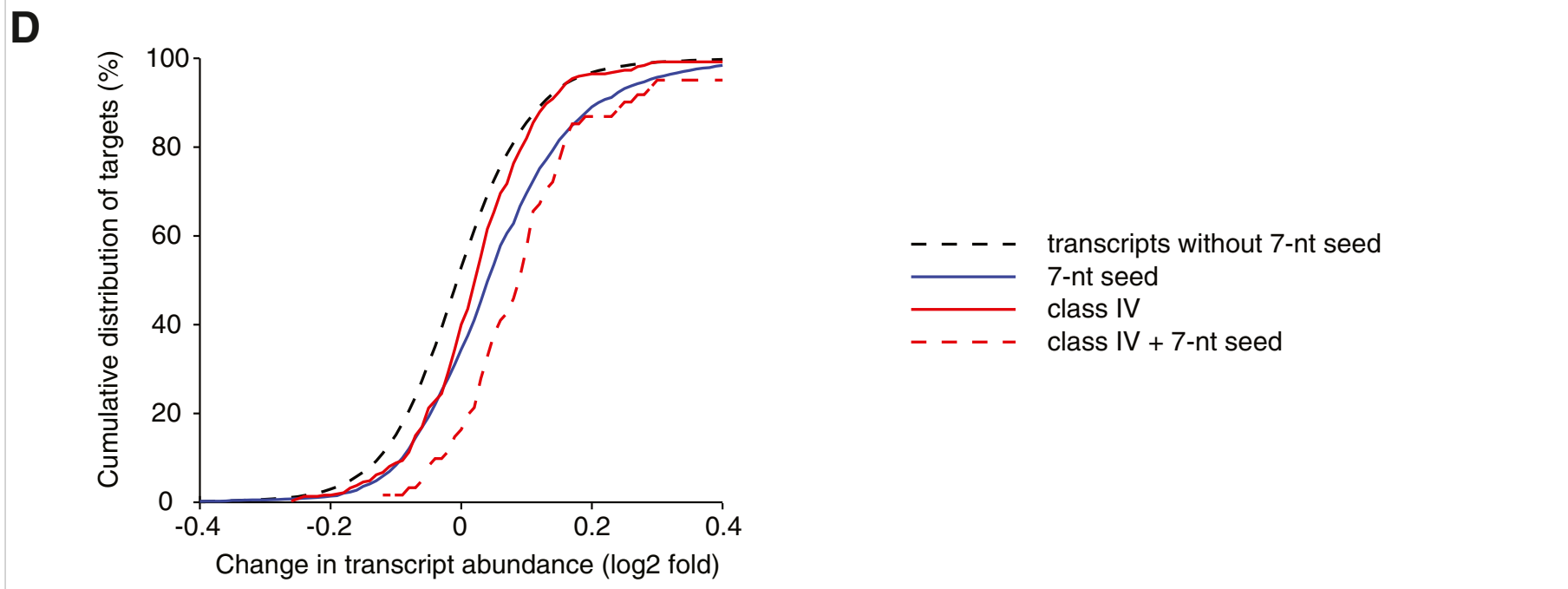
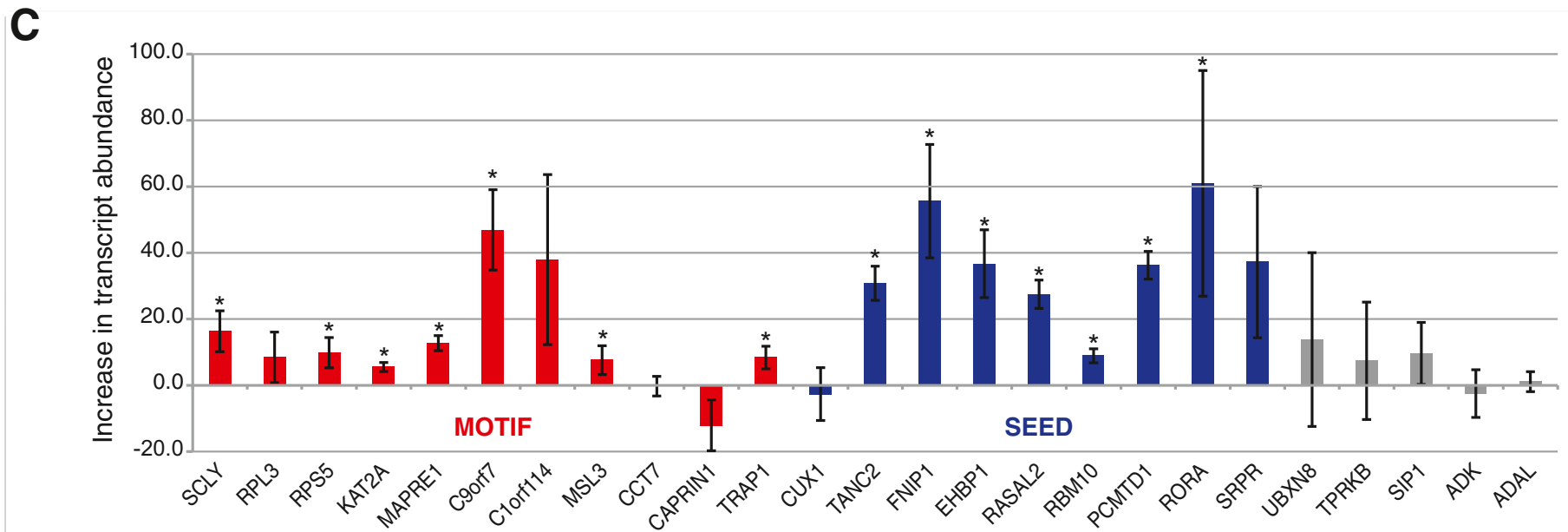
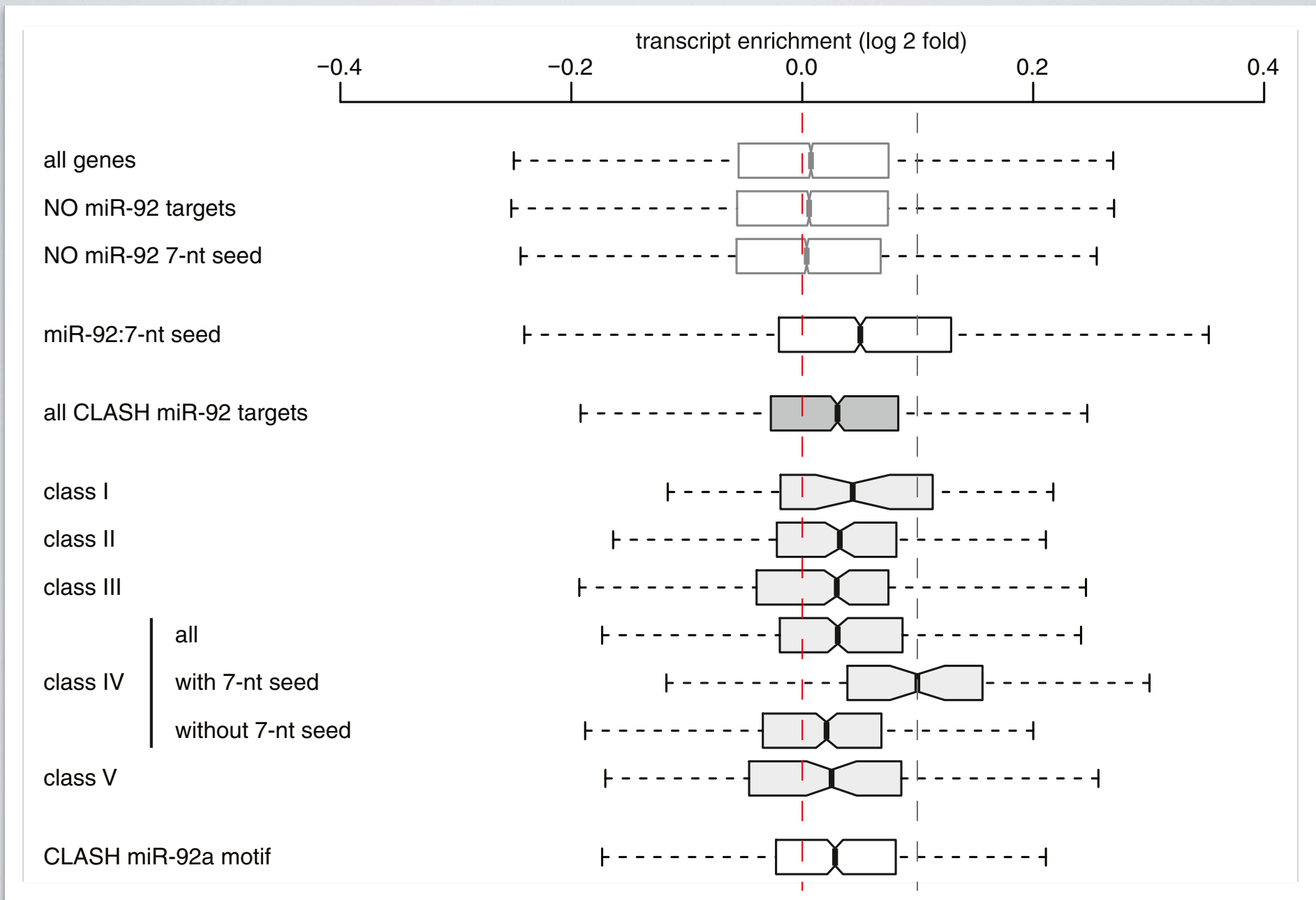


Figure 5



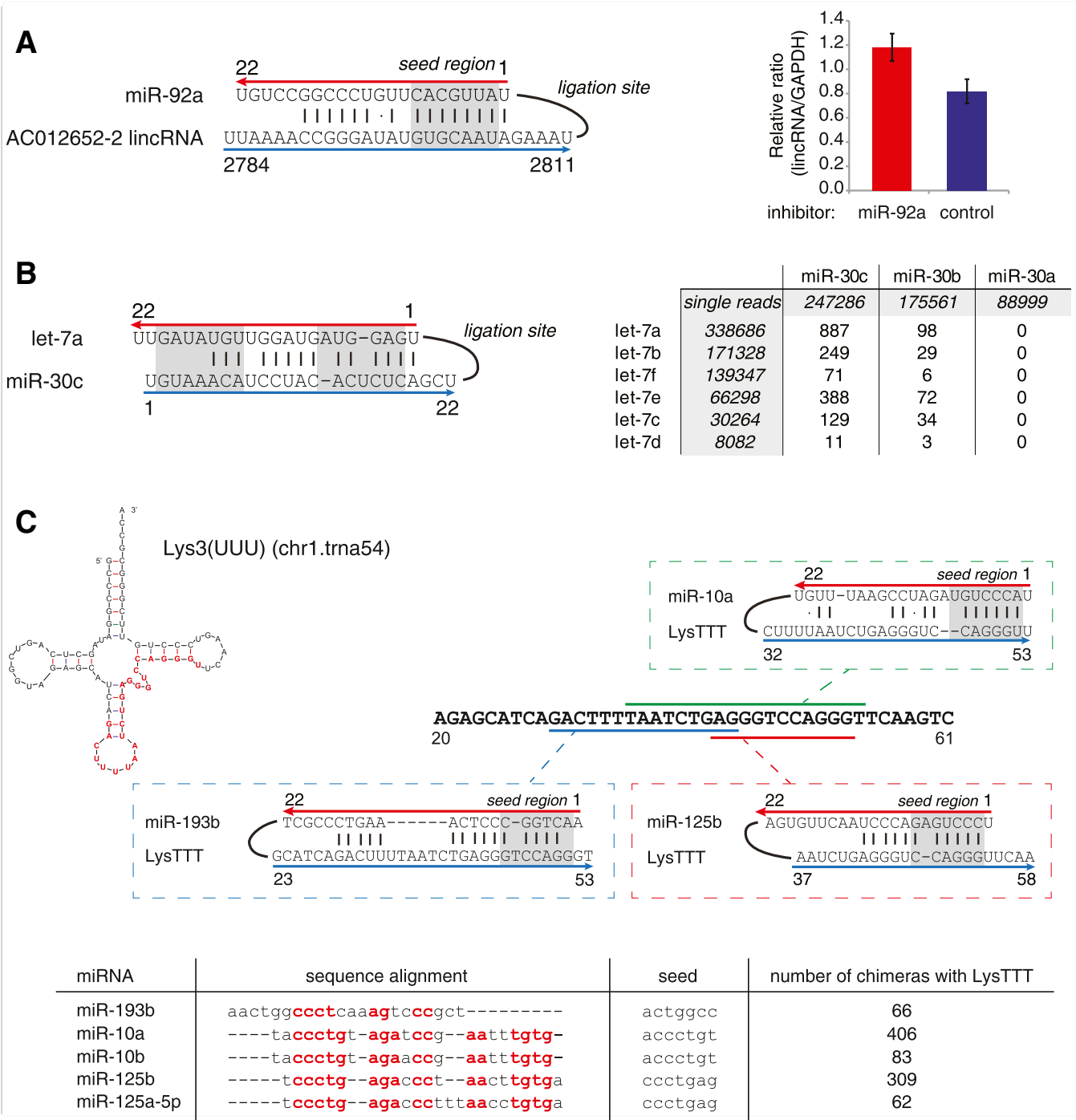


Figure 6

cliffs notes

- report **18,500** miRNA-mRNA interactions
- only around **37%** of seed interactions involve **uninterrupted base pairing**
- **16%** of all interactions **do not involve the seed region**
- “competing endogenous RNA” from pseudogenes/lincRNAs **AND abundant stable RNAs** such as tRNA/rRNA to buffer temporary excess of particular miRNAs