Report on Transcriptome Analysis

modENCODE Joint AWG call 13 July 2012

Datasets

- agreed-upon "expression compendium"
 - total RNA
 - ENCODE Tier 1
- developmental time courses (worm, fly)
- matched embryonic datasets

Transcription Paper Outline

- Comparison of protein-coding genes
 - Comparison with existing annotations (Hillier, Davis, Brown)
 - Splicing complexity (Graveley)
 - Comparison of select orthologs (Mortazavi, Harrow, Celniker)
- Comparison on non-coding RNAs (Brown, Lai, Gerstein, Guigo, Samsonova)
- Comparison of pseudogenes (Gerstein)
- Analysis of relationship of upstream regions to transcript level (Gerstein, Weng)
- Expression clustering (Brenner, Gerstein)

Comparison with existing annotations

- Because of the difficulty of assembling full transcripts with short reads and comparing their expression across species, we will focus on comparing transcript elements:
 - Transcript Start Sites (TSSs)
 - Transcript End Sites (TESs)
 - Splice Junctions (SJ)
 - de novo exons
 - de novo genes
 - de novo transcripts
 - Expression values for each above element
 - Expression values for the annotations

Number of protein-coding genes



Finding all isoforms of a gene can be difficult





C. elegans refseq models and spliced ESTs



Analysis of Splicing Complexity



Brenton Graveley

Analysis of Splicing Complexity

• For all three species, compare motifs and conservation at splice sites for constitutive vs. alternative exons, and highly switching vs. low switching.

• Analyze number of isoforms per gene.

Highlight outliers (Dscam, etc.)



Number of isoforms

Brenton Graveley

Comparison of select orthologs

Case Study: DUT / Dut / dUTPase / dut-1

Human DUT CCD 545256.1 -CCD 945255.1 --11-11 CCD \$32231.1 -11-1 CCDS set -11 protein coding < 800-TUG -1111 protein coding DUT-005 > -n-n T-001 -11-1 DUT-201 > protein coding -m-10 OUT-009 > 1111 protein coding DUT-002 > **0***0 nonsense mediated decay -m-m____ DUIT COS DUT-004 > protein coding protein coding DUT-010 > -11-1 protein coding DUT-012 > processed transcript cm-4 DUT-011 > retained intron 0 < Y_RNA 421-201 misc RNA < RP11-154J22.1-002 antisense CHAP11-154/22.1-001

Mouse Dut



Fly dUTPase



Worm dut-1



Adam Frankish

Comparison of non-coding RNAs

- How much of the nc genome is transcribed?
 - per megabase
 - across entire agreed-upon "expression compendium"
 - in ~matched embryonic stages
 - Ubiquitous vs Stage- / Cell-line specific transcription
- You cannot directly compare annotations (Gencode vs Flybase vs Wormbase)
- so, use a tiered approach; build a table or pie chart
 - first compare the existing annotations
 - incRNA algorithm
 - breakdown by RNA class
 - de novo mapping / TAR calling
 - issues: repeats, multi-mapped vs unique reads

Comparison of existing annotations

Number of short ncRNA



rRNA, tRNA, miRNA, snRNA, snoRNA (! mouse excludes tRNA)

Adam Frankish

incRNA algorithm



Results for known types of ncRNAs:



Comparison of pseudogenes

• Pseudogenes annotated using automated pipelines intersected with manual curation



	Human – GENCODE	Worm	Fly
Total	11240 (14112*)	1198	529
Duplicated	2158	538	119
Processed	8715	255	95
Ambiguous	23	405	315
Others**	344		

* Estimated total number of pseudogenes in human genome.

** Including Unitary (138), IG (161) TR V (21) and polymorphic (24) pseudogenes

*Transcribed Pseudogenes

Human

Worm



*Transcription Factor Binding Sites



- TFBS were selected within 2kb upstream of the pseudogene start site
- 95 (58) duplicated and 29 (20) processed pseudogenes had TFBS in the upstream region

Analysis of relationship of upstream regions to transcript level

His. mods around TSS & TTS are clearly related to level of gene expression, in a position-dependent fashion



Correlation between Signal and expression

Application of HM model in 3 species: Consistent Performance

>50% of variation of expression levels can be explained by HMs







Chao Cheng

Expression clustering of protein-coding and ncRNA genes in embryo development

Species	Developm ental stages	Protein- coding genes*	Non- coding RNAs*	Co- expression modules**
Worm (C. elegans)	111	9114	855	69
Fly (D. mel.)	50	8340	357	46
 * >80% valid samples, coeff. of variance > 1 in the modENCODE finalized datasets in June 2012 ** clustering via weighted gene co-expression network analysis (WGCNA) 				

Many co-expression modules are enriched with ncRNAs (red circles).





Daifeng Wang

Influence of ncRNA hubs on protein-coding co-expression modules

Influential ncRNAs (high network centrality) exist in modules NOT enriched with ncRNAs (blue circles).



Developmental stage mapping between worm and fly based on co-expression clustering of orthologs

- Gene expression threshold: FPKM >=1 and z >= 1.5
- Significance calculated from fraction of orthologs co-expressed between pairs of stages compared to hypergeometric expectation
- Cluster numbering facilitates follow-on analysis:



Developmental stage mapping between worm and fly based on co-expression clustering of orthologs



Jingyi Jessica Li, Peter Bickel, Haiyan Huang, Steven Brenner

END

Production Stats - Worm

	Samples	Total Reads	Total Unique Reads
Embryonic Time Course	106	1,633,419,670	1,031,557,649
Life Stages	70	2,401,311,389	1,420,342,487
Other Species	54	1,779,775,463	946,431,824
Pathogens	11	702,645,329	489,536,643
Tissues	183	3,560,398,393	1,322,552,917
Totals		10,077,550,244	5,210,421,520

Production Stats - Fly

Experiment	Samples	Total Reads	Total Unique Reads	Total Unique bp
Cell Lines	25	1,677,980,920	1,272,452,612	96,706,398,512
Tissues	29	4,265,585,752	3,667,365,400	278,719,770,400
Treatment	21	6,495,812,560	4,949,215,447	376,140,373,972
Poly(A) Tail Enrichment	29	845,610,153	638,882,610	48,555,078,360
Developmental Time Course*	30	3,538,880,404	2,282,408,273	171,180,620,475
Genome Resequencing	25	943,927,826	N/A	71,738,514,776
Total	247	17,767,797,615	12,810,324,342	1,043,040,756,495



Comparison of Fly Stages

Jingyi Jessica Li, Peter Bickel, Haiyan Huang, Steven Brenner



Comparison of Worm Stages